

Adipose Tissue Dysfunction in PCOS

Ananya Aparupa and Rita Singh*

Division of Molecular Endocrinology and Reproduction, Department of Zoology, University of Delhi, Delhi - 110007, India; ritas@zoology.du.ac.in

Abstract

Polycystic Ovary Syndrome (PCOS) is one of the most common endocrine diseases among women of reproductive age; however, its aetiology is unclear. PCOS is linked to many metabolic manifestations and alterations such as obesity, insulin resistance, and cardiovascular diseases (CVD). Women with PCOS have intra-ovarian and systemic changes in their metabolite levels. Adipose tissue dysfunction plays a significant role in the pathophysiology of PCOS. Adipose tissue growth is disrupted by metabolic stress, leading to hypertrophy of adipocytes, which begin to express stress signals. Adipose tissue secretes autocrine and paracrine factors, called adipokines or adipocytokines. Adiponectin is an adipocyte-derived protein abundant in the bloodstream. Plasma adiponectin concentration is low in women with PCOS, obesity, CVD, and hypertension. Other adipocytokines with altered secretion in PCOS include leptin, resistin, apelin, visfatin, IL-6, IL-8, and TNF- α . Hormonal imbalance, untimely action of high LH, and consequent hyperandrogenism in women with PCOS may cause metabolic defects associated with adipose tissue dysfunction; however, there are no reports on the role of higher LH levels in adipose dysfunction and altered adipokine secretion. New medications with therapeutic potential have been developed that target adipokines for the treatment of PCOS. This review discusses the association between PCOS and altered adipokine production as a consequence of adipose dysfunction.

Keywords: Adipocyte, Adiponectin, Adipose Tissue, Follicle Stimulating Hormone, Luteinizing Hormone, PCOS

1. Introduction

Polycystic Ovary Syndrome (PCOS), or Stein-Leventhal syndrome, is a complex endocrine abnormality that affects 5 to 10% of women of reproductive age and accounts for 50-70 % of cases with anovulatory infertility. It is the most common endocrine disorder in women with menstrual irregularities and anovulation. PCOS is characterized by hyperandrogenism, anovulation, polycystic morphology of the ovary, and menstrual irregularities¹⁻³ [Figure 1]. One of the main endocrinological disturbances in women with PCOS is the increased levels of Luteinizing Hormone (LH), and consequently a high LH:FSH ratio¹⁻⁵. Neuroendocrine defects in estrogen and progesterone feedback mechanisms lead to gonadotropin imbalance and hyperandrogenism in women⁶. High androgen levels also alter the hypothalamic inhibitory feedback of progesterone. Along with the reproductive anomalies observed in PCOS, metabolic defects including obesity, insulin resistance, T2DM, and CVD are also strongly

associated with PCOS [Figure 1]. Women with PCOS also have an elevated risk of endometrial cancer⁷ and early atherosclerosis⁸.

In addition to overall obesity, women with PCOS are more likely to exhibit an abdominal or visceral fat distribution⁹. These depots are associated with an unfavorable metabolic profile and insulin resistance¹⁰. There is strong evidence linking androgen excess to insulin resistance¹⁰. Furthermore, hyperinsulinemia stimulates ovarian androgen production and decreases the hepatic production of Sex Hormone-Binding Globulin (SHBG)^{4,10,11}. Some studies have strongly suggested that genetic factors play an important role in PCOS. Many potential genes implicated in insulin resistance, secretion, and ovarian and adrenal steroidogenesis have been positively associated with PCOS^{10,12}. β -cell dysfunction in addition to peripheral insulin resistance is reported in women with PCOS^{4,13}.

