



In Silico Computational Analysis of *Citrus aurantium* of *Thurunji Manapagu* in Inhibiting the Receptor Target of Angiotensin-converting Enzyme Against *Uratha Pitha Vatham* (Hypertension)

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Abstract

Citrus aurantium (*Thurunji*) is a tree cultivated in various parts of the world. The fruit pulp is used to make juices, marmalade and pickles. It is commonly used by tribes to treat digestive disorders, constipation, heaviness in the chest, prolapse of the uterus, anorexia, chest pain, cold, and cough. In the Siddha system of medicine, *Citrus aurantium* fruit pulp is used to prepare a formulation, syrup in consistency named *Thurunji manapaagu* indicated for diseases due to deranged *pitha* humor. According to the Siddha system of medicine Hypertension termed *Uratha Pitha Vatham* is an ailment that occurs due to an elevated *pitham*. Considering the facts an attempt of *in silico* docking study was done with the selected phytochemicals such as Linalool, Apigenin, Ichangin, Kaempferol, Luteolin, Limonin, Limonene, Myrcene, Diosmetin and Tangeretin of *Citrus aurantium* against Angiotensin Converting Enzyme (ACE) and the results are detailed in this article.

Keywords: Antihypertensive, *Citrus aurantium*, Molecular Docking, Siddha Medicine, *Uratha Pitha Vatham*

1. Introduction

Citrus aurantium (Bitter orange), also called Seville orange or sour orange, is an evergreen tree cultivated all over the world. It bears spherical fruits with a thick, orange or sometimes green peel. The juicy pulp tastes sweet or sour, is orange in colour used in the treatment of gastrointestinal problems, abdominal distension heaviness in the chest, uterine prolapse, treatment of anorexia, chest pains, colds, coughs, etc. The fruit and other parts of the plant are being used in traditional systems of medicines for the cure of different ailments. The phytochemical composition of *C. aurantium* includes vitamins, minerals, phenolic

compounds, and terpenoids play a major role in its health-promoting effects¹⁻⁴. The fruit pulp is rich in phytochemicals (Limonoid glucosides) such as limonin, nomilinin, nomilin, obacunone, ichangin, and flavanols kaempferol, quercetin, free flavones such as apigenin, luteolin, tangeretin and diosmetin⁵⁻⁸.

The pharmacological profile of the fruit indicates limonene, protective against lesions induced by ethanol and nonsteroidal anti-inflammatory drugs in rats by increasing the production of gastric mucus⁹.

In vitro assays had proved the antimicrobial activity of *C. aurantium* juice against *Salmonella* species and *Listeria monocytogenes*. The antioxidant, cytotoxic, anxiolytic, and antidiabetic effects of *C.*

aurantium juice were established by various studies. It has been also used for weight loss and to enhance sports performance¹⁰⁻¹⁶. A drink called Zhi-Zi-Hou-Po (ZZHPD) prepared with *C. aurantium*, *Gardenia jasminoides* and *Magnolia officinalis* has been used to treat depression-like symptoms¹⁷.

1.1 Uratha Pitha Vaatham

The symptoms of Hypertension are more similar to those of *Uratha Pitha Vatham* elaborated in Siddha literature *Yugi vaithiya Cinthamani* – 800. *Uratha pitha vaatham* is one among the 42 types of *azhal noi* that arise due to the increased *azhal kutram* caused by the heat generated in the abdomen and moved to the neck region, which affects the healthiness of blood. The *azhal kutram* is affected due to increased intake of sour, spicy and salty foods, frequent anger, mental stress, insomnia, eating of undercooked food and exposure to heat like flame and sunlight.

According to Siddha's basic principles, human beings are incorporated with 96 principles (*thathuvangal*), which include physical, functional, psychological and intellectual components and the diagnosis is based on Seven physical constituents (*Ezhu udal thathukal*), Eight investigatory tools (*Envagai thervu*), Three vital forces (*Mukkuttram*). In *Uratha pitha vaatham* - the *azhal kuttram* (*Anar pitham*, *Saadhaga pitha*, *Prasaga pitham*, *Aalosaga pitham*) gets affected which continuously tends to affect the *vali kuttram* (*Uyirkaal*, *Paravukaal*, *Nadukaal*) and as a result will increase the blood pressure¹⁸.

1.2 Hypertension

Hypertension is the increase in Blood pressure, known to end in cardiovascular and renal complications¹⁹. It was estimated that 1.28 billion adults aged 30-79 have hypertension globally. Considering this fact, the World Health Organization (WHO) released a new guideline regarding the pharmacological management of hypertension in adults during 2021²⁰.

Thurinji manapaagu is a Siddha formulation cited in the traditional literature book *Siddha vaithiya thirattu* used to treat the *Pitha thodam* and diseases preceding the *Pitha thodam*²¹. *Thurunji* (*Citrus aurantium*) belongs to the family Rutaceae. Besides vitamin C, flavonoids and volatile oil, the other phytoconstituents of citrus fruits are phenethylamine alkaloids like octopamine, synephrine, tyramine, etc. Fruit extract of *Citrus aurantium* has astringent, antioxidant, carminative

and spasmolytic activity. It is used in gastrointestinal disorders, sleep disturbances, cardiovascular diseases and cancer treatments²².

The Renin Angiotensin System (RAS) cascade is responsible for the regulation of blood pressure²³. Therapeutic agents that can block RAS initiation through inhibition of ACE, angiotensin receptor inhibitors and renin inhibitors are commonly used in Hypertension management²⁴.

Molecular docking or ligand-protein docking is usually performed between a small molecule and a target macromolecule. It has a variety of applications in drug discovery, including structural activity study, lead optimization, discovery of potential leads to facilitate predictions in mutagenesis studies, assisting X-ray crystallography, and designing chemical mechanisms²⁵. Considering the facts mentioned, selected phytochemicals of *Thurunji* (*Citrus aurantium*) were subjected to molecular docking and the outcome is discussed in this article.

2 Materials and Methods

2.1 Preparation of Thurunji Manapaagu

The investigational drug *Thurunji Manapaagu* is an herbal formulation prepared using fresh fruit juice of *Citrus aurantium* and sugar obtained from *Saccharum officianlae*. The juice is extracted from *Citrus aurantium* fruit, filtered and sugar added in the ratio 1:2 boiled to *Manapaagu* (syrup) consistency²¹.

2.2 In silico Method - Docking Study

Molecular docking was done computationally to identify the architecture of compounds generated by two or more distinct molecules. It plays a critical role in rational drug design by anticipating the alignment of small molecules of therapeutic compounds, their affinity and activity²⁶. Preferring with minimal side effects, use of phytochemicals has emerged as one of the holistic alternative approaches²⁷.

Screening molecules with potential bioactivity is quite costly and may consume time, which could be overcome by Computer-Aided Drug Design (CADD). In recent years, chemoinformatics, molecular docking, and artificial intelligence have considerably increased in designing a drug, its development and discovery^{28,29}. *In silico* molecular docking virtually predicts the binding efficacy and the structure-based drug design³⁰. It also

provides details on structure-activity relationships, mode of action, and analysis of interaction between protein and ligand³⁰. Therefore, *in silico* analysis would result in providing better chances of discovering suitable drug candidates in considerably short duration and less expense. Nowadays, several molecular docking approaches based on molecular structure and ligands are available to facilitate drug discovery^{28,30}.

2.2.1 Retrieval of Drug Compound-Captopril

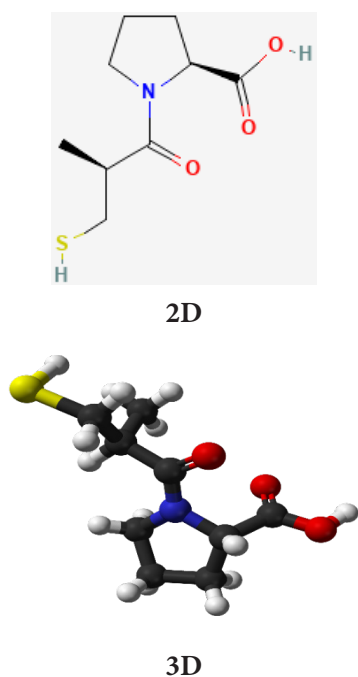


Figure 1. 2D and 3D structure models of drug compound - Captopril.

Captopril (C₉H₁₅NO₃S) (structure given in Figures 1 and 2) is an analogue of proline that possesses antihypertensive and antineoplastic activity by competitively inhibiting ACE and reduces the level of Angiotensin II by promoting plasma renin activity and suppressing aldosterone secretion.

2.2.2 Retrieval of Phytochemical Data

Phytochemical data of components such as Linalool, Limonene, Myrcene, Apigenin, Ichangin, Kaempferols, Luteolin, Diosmetin Limonin, Tangeretin are retrieved from published articles³¹ using search engines, IMPPAT (Indian Medicinal Plants, Phytochemistry and Therapeutics). IMPPAT is a database that contains the 2D and 3D structures of phytochemical constituents present in Indian medicinal plants.

Captopril

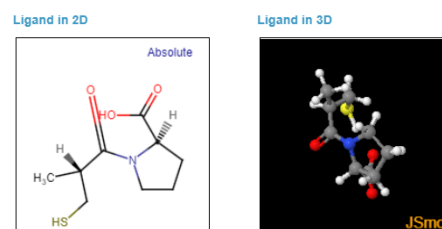


Figure 2. 2D, 3D pictures of standard Captopril.

2.2.3 Ligand Preparation

A literature survey conducted using the search engines PubMed and Google Scholar to retrieve the phytochemicals present in the drug to be subjected to docking, and ten lead phytocomponents were identified and retrieved (Table 1). The 3D structure of the ligands was built using ChemDraw prof online tool version 12.0. The ligands were prepared with Merck Molecular Force Field 94 (MMFF94). The ligand properties of the selected compounds, molecular weight, molecular formula, H bond donor, receptor, rotatable bonds for docking against ACE (1O86), and of the standard drug captopril are shown in Table 1.

Table 2 represents 2D and 3D pictures of selected ligands of *Cirtus aurantium*.

2.2.4 Protein Receptor Preparation

Three dimensional protein structure of target protein ACE(PDB)1086 given in Figure 3 was retrieved from the online repository of Protein Data Bank (www.rcsb.org/pdb) and subjected to a protein cleanup process before docking simulation. Essential missing hydrogen atoms Kollman charges were added using the MGL AutoDock Tools 1.5.7, and the statistical distribution of the combinations of the amino acid backbone dihedral Ø (Phi) angles and ψ (Psi) angle were identified by subjecting to Ramachandran plot analysis using Rampage. Before the docking simulation, the water molecules were removed, and the protein clean geometry procedure was done. Orientation of the lead molecules concerning the target protein and their best dock pose was selected based on the interaction study analysis using the Auto dock version 4 program³².

2.2.5 Molecular Docking

To define the rotatable bonds, Gasteiger partial charges were added, and non-polar hydrogen atoms were merged

Table 1. Ligand properties of selected phytochemicals for docking against ACE (1O86) and captopril

S. No.	Compound	Molar weight g/mol	Molecular Formula	H Bond Donor	H Bond Acceptor	Rotatable bonds
1.	Linalool	154.25 g/mol	C ₁₀ H ₁₈ O	1	1	4
2.	Limonene	136.23 g/mol	C ₁₀ H ₁₆	0	0	1
3.	Myrcene	136.238 g/mol	C ₁₀ H ₁₆	0	0	4
4.	Ichangin	488.5 g/mol	C ₂₆ H ₃₂ O ₉	2	9	2
5.	Kaempferol	286.239 g/mol	C ₁₅ H ₁₀ O ₆	4	6	1
6.	Luteolin	286.24g/mol	C ₁₅ H ₁₀ O ₆	4	6	1
7.	Diosmetin	300.26 g/mol	C ₁₆ H ₁₂ O ₆	3	6	2
8.	Limonin	470.5 g/mol	C ₂₆ H ₃₀ O ₈	0	8	1
9.	Tangeretin	372.4 g/mol	C ₂₀ H ₂₀ O ₇	0	7	6
10.	Captopril	217.29 g/mol	C ₉ H ₁₅ NO ₃ S	2	4	3

Ligand Properties of the Compounds selected for docking against Acetyl Cholinesterase (1B41)