Metabolic abnormalities and obesity are reported in 50-60 % of women with PCOS, strongly associated

*Author for correspondence

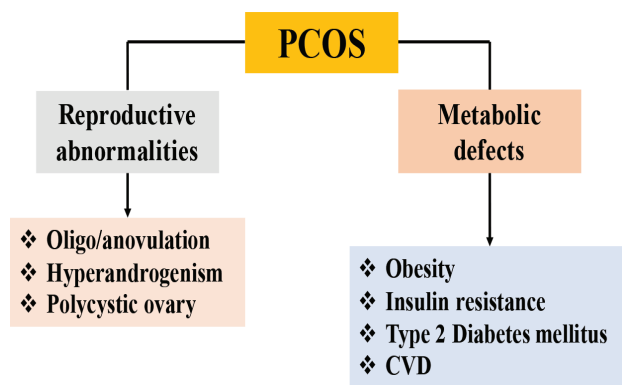


Figure 1. PCOS is characterized by reproductive abnormalities that lead to infertility, and these women are more likely to develop metabolic disorders.

with a greater waist-to-hip ratio, larger adipocytes, decreased adiponectin levels, and lower lipoprotein lipase activity^{8,14}. In women with PCOS, the prevalence of dyslipidemia was reported to be 70%. Low-density lipoprotein cholesterol, free fatty acid, and triglyceride levels are elevated in women with PCOS¹⁵. On the contrary, high-density lipoprotein cholesterol levels are reduced in women with PCOS^{16,17}. Numerous studies have established that Non-Alcoholic Fatty Liver Disease (NAFLD) is highly prevalent in women with PCOS¹⁸. The close association between PCOS and metabolic syndrome has led to the assumption that dysregulation of adipose tissue function may contribute to many of the metabolic abnormalities observed in women with PCOS, as it is known to occur in metabolic syndrome. Here, we discuss the association of altered adipokine production as a consequence of adipose dysfunction in women with PCOS.

2. Methods

We searched PubMed for articles published in English. The search terms used were as follows: polycystic ovary syndrome, polycystic ovary disease, LH, hCG, insulin resistance, adipocytes, White Adipose Tissue (WAT), adipocytokines, adipose tissue dysfunction, obesity, visceral obesity, and metabolic syndrome. Studies that evaluated adipose tissue function in women with PCOS identified by the National Institute of Health (1990), Rotterdam Criteria (2003), and/or Androgen Excess and PCOS Society (AE-PCOS) criteria (2006) and compared findings with controls with normal Body Mass Index (BMI) were considered eligible for inclusion. We

broadened the search to include references from the retrieved articles.

3. Adipose Tissue Dysfunction in PCOS

Adipose tissue is a loose connective tissue composed of mature adipocytes, pre-adipocytes, mesenchymal cells, and various immune cells. Mesenchymal cells differentiate into preadipocytes and mature adipocytes¹⁹. The main function of adipose tissue is to insulate the body, store Free Fatty Acids (FFAs) after food consumption, and release FFAs during a fasting state to maintain a suitable energy level. After triglyceride hydrolysis, FFAs are absorbed from the circulation in adipose tissue during the postprandial period. Following the hydrolysis of TG from triglyceride-rich lipoproteins (chylomicrons, Very Low-Density Lipoprotein Cholesterol (VLDL-C), and their remnants) by lipoprotein lipase, adipose tissue absorbs FFAs from the blood during the postprandial period. Hormone-Sensitive Lipase (HSL) breaks down intracellular TG to release fatty acids. Changes in adipose tissue growth occur owing to the differentiation and proliferation of adipocyte progenitors, which increase the number of adipocytes and replace larger, older, and defective adipocytes. In response to chronic changes in energy balance and nutrient content, the proliferation of preadipocytes, differentiation into mature adipocytes, and adipocyte growth are affected, leading to hypertrophy, and eventually adipocyte apoptosis and/or necrosis²⁰. Furthermore, nutritional status alters the rates of angiogenesis, extracellular matrix remodeling, and relative distribution of the immune cell population in adipose tissue²¹. The cells collectively work as endocrine organs, secreting a variety of signaling chemicals that influence feeding behavior, energy expenditure, metabolism, reproduction, and endocrine and immunological functions²². As the size and number of fat cells are associated with insulin sensitivity, glucose uptake, and fatty acid uptake, alterations in the function and cellular makeup of fat tissue can alter the metabolic state and result in medical complications^{23,24}.