Table 2. 2D and 3D structure of selected ligands of *Citrus aurantium* compounds

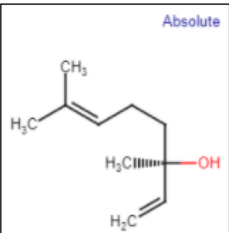
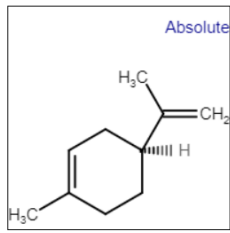
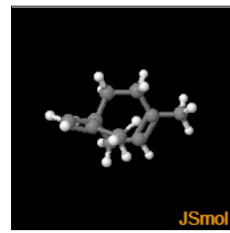
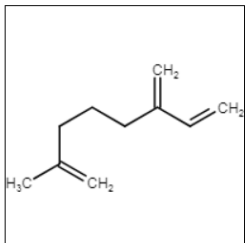
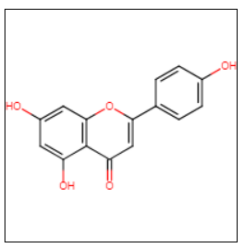
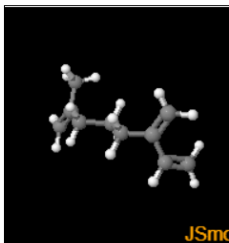
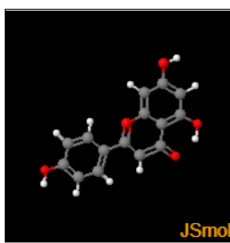
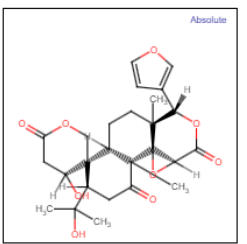
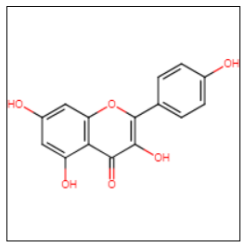
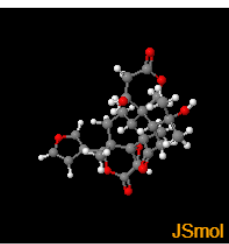
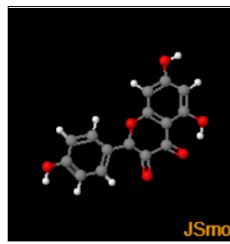
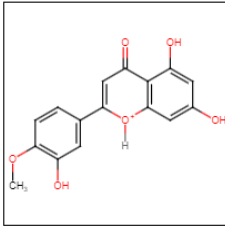
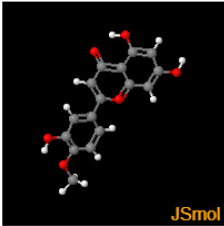
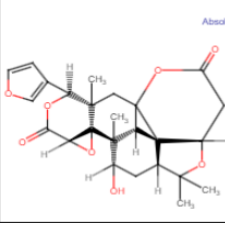
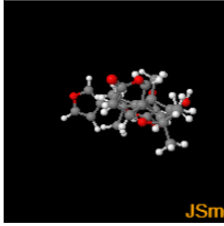
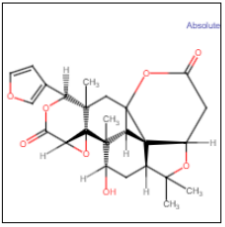
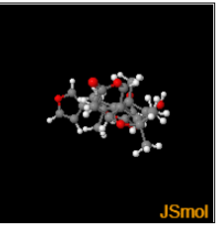
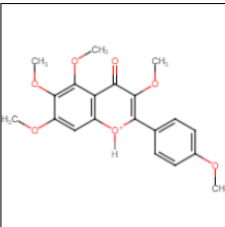
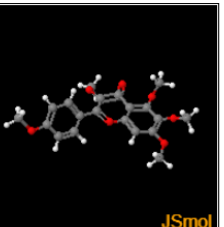
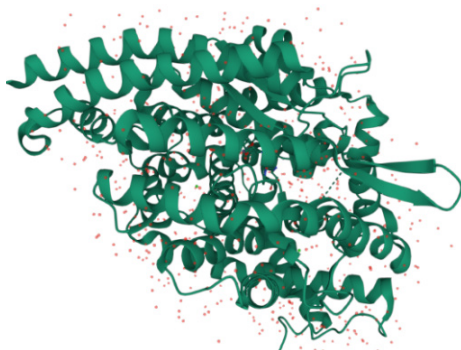
2D and 3D Structure of selected phytocomponents			
Linalool Ligand in 2D 		Limonene Ligand in 2D 	
Ligand in 3D 		Ligand in 3D 	
Myrcene Ligand in 2D 		Apigenin Ligand in 2D 	
Ligand in 3D 		Ligand in 3D 	
Ichangin Ligand in 2D 		Kaempferols Ligand in 2D 	
Ligand in 3D 		Ligand in 3D 	

Table 2. to be continued...

Luteolin Ligand in 2D  Ligand in 3D  JSmol	Diosmetin Ligand in 2D  Ligand in 3D  JSmol
Limonin Ligand in 2D  Ligand in 3D  JSmol	Tangeretin Ligand in 2D  Ligand in 3D  JSmol

Human Angiotensin Converting Enzyme (1O86)

**Figure 3.** 3D protein structure of Human angiotensin converting enzyme (1O86).

with ligand atoms. The retrieved phytochemicals Linalool, Apigenin, Ichangin, Kaempferol, Luteolin, Limonin, Limonene, Myrcene, Diosmetin and Tangeretin were subjected to docking calculations against ACE using AutoDock 4. With the aid of the AutoDock 4 virtual screening tool, required parameters were added, and with the help of the Autogrid program, Affinity (grid) maps with $60 \times 60 \times 60$ Å grid points, 0.375 Å spacing were generated. To calculate the van der Waals and the electrostatic terms, the AutoDock parameter set- and distance-dependent dielectric functions were used. The Lamarckian genetic algorithm and the Solis and Wets local search method were used for

docking simulations and Torsions were released, setting the Initial position, orientation, and torsions of the ligand molecules randomly. Each docking experiment was subjected to two different runs that were fixed to terminate after energy evaluations of 250,000 with a population size of 150 and applying a translational step of 0.2 Å, and quaternion and torsion steps of 5^{33} .

2.2.6 Rigid Docking

To get a group of permissible orientations and conformations for the ligand at the binding site. Computational docking was carried out at the active site of spike protein of the selected phytochemicals with MGL AutoDock Tool-Autodock Vina 1.1.2.

2.2.7 Visualization

The Discovery Studio Visualizer program version 2016 was used to analyse the interaction of hydrogen bonds and aminoacids.

3. Results

Ten bioactive lead compounds of *Citrus aurantium* fruit, such as Linalool, Apigenin, Ichangin, Kaempferol, Luteolin, Limonin and Tangeretin were retrieved. Their interactions with the receptor site of amino acid residues present on ACE was determined. The details are tabulated in Tables 3-5.

Table 3. Summary of *in silico* analysis of phytochemicals against ACE (1O86)

Compounds	Binding Free energy Kcal/mol	Inhibition constant Ki μM (*mM)(**nM)	Electrostatic energy Kcal/mol	Intermolecular energy Kcal/mol	Total Interaction Surface
Linalool	-4.04 kcal/mol	1.10 mM	-0.16 kcal/mol	-5.49 kcal/mol	534.936
Limonene	-4.45 kcal/mol	546.45 μM	-0.02 kcal/mol	-4.75 kcal/mol	436.552
Myrcene	-4.38 kcal/mol	616.81 μM	-0.01 kcal/mol	-5.52 kcal/mol	480.415
Apigenin	-5.10 kcal/mol	182.65 μM	-0.74 kcal/mol	-6.60 kcal/mol	731.524
Ichangin	-8.45 kcal/mol	639.78 μM	-0.11 kcal/mol	-7.62 kcal/mol	1049.019
Kaempferol	-5.34 kcal/mol	121.82 μM	-0.10 kcal/mol	-5.71 kcal/mol	704.237
Luteolin	-6.66 kcal/mol	13.17 μM	-0.60 kcal/mol	-6.29 kcal/mol	707.988
Diosmetin	-6.30 kcal/mol	24.25 μM	-0.18 kcal/mol	-6.61 kcal/mol	650.991
Limonin	-7.23 kcal/mol	5.01 μM	-0.13 kcal/mol	-7.47 kcal/mol	1022.766
Tangeretin	-5.00 kcal/mol	216.20 μM	-0.17 kcal/mol	-6.53 kcal/mol	829.921
Captopril	-5.09 kcal/mol	186.95 μM	-0.61 kcal/mol	-5.13 kcal/mol	504.357

Table 4. Interaction of amino acid residue of phytochemicals and captopril ACE

S. No.	Protein	Phyto chemical	Affinity	RMSD (I. B.)	RMSB (U. B.)	No. of Hydrogen Bonds	Ligand – Allele Interaction (Non-bound)	Distance	Category
a	ACE2	Linalool	-5.3	9.227	11.55	1	:UNL1:H - A:GLN102:O	1.99989	Hydrogen Bond
							:UNL1 - A:LEU73	4.88994	Hydrophobic
							:UNL1 - A:LEU100	5.49351	Hydrophobic
							:UNL1:C - A:ALA99	4.48489	Hydrophobic
							:UNL1:C - A:LEU100	4.68179	Hydrophobic
							:UNL1:C - A:LEU391	4.93524	Hydrophobic
b	ACE2	Limonene	-5.3	37.54	38.868	0	:UNL1:C - A:LEU95	4.65071	Hydrophobic
							:UNL1:C - A:VAL209	5.16234	Hydrophobic
							:UNL1:C - A:VAL212	4.44133	Hydrophobic
							:UNL1:C - A:PRO565	4.6178	Hydrophobic
							:UNL1:C - A:VAL209	4.81661	Hydrophobic
							A:LEU95 - :UNL1	4.34903	Hydrophobic
							A:VAL209 - :UNL1	4.05884	Hydrophobic
							A:PRO565 - :UNL1	4.4849	Hydrophobic
							A:TRP566 - :UNL1:C	4.92793	Hydrophobic
c	ACE2	Myrcene	-5.1	50.354	51.875	0	:UNL1 - A:LEU95	4.86109	Hydrophobic
							:UNL1:C - A:LEU95	4.13041	Hydrophobic
							:UNL1:C - A:VAL209	4.48599	Hydrophobic
							:UNL1:C - A:PRO565	3.93989	Hydrophobic
							A:LYS562 - :UNL1	4.96802	Hydrophobic

Table 4. *to be continued...*

S. No.	Protein	Phyto chemical	Affinity	RMSD (I. B.)	RMSB (U. B.)	No. of Hydrogen Bonds	Ligand – Allele Interaction (Non-bound)	Distance	Category
d	ACE2	Apigenin	-7.4	34.271	37.007	4	:UNK0:H - A:ASP509:OD2	2.76684	Hydrogen Bond
							A:SER511:HN - :UNK0:O	1.95281	Hydrogen Bond
							A:SER511:HG - :UNK0:O	2.47835	Hydrogen Bond
							A:ARG514:HH12 - :UNK0:O	2.68592	Hydrogen Bond
							:UNK0 - A:TRP203	5.54802	Hydrophobic
							A:TRP203 - :UNK0	4.6941	Hydrophobic
							A:TRP203 - :UNK0	4.29446	Hydrophobic
							A:GLY205:C,O; ASP206:N - :UNK0	4.43928	Hydrophobic
e	ACE2	Ichangin	-9.2	0	0	6	:UNL1:H - A:ASP382:OD1	2.42224	Hydrogen Bond
							:UNL1:H - A:ASP382:OD1	2.83962	Hydrogen Bond
							:UNL1:H - A:TYR385:OH	3.09578	Hydrogen Bond
							A:TYR385:HH - :UNL1:O	2.83545	Hydrogen Bond
							A:ASN394:HD21 - :UNL1:O	2.66288	Hydrogen Bond
							A:HIS401:HE2 - :UNL1:O	2.48791	Hydrogen Bond
							:UNL1:C - A:HIS401	3.97227	Hydrophobic
							:UNL1:C - A:HIS401	3.83153	Hydrophobic
f	ACE2	Kaempferols	-8.9	8.524	13.03	9	:UNK0:H - A:ASP206:OD1	2.43929	Hydrogen Bond
							:UNK0:H - :UNK0:O	1.7923	Hydrogen Bond
							:UNK0:H - A:SER511:OG	2.71878	Hydrogen Bond
							:UNK0:H - A:ASP509:OD2	2.55348	Hydrogen Bond
							A:TYR199:HH - :UNK0:O	2.61501	Hydrogen Bond
							A:SER511:HN - :UNK0:O	1.8683	Hydrogen Bond
							A:SER511:HG - :UNK0:O	2.59959	Hydrogen Bond
							A:ARG514:HH12 - :UNK0:O	2.59917	Hydrogen Bond
							A:ASP206:OD1 - :UNK0	4.42701	Electrostatic
							:UNK0 - A:TRP203	5.56745	Hydrophobic
							A:TRP203 - :UNK0	4.85631	Hydrophobic
							A:TRP203 - :UNK0	4.33686	Hydrophobic
g	ACE2	Luteolin	-8.2	15.289	17.927	5	:UNL1:H - :UNL1:O	1.80092	Hydrogen Bond
							:UNL1:H - A:GLU398:O	2.29594	Hydrogen Bond
							:UNL1:H - A:ALA348:O	2.69512	Hydrogen Bond
							:UNL1:H - A:ASP382:OD2	2.80261	Hydrogen Bond
							A:ARG514:HE - :UNL1:O	2.13238	Hydrogen Bond
							A:HIS401:NE2 - :UNL1	3.6884	Electrostatic
							A:HIS401 - :UNL1	5.2828	Hydrophobic
							A:HIS401 - :UNL1	3.79672	Hydrophobic