Adipose tissue dysfunction is referred to as a state of hypersecretion of adipocytokines, which are pro-inflammatory, pro-atherogenic, and pro-diabetic, along with reduced anti-inflammatory and adiponectin secretion. An imbalance between pro- and anti-inflammatory adipokines can lead to insulin resistance, low-grade inflammation, and hypertension. Both lean and obese

women with PCOS have abnormal adipose tissue shape, which is considered a major characteristic of adipose tissue dysfunction^{14,25-27}. Adipocytes have the ability to store lipids, perceive regional and systemic cues, and respond by controlling the amount of energy mobilized by secreting paracrine and endocrine hormones. In addition to their function in glucose and lipid metabolism, adipocytes release endocrine factors that control glucose homeostasis, insulin sensitivity, inflammation, and tissue repair²⁸. Adipocytes release exosomal microRNAs, peptides (adipokines), and lipids (lipokines)²⁸, and dysregulated adipocytokine production leads to the development of common metabolic disorders^{29,30}. Accumulation of adipose tissue increases the production and secretion of Tumor Necrosis Factor α (TNF- α) and resistin, which can play an important role in the development of insulin resistance in obese patients^{29,30}. In obesity, adipocytes become enlarged to store excess energy. Adipocyte hypertrophy (an increase in adipocyte cell size) is caused by changes in adipocyte storage or lipolytic capacity. Under these conditions, adipocytes in adipose tissue are hypertrophic, insulin-resistant, unable to store triglycerides, and exhibit reduced energy expenditure. As a result, fatty acids are released into the bloodstream and build up in other organs, disrupting the metabolism which is a symptom of chronic metabolic illnesses like T2DM and CVD²⁸ [Figure 2].

Adipose tissue is characterized by hypertrophic adipocytes, a pro-inflammatory gene expression profile, dysregulated secretion of adipose-specific proteins, cytokines, and other proteins, and an increase in the macrophage and immune cell population in obese people^{31,32}. Similar alterations were observed in adipose tissues of women with PCOS. Additionally, hypertrophic adipocytes lead to decreased adiponectin³². In PCOS, the pro-inflammatory condition of obesity promotes insulin resistance and atherosclerosis, and adipose tissue expansion leads to hypoxia-related adipocyte death, which induces an influx of Mononuclear Cells (MNC) into the stromal-vascular compartment²¹. MNCs undergo morphological changes and develop into macrophages. The primary source of TNF α and interleukin 6 (IL-6) synthesis in adipose tissue is macrophages, which also promote cytokine production in adipocytes³³. Studies have demonstrated that IL-6 and TNF- α levels are elevated in PCOS^{34,35}. Adipocytes larger than 75 μm are associated with insulin resistance, increased insulin levels, and metabolic syndrome³⁶. Adipocyte diameter is usually increased by approximately 25% in women with PCOS compared to obese control women without PCOS^{26,37}. Hypertrophic adipocytes are insulin-resistant³¹. Insulin receptor phosphorylation in adipocytes is reduced in women with PCOS, resulting in decreased GLUT4