Table 4. to be continued...

S. No.	Protein	Phyto chemical	Affinity	RMSD (I. B.)	RMSB (U. B.)	No. of Hydrogen Bonds	Ligand – Allele Interaction (Non-bound)	Distance	Category
h	ACE2	Diosmetin	-8.2	13.366	15.315	7	:UNL1:H - :UNL1:O	1.80115	Hydrogen Bond
							:UNL1:H - A:ASN394:OD1	2.7819	Hydrogen Bond
							:UNL1:H - A:ASP206:OD1	2.67978	Hydrogen Bond
							A:GLY395:HN - :UNL1:O	2.63704	Hydrogen Bond
							A:ARG514:HE - :UNL1:O	2.93816	Hydrogen Bond
							A:ARG514:HH21 - :UNL1:O	2.26149	Hydrogen Bond
							A:HIS401:CD2 - :UNL1:O	3.46778	Hydrogen Bond
							A:ASN397:C,O;GLU398:N - :UNL1	4.62598	Hydrophobic
							A:ASN397:C,O;GLU398:N - :UNL1	4.16251	Hydrophobic
i	ACE2	Limonin	-8.9	1.8	7.878	0	:UNL1:C - A:PRO415	4.95867	Hydrophobic
							:UNL1:C - A:ILE291	5.03567	Hydrophobic
							A:ILE291 - :UNL1	4.1403	Hydrophobic
							A:PRO415 - :UNL1	4.97759	Hydrophobic
							A:PHE438 - :UNL1	4.50987	Hydrophobic
							A:PHE438 - :UNL1:C	5.03793	Hydrophobic
j	ACE2	Tangeretin	-8.3	3.225	8.494	6	A:GLN102:HE22 - :UNL1:O	2.19866	Hydrogen bond
							A:TYR196:HH - :UNL1:O	2.411	Hydrogen bond
							A:LYS562:HZ1 - :UNL1:O	2.2789	Hydrogen bond
							:UNL1:C - A:ASP206:O	3.5189	Hydrogen bond
							:UNL1:C - A:GLN102:OE1	3.47682	Hydrogen bond
							A:ASN210:HN - :UNL1	3.23243	Hydrogen bond
							:UNL1:C - A:LEU95	5.12456	Hydrophobic
							:UNL1:C - A:VAL209	3.73609	Hydrophobic
							:UNL1:C - A:VAL212	4.66831	Hydrophobic
							:UNL1:C - A:PRO565	3.659	Hydrophobic
							:UNL1 - A:LEU95	4.88785	Hydrophobic
							:UNL1 - A:VAL209	4.93861	Hydrophobic

Table 4 shows the amino acid residue interaction of (a) Linalool, b) Limonene, c) Myrcene, d) Apigenin, e) Ichangin, f) Kaempferol, g) Luteolin, h) Diosmetin, i) Limonin, j) Taneretin and captopril against the structure of ACE in crystal form.

4. Discussion

Hypertension is one of the non-communicable diseases in high prevalence in our country, causing complications of the cardiovascular system and also a

Table 5. 2D interaction plot analysis and docking pose of selected phytochemicals present in *Citrus aurantium* with ACE

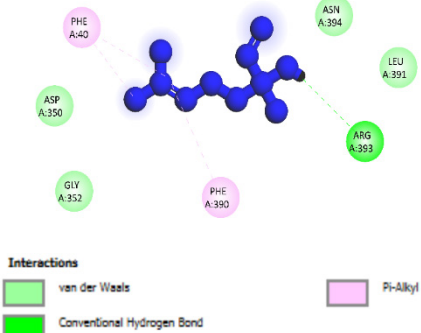
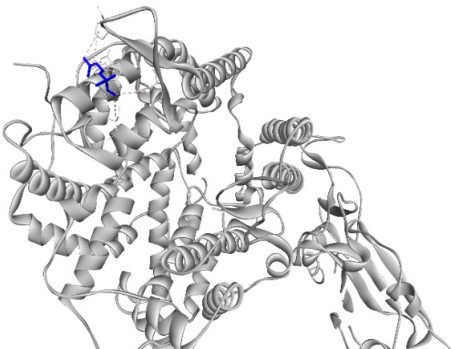
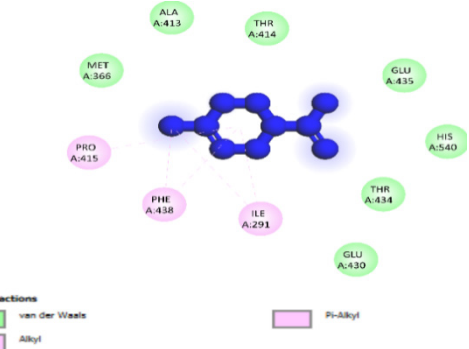
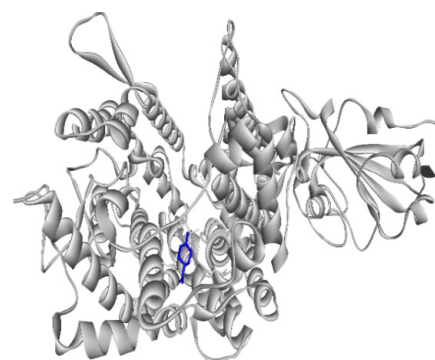
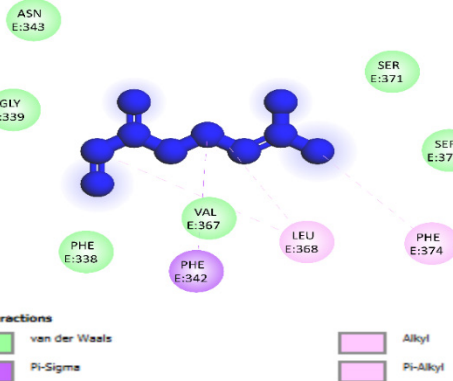
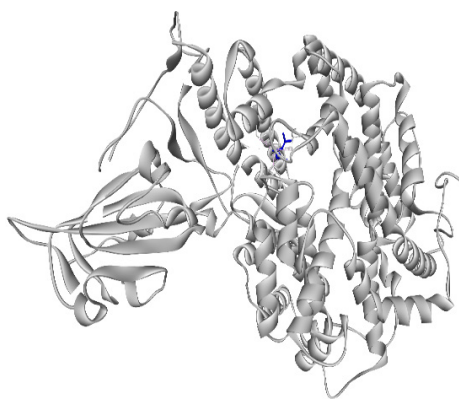
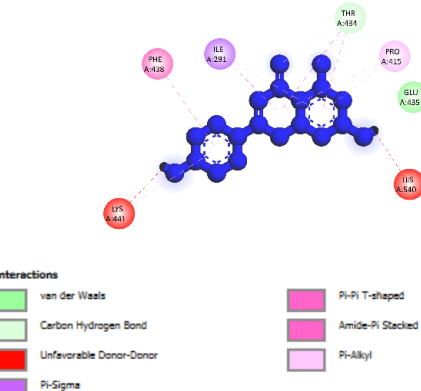
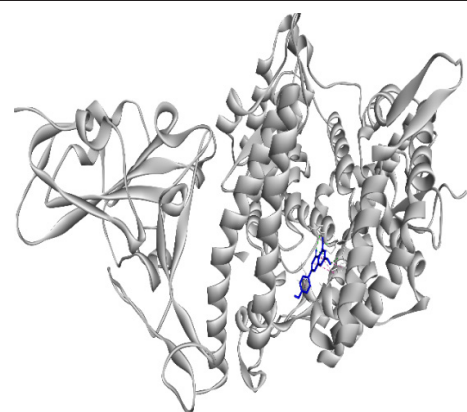
Phytochemicals of <i>Citrus aurantium</i>	2D Interaction plot analysis	Docking pose phytochemicals with ACE
Linalool	 Interactions - van der Waals - Conventional Hydrogen Bond - Pi-Alkyl	
Limonene	 Interactions - van der Waals - Alkyl - Pi-Alkyl	
Myrcene	 Interactions - van der Waals - Pi-Sigma - Alkyl - Pi-Alkyl	
d) Apigenin	 Interactions - van der Waals - Carbon Hydrogen Bond - Unfavorable Donor-Donor - Pi-Sigma - Pi-Pi T-shaped - Amide-Pi Stacked - Pi-Alkyl	

Table 5. to be continued...

Phytochemicals of <i>Citrus aurantium</i>	2D Interaction plot analysis	Docking pose phytochemicals with ACE
e) Ichangin	Interactions - van der Waals - Conventional Hydrogen Bond - Pi-Sigma	
f) Kaempferols	Interactions - van der Waals - Conventional Hydrogen Bond - Pi-Sigma - Pi-Alkyl	
g) Luteolin	Interactions - van der Waals - Conventional Hydrogen Bond - Unfavorable Donor-Donor - Pi-Sigma - Pi-Alkyl	
h) Diosmetin	Interactions - van der Waals - Conventional Hydrogen Bond - Carbon Hydrogen Bond - Pi-Sigma - Pi-Alkyl	

Table 5. to be continued...

Phytochemicals of <i>Citrus aurantium</i>	2D Interaction plot analysis	Docking pose phytochemicals with ACE
i) Limonin		
j) Tangeretin		

high mortality rate among the middle aged as well as elderly population. The study was carried out to find the efficacy of the molecules with lead activity to bring out the core bioactive amino acid residues, GLU162, LYS511, HIS513, TYR520, and TYR523, which mediates the enzymatic action of the ACE. Since ACE is involved in converting Angiotensin I to II, which constricts blood vessels and raises blood pressure, reducing the enzyme concentration of ACE will inhibit the conversion and in turn could be effective in controlling the elevation of blood pressure³⁴.

The enzymatic action of ACE could be reduced by occupying the receptor sites. Phytochemicals that have the potency to inhibit the enzyme ACE could

lower the high blood pressure. Captopril is a synthetic antihypertensive drug developed by structural modification of venom of Brazilian viper acts as an ACE inhibitor. Long-term use of ACE inhibitors were known to produce serious irreversible side effects, such as lightheadedness, cough, and angio edema. Such effect could be overcome by replace synthetic drugs with natural medicine. Natural ACE inhibitors of plant origin were known to inhibit ACE. The mechanism of such biocomponents resemble synthetic ACE inhibitors³⁵.

In Siddha literature single herbs such as *Cuminum cyminum*, *Glyzrrhiza glabra*, *Citrus limon*, *Vetiveria zizanooides*, *Eletaria cardamum*, *Terminalia bellarica*,

Oldenlandia umbellata, *Allium cepa* and prepared medicines like *Thiratchadi kudineer*, *Nerunjil kudineer*, *Seeraga chooranam*, *Maruthampattai chooranam*, *Inji ilagam*, *Kesari ilagam* are also recommended for the hypertension management. The same has been emphasized in the Siddha treatment guidelines for Hypertension published by Siddha Clinical Research Institute, Chennai¹⁸. The selected fruit is used in the treatment of hypertension by the people of Edo state, Enderta, Northern Ethiopia and Nigeria^{36,37}. Other than the suggested pharmacological activity, the phyto components of *Citrus aurantium* are said to have protective activity against gastrointestinal lesions induced by alcohol and NSAIDS in rats⁹ and antimicrobial activity against pathogens of *Salmonella species*. Also, its antidiabetic effects were well established by various studies. Extract of *C. aurantium* has been commonly used in dietary supplements for weight loss and to enhance sports performance¹⁰⁻¹⁶.