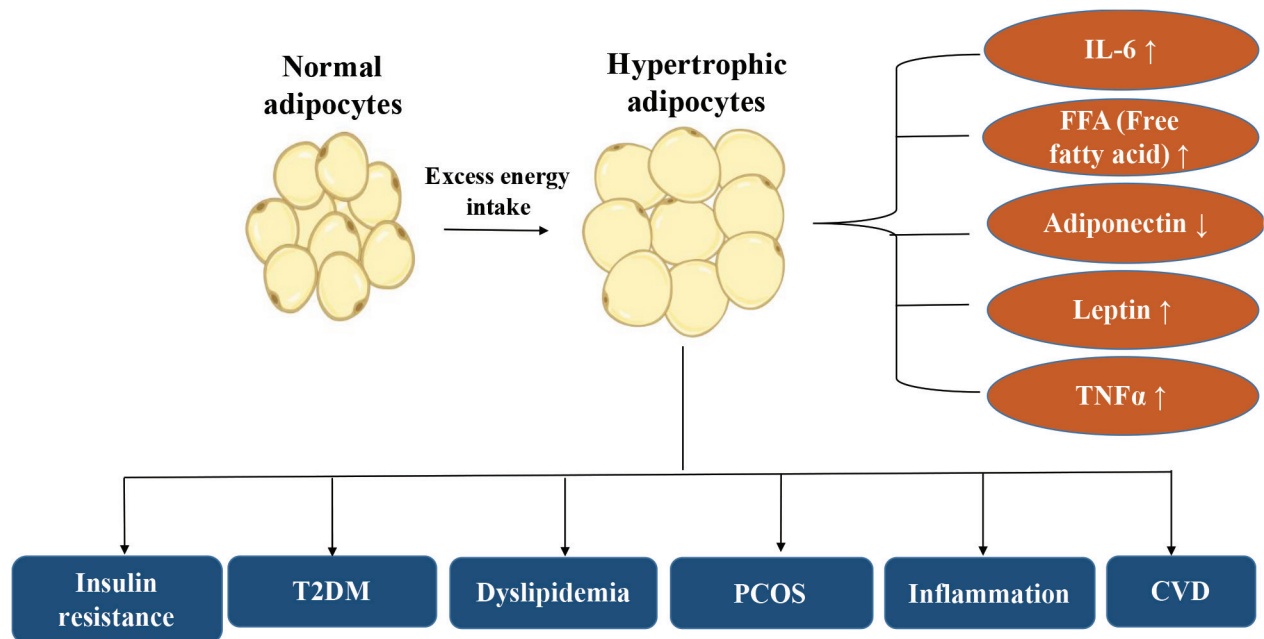


Figure 2. Due to excess energy intake, adipocytes become hypertrophic, insulin resistant, and unable to store triglycerides and fatty acids that are released into the bloodstream and build up in other organs, leading to ectopic lipid accumulation and disruption of metabolism, symptoms of chronic metabolic illnesses such as type 2 diabetes mellitus, PCOS, and CVDs.

translocation and altered insulin-dependent glucose uptake³⁸. A study conducted by Chang *et al.* revealed that women with PCOS have lower insulin-stimulated serine phosphorylation of Glycogen Synthase Kinase-3 β (GSK3 β)¹⁹. Adipocytes from women have increased activity and tyrosine phosphorylation of GSK3 β . These findings imply that GSK3 β is hyperactivated in PCOS³⁹. Subcutaneous abdominal preadipocytes cultivated *in vitro* from women with PCOS and controls were studied by Corbould and Dunaif⁴⁰. They found no differences in basal glycogen synthesis or glucose transport in preadipocytes in women with PCOS and controls, proving that, unlike skeletal muscle, the adipocyte lineage from PCOS women does not appear to have intrinsic abnormalities in insulin sensitivity⁴⁰. The correlation between hypertrophic adipocytes and insulin resistance *in vivo* is therefore probably caused by external factors such as elevated circulating testosterone levels⁴¹. In patients with diabetes or glucose intolerance, adipocyte size is increased compared to those with normal glucose tolerance^{41,42}.

Studies have shown that subcutaneous adipocytes from women with PCOS have lower levels of Hormone-Sensitive Lipase (HSL) expression and activity, as well as decreased catecholamine-mediated lipolysis. On the other hand, women with PCOS and hyperinsulinemia have increased catecholamine-induced lipolysis in adipocytes, raising blood-free fatty acids and causing dyslipidemia. As a result, the liver produces more free fatty acids, which increases the blood levels of Very Low-Density Lipoprotein (VLDL) and Triglycerides (TG)⁴³. Paradoxically, some studies have shown that subcutaneous adipocytes from PCOS women have lower levels of hormone-sensitive lipase expression and activity as well as decreased catecholamine-mediated lipolysis^{37,27}. A study conducted by EK *et al.* revealed marked lipolytic catecholamine resistance in lean PCOS women compared with healthy non-obese control women, which includes a 7-fold decrease in lipolytic sensitivity, and maximal lipolytic reaction to endogenous catecholamine noradrenaline stimulus was reduced by approximately 35%²⁷. The expression of CD36/Fatty acid translocase, a key membrane glycoprotein involved in the transportation of fatty acids, is upregulated in adipocytes of women with PCOS, and its expression correlates with insulin resistance^{44,45}. Adipose tissue produces pro-inflammatory cytokines and secretes angiotensin II, which increases NADPH

oxidase activity. NADPH oxidase is the main pathway for Reactive Oxygen Species (ROS) production in adipocytes⁴⁶.

Using microarray expression profiling analysis, Lee *et al.* studied the adipocytes of obese and non-obese subjects³¹. In contrast to non-obese individuals, 52 of 54 inflammatory/immune response genes were upregulated in the adipocytes of obese individuals³¹. Studies have also revealed that PCOS alters the gene expression profile of adipose tissues. Several genes are aberrantly expressed in the omental adipose tissue of women with PCOS⁴⁷. Variations in genes related to immunological response, cell development, metabolic syndrome, lipid metabolism, and insulin signaling have been discovered⁴⁷. Insulin resistance in PCOS may be caused by the dysregulation of factors secreted by hypertrophic adipocytes⁴¹.

Studies have reported that hyperandrogenism increases adipocyte size in the adipose tissue of women with PCOS, which can lead to adipose tissue dysfunction⁴⁸. These findings were supported by studies in rodents, which demonstrated that overexposure to androgens during the early postnatal and peri-puberty stages was linked to hypertrophic adipocytes in subcutaneous and visceral adipose tissue and insulin resistance⁴⁹⁻⁵¹. However, in female sheep and monkeys, prenatal exposure to testosterone has been linked to reduced adipocyte size^{52,53}. These contrasting results may be due to the species or nature of the androgens. Human pre-adipocytes have also shown androgen-driven suppression of adipocyte development, and anti-androgenic drugs were able to partially reverse this effect⁵⁴. However, some studies have revealed that the parameters most strongly linked to insulin resistance in women with PCOS are increased adipocyte size, reduced adiponectin levels, and increased waist circumference, but not androgen excess²⁶.

4. Adiponectin and PCOS

White adipocytes are important secretory cells that release various fatty acids, lipids, and proteins. Leptin and adiponectin are the most significant protein hormones produced and released by adipocytes⁵⁵. Adiponectin is a 224-amino acid protein with a molecular weight of 30 kDa that was first discovered in 1995⁵⁶. It is also known as adipocyte complement-related protein 30 kDa (ACRP30). Women were shown to have significantly higher adiponectin levels than men⁵⁷. Mice also show similar sexual dimorphism⁵⁸, wherein tissue adiponectin levels have been reported to rise

early in puberty, which can further increase post-castration in males or ovariectomy in females. Adiponectin expression is inhibited by estrogen *in vitro*. The human adiponectin gene is located on the chromosome 3q27. In a native French cohort, Froguel *et al.* identified a human chromosome 3q27 locus that is associated with diabetes susceptibility⁵⁹, and Comuzzie *et al.* showed that this locus is strongly associated with metabolic syndrome in individuals of European descent⁶⁰. Adiponectin is a key factor in regulating insulin sensitivity. Adiponectin promotes beta-oxidation of fatty acids by myocytes, decreases plasma fatty acid levels⁶¹, and inhibits hepatic glucose synthesis⁶². Adiponectin was positively correlated with High-Density Lipoprotein (HDL) and inversely correlated with Body Mass Index (BMI), percentage of body fat, waist-to-hip ratio, glucose, insulin, and triglycerides. Hypoadiponectinemia is closely associated with IR^{42,63} and high adiponectin levels are associated with low insulin resistance and a healthy lipid profile⁶³. Recombinant adiponectin treatment improves insulin resistance in obese mice with low circulating adiponectin levels⁶⁴ and prevents the onset of diet-induced insulin resistance⁶⁵. Circulating adiponectin levels are decreased in obesity⁵⁷, T2DM⁶⁶ CVD, and hypertension⁶⁷.