Among 32 phytochemicals, components such as Linalool, Limonene, Myrcene, Apigenin, Ichangin, Kaempferols, Luteolin, Diosmetin, Limonin, Tangeretin were selected based on their activity from published articles³². Phytochemical data, including the molecular structure of the selected compounds were retrieved using search engines, IMPPAT (Indian Medicinal Plants, Phytochemistry and Therapeutics). Since pharmacological screening of all the phytochemicals is time-consuming and costly, molecular docking of selected phytochemicals was done with computer-aided drug design, the results are mentioned in Tables 4 and 5.

The hydrogen bond counts present in the phytoconstituent is said to have a favourable contribution to affinity towards the target protein. Of the ten bioactive lead compounds retrieved from the herb *Citrus aurantium*, Kaempferol has the highest number, i.e. 9 interactions with the active site of the target enzyme ACE when compared to Captopril which reveals 8 interactions over the target enzyme. Diosmetin is ranked second with 7 interactions, whereas Tangeretin and Ichangin are with 6 interactions. Luteolin Apigenin and Linalool have 5, 4 and 1 hydrogen bonds, respectively. Limonin, Limonene and Myrcene possess only hydrophobic bonds that are said to be unfavourable for contribution towards affinity. Kaempferol has a binding affinity of -8.9, Limonin

-8.9, Ichangin- 9.2, Tangeretin -8.3, Diosmetin and Luteolin - 8.2, Apigenin -7.4, Limonene and Linalool -5.3 Myrcene -5.1.

5. Conclusion

The docking study results prove that the phytochemical Kaempferol shows about 9 interactions against the active site of ACE. Diosmetin, Tangeretin and Ichangin also have more than 5 hydrogen bonds for active interaction. Results of the *in silico* analysis suggest that the phytochemical Kaempferol, Diosmetin, Tangeretin and Ichangin present in *Citrus aurantium*, the prime ingredient of *Thurunji Manapaagu* reveals effective binding to the active receptor site of ACE that supports the conclusion that *Thurunji Manapaagu* could have a promising anti-hypertensive activity.

6. Acknowledgement

The authors wish to acknowledge the support and guidance of the Head of Institute of Siddha Clinical Research Unit, Tirupati.

7. References

1. Gentis N, Licht A, Boura A, De Franceschi D, Win Z, Aung DW, Dupont-Nivet G. Fossil wood from the lower Miocene of Myanmar (Natma Formation): palaeoenvironmental and biogeographic implications. *Geodiversitas*. 2022; 44(28): 853-909. <https://doi.org/10.5252/geodiversitas2022v44a28>
2. Khan H, Nabavi SM, Sureddi A, Mehterov N, Gulei D, Berindan-Neagoe I, *et al.* Therapeutic potential of songorine, a diterpenoid alkaloid of the genus *Aconitum*. *European Journal of Medicinal Chemistry*. 2018; 153:29-33. <https://doi.org/10.1016/j.ejmech.2017.10.065>
3. Nabavi SF, Khan H, D'onofrio G, Šamec D, Shirooie S, Dehpour AR, *et al.* Apigenin as neuroprotective agent: of mice and men. *Pharmacological Research*. 2018; 128:359-65. <https://doi.org/10.1016/j.phrs.2017.10.008>
4. Khan H, Nabavi SM, Habtemariam S. Anti-diabetic potential of peptides: Future prospects as therapeutic agents. *Life Sciences*. 2018; 193:153-8. <https://doi.org/10.1016/j.lfs.2017.10.025>
5. Lim SW, Lee DR, Choi BK, Kim HS, Yang SH, Suh JW, *et al.* Protective effects of a polymethoxy flavonoids-rich *Citrus aurantium* peel extract on liver fibrosis induced by bile duct ligation in mice. *Asian Pacific Journal of Tropical Medicine*. 2016; 9(12):1158-64. <https://doi.org/10.1016/j.apjtm.2016.10.009>