Increased adiponectin levels have been linked to improved glucose homeostasis, smaller adipocytes, and less visceral fat, all of which have protective functions against metabolic health²⁵. Furthermore, increased paternal adiponectin transcription has been shown to protect against adipose tissue dysfunction²⁵. In women with PCOS, lowered adiponectin concentrations are closely related to Heart Rate Recovery (HRR) blunt⁶⁸. Adiponectin has been reported to have anti-inflammatory, anti-atherogenic, and insulin-sensitizing activities⁴². Women with PCOS have significantly decreased adiponectin levels⁶⁹⁻⁷² linked to increased inflammation and higher adipokine levels⁷³, implying that dyslipidemia is directly associated with inflammation. Women with PCOS have considerably lower Adiponectin/Leptin (A/L) and higher HOMA/Adiponectin (H/A) ratios, respectively⁷⁴. A recent study demonstrated that the A/L ratio is markedly reduced in women with PCOS; however, there is no correlation with BMI or IR⁷⁵. Adiponectin expression is reduced by androgens by reducing their secretion⁷⁶. Although adiponectin does not appear to be directly involved in the pathophysiology of PCOS, it may interact with steroid production or its action in women with PCOS⁷⁷. Adiponectin levels may help identify high-risk for PCOS in women since low adiponectin levels can serve as a predictor of the development of T2DM and CVD in women.

In some studies, adiponectin secretion was shown to be independent of BMI or IR. As both lean and obese women with PCOS have abnormal adipose tissue morphology independent of BMI or IR, there seems to be another factor(s) influencing adipose tissue biology in PCOS. *In vitro* studies have demonstrated that differentiated pre-adipocytes and human adipocytes express LHCGR mRNA. Furthermore, fully differentiated adipocytes had a three-fold higher level of LHCGR mRNA than confluent or early differentiating preadipocytes. Thus, human preadipocytes are directly influenced by hCG *in vitro*, which affects their ability to proliferate and differentiate⁷⁸. As LH levels are higher in women with PCOS, there may be a possible link between higher LH levels and adipose tissue dysfunction in PCOS [Figure 3].

Additionally, higher LH levels, and a high LH:FSH ratio may contribute to the hyperandrogenism associated with adipose dysfunction in PCOS. Androgens influence adipocyte size and activity. Hyperandrogenism has been linked to lower adiponectin levels, leading to insulin resistance, as adiponectin is an insulin sensitizer⁶⁹. In women with PCOS, increased adipocyte size and decreased adiponectin levels are characteristics most closely linked to insulin resistance²⁶. Adiponectin overexpression is linked to reduced adipocyte size, increased mitochondrial density in adipocytes, and the transcriptional upregulation of factors involved in lipid storage^{79,80}.

5. Role of Other Adipocytokines in PCOS

Adipokines, also known as adipocytokines, are a class of bioactive proteins and immunological agents secreted by the adipose tissue. Adipokines play crucial roles in the regulation of glucose and lipid metabolism, energy homeostasis, insulin sensitivity, immunity, and inflammation. Adipokines are categorized as inflammatory or anti-inflammatory. The adipokines that are increased in obese individuals are classified as inflammatory adipokines. Numerous studies have demonstrated the role of adipokines in the pathophysiology of PCOS. Altered adipokine secretion leads to IR in obese individuals, and IR may be an important link between adipokines and polycystic ovaries. A study conducted by Mahde *et al.* revealed that IL-6, IL-18, TNF- α , RBP-4, Resistin, leptin, insulin, LH, testosterone, and free testosterone levels were elevated in women with PCOS⁸¹. Another study conducted on the Indian population by Ram *et*