6. Zhao HY, Yang L, Wei J, Huang M, Jiang JG. Bioactivity evaluations of ingredients extracted from the flowers of *Citrus aurantium* L. var. *amara* Engl. Food chemistry. 2012; 135(4):2175-81. <https://doi.org/10.1016/j.foodchem.2012.07.018>
7. Costa CA, Cury TC, Cassettari BO, Takahira RK, Flório JC, Costa M. *Citrus aurantium* L. essential oil exhibits anxiolytic-like activity mediated by 5-HT 1A-receptors and reduces cholesterol after repeated oral treatment. BMC Complementary and Alternative Medicine. 2013; 13. <https://doi.org/10.1186/1472-6882-13-42>
8. Akhlaghi M, Shabani G, Rafeian-Kopaei M, Parvin N, Saadat M, Akhlaghi M. *Citrus aurantium* blossom and preoperative anxiety. Revista Brasileira de Anestesiologia. 2011;61:707-12. <https://doi.org/10.1590/S0034-70942011000600002>
9. Moraes TM, Kushima H, Moleiro FC, Santos RC, Rocha LR, Marques MO, et al. Effects of limonene and essential oil from *Citrus aurantium* on gastric mucosa: Role of prostaglandins and gastric mucus secretion. Chemico-Biological Interactions. 2009; 180(3):499-505. <https://doi.org/10.1016/j.cbi.2009.04.006>
10. Karabiyikli Ş, Değirmenci H, Karapınar M. Inhibitory effect of sour orange (*Citrus aurantium*) juice on *Salmonella typhimurium* and *Listeria monocytogenes*. LWT-Food Science and Technology. 2014; 55(2):421-5. <https://doi.org/10.1016/j.lwt.2013.10.037>
11. Colker CM, Kaiman DS, Torina GC, Perlis T, Street C. Effects of *Citrus aurantium* extract, caffeine, and St. John's wort on body fat loss, lipid levels, and mood states in overweight healthy adults. 1999; 60(3):145-153. [https://doi.org/10.1016/S0011-393X\(00\)88523-9](https://doi.org/10.1016/S0011-393X(00)88523-9)
12. Stohs SJ, Preuss HG, Shara M. Issues regarding a FACT review paper on the efficacy of herbal supplements containing *Citrus aurantium* and synephrine alkaloids for the management of overweight and obesity. Focus on Alternative and Complementary Therapies. 2013; 1(18):45-7. <https://doi.org/10.1111/fct.12005>
13. Haaz S, Fontaine KR, Cutter G, Limdi N, Perumean-Chaney S, Allison DB. *Citrus aurantium* and synephrine alkaloids in the treatment of overweight and obesity: An update. Obesity reviews. 2006; 7(1):79-88. <https://doi.org/10.1111/j.1467-789X.2006.00195.x>
14. Fabio A, Cermelli C, Fabio G, Nicoletti P, Quaglio P. Screening of the antibacterial effects of a variety of essential oils on microorganisms responsible for respiratory infections. Phytotherapy Research. 2007; 21(4):374-7. <https://doi.org/10.1002/ptr.1968>
15. Oliveira SAC, Zambrana JRM., Di Iorio FBR, Pereira CA, Jorge AOC. The antimicrobial effects of *Citrus limonum* and *Citrus aurantium* essential oils on multi-species biofilms. 2014; 28(1):22-7. <https://doi.org/10.1590/S1806-83242013005000024>
16. Meléndez PA, Capriles VA. Antibacterial properties of tropical plants from Puerto Rico. 2006; 13(4):272-6. <https://doi.org/10.1016/j.phymed.2004.11.009>
17. Xing H, Zhang K, Zhang R, Shi H, Bi K, Chen X. Antidepressant-like effect of the water extract of the fixed combination of *Gardenia jasminoides*, *Citrus aurantium* and *Magnolia officinalis* in a rat model of chronic unpredictable mild stress. Phytomedicine. 2015; 22(13):1178-85. <https://doi.org/10.1016/j.phymed.2015.09.004>
18. Siddha Central Research Institute. Siddha treatment guideline for selected Non-communicable Diseases. Chennai: Siddha Central Research Institute; 2014. p. 245-63.
19. Staessen JA, Wang J, Bianchi G, Birkenhäger WH. Essential Hypertension. The Lancet. 2003; 361(9369):1629-41. [https://doi.org/10.1016/S0140-6736\(03\)13302-8](https://doi.org/10.1016/S0140-6736(03)13302-8)
20. World Health Organization. Hypertension. Available from: https://www.who.int/health-topics/hypertension#tab=tab_1
26. Uthamarayan KN, Mudhaliyar KSM. Siddha Vaidhiya Thirattu, Department of Indian Medicine and Homeopathy, Chennai-106. First Edition. Pp. 259.
22. Suryawanshi JA. An overview of *Citrus aurantium* used in treatment of various diseases. African Journal of Plant Science. 2011; 5(7):390-395.
23. Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, et al. A novel Angiotensin-Converting Enzyme-related carboxypeptidase (ACE2) converts Angiotensin I to Angiotensin 1-9. Circulation Research. 2000; 87(5):e1-e9. <https://doi.org/10.1161/01.RES.87.5.e1>
24. Yusuf S, Sleight P, Pogue JF, Bosch J, Davies R, Dagenais G. Effects of an Angiotensin-Converting-Enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. The New England journal of medicine. 2000; 342(3):145-153. <https://doi.org/10.1056/NEJM200001203420301>
25. Pozzan A. Molecular descriptors and methods for ligand based virtual high throughput screening in drug discovery. Current Pharmaceutical Design. 2006; 12(17):2099-2110. <https://doi.org/10.2174/13816120677585247>
26. Morris GM, Lim-Wilby M. Molecular modeling of proteins. Methods in Molecular Biology. 2008; 443:365-82. https://doi.org/10.1007/978-1-59745-177-2_19
27. AlSheikh HM, Sultan I, Kumar V, Rather IA, Al-Sheikh H, Tasleem Jan A, et al. Plant-based phytochemicals as possible alternative to antibiotics in combating bacterial drug resistance. Antibiotics. 2020; 9(8):480. <https://doi.org/10.3390/antibiotics9080480>
28. Pinzi L, Rastelli G. Molecular docking: Shifting paradigms in drug discovery. International Journal of Molecular Sciences. 2019; 20(18):433. <https://doi.org/10.3390/ijms20184331>
29. Jia Z, Song X, Shi J, Wang W, He K. Transcriptome-based drug repositioning for coronavirus disease 2019 (COVID-19). Pathogens and Disease. 2020; 78(4). <https://doi.org/10.1093/femspd/ftaa036>

30. Abdullahi M, Adeniji SE. *In-silico* molecular docking and ADME/pharmacokinetic prediction studies of some novel carboxamide derivatives as anti-tubercular agents. *Chemistry Africa*. 2020; 3(4):989-1000. <https://doi.org/10.1007/s42250-020-00162-3>
31. Abdullahi M, Shallangwa GA, Uzairu A. *In silico* QSAR and molecular docking simulation of some novel aryl sulfonamide derivatives as inhibitors of H5N1 influenza A virus subtype. *Beni-Suef University Journal of Basic and Applied Sciences*. 2020; 9(1). <https://doi.org/10.1186/s43088-019-0023-y>
32. Suntar I, Khan H, Patel S, Celano R, Rastrelli L. An overview on *Citrus aurantium* L.: Its functions as food ingredient and therapeutic agent. *Oxidative Medicine and Cellular longevity*. 2018; 2018. <https://doi.org/10.1155/2018/7864269>
33. Morris GM, Goodsell DS, Halliday RS, Huey R, Hart WE, Belew RK, *et al*. Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *Journal of Computational Chemistry*. 1998; 19(14):1639-1662. [https://doi.org/10.1002/\(SICI\)1096-987X\(19981115\)19:14<1639::AID-JCC10>3.0.CO;2-B](https://doi.org/10.1002/(SICI)1096-987X(19981115)19:14<1639::AID-JCC10>3.0.CO;2-B)
34. Bikadi Z, Hazai E. Application of the PM6 semi-empirical method to modeling proteins enhances docking accuracy of AutoDock. *Journal of Cheminformatics*. 2009; 1:1-6. <https://doi.org/10.1186/1758-2946-1-15>
35. Passos-Silva DG, Brandan E, Santos RA. Angiotensins as therapeutic targets beyond heart disease. *Trends in Pharmacological Sciences*. 2015; 36(5):310-20. <https://doi.org/10.1016/j.tips.2015.03.001>
36. Chakraborty R, Roy S. Angiotensin-converting enzyme inhibitors from plants: A review of their diversity, modes of action, prospects, and concerns in the management of diabetes-centric complications. *Journal of Integrative Medicine*. 2021; 19(6):478-92. <https://doi.org/10.1016/j.joim.2021.09.006>
37. Mensah JK, Okoli RI, Turay AA, Ogie-Odia EA. Phytochemical analysis of medicinal plants used for the management of hypertension by Esan people of Edo state, Nigeria. *Ethnobotanical Leaflets*. 2009; 2009(10):7.