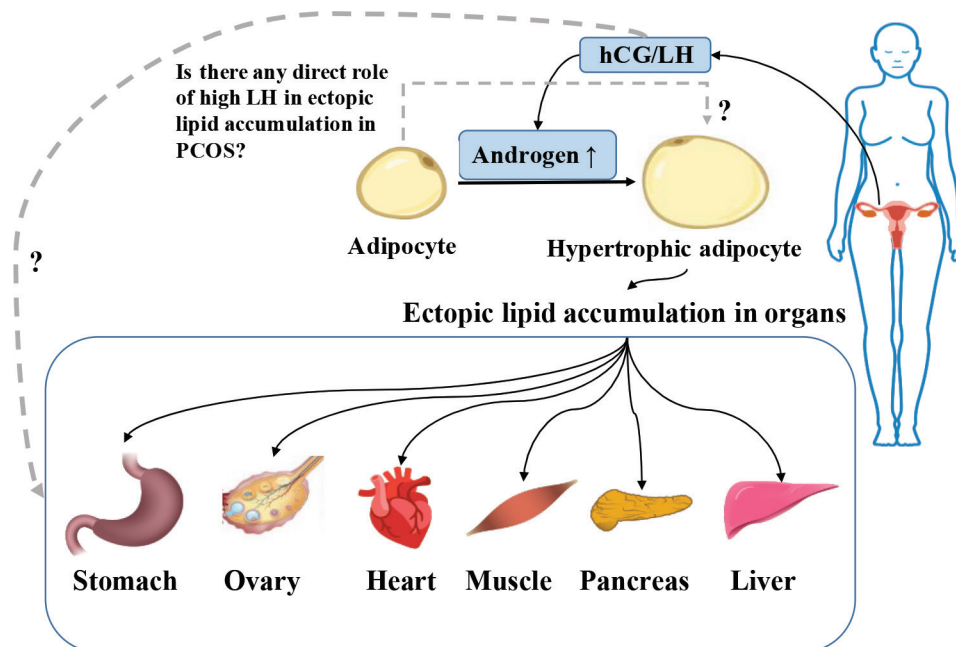


Figure 3. Hyperandrogenism is linked to adipose tissue dysfunction, leading to hypertrophic adipocytes and decreased adiponectin secretion, which leads to ectopic lipid accumulation. Here, we illustrated the possible direct role of hCG/LH in abnormal adipose tissue morphology and adipose tissue dysfunction in PCOS.

al. revealed that underweight, overweight, obese, and morbidly obese PCOS women had elevated serum leptin and serum TNF- α , IL-6, and IL-8 levels compared to women with normal weight and underweight non-PCOS. Leptin plays a key role in regulating female reproduction, and leptin resistance in underweight and obese women with PCOS may be caused by impaired leptin expression in ovaries. Elevated TNF- α , IL-6, and IL-8 levels are strongly correlated with Insulin resistance⁸². Studies have demonstrated that TNF- α induces IR in animal models and cultured cells. Expression of TNF- α is also significantly elevated in the adipose tissue of obese human patients, which is strongly associated with hyperinsulinemia²⁹. Hence, overproduction of TNF- α by adipose tissue contributes to insulin resistance. A recent study shows that both normal-weight and obese PCOS women have lower plasma omentin-1 levels than non-PCOS women⁸³. Omentin-1 is a novel adipokine produced by the visceral white adipose tissue. It has anti-inflammatory, anti-obesity, and antidiabetic effects⁸⁴. Apelin is an adipokine that contributes to the endocrine regulation of glucose metabolism. There are conflicting reports on serum apelin levels in women with PCOS. Apelin might be a contributing factor in the development of PCOS⁸⁵. Chemerin is an adipokine that induces insulin resistance and modulates adipogenesis and adipocyte

metabolism. Women with PCOS have elevated chemerin levels compared with controls⁸⁶. Retinol-Binding Protein 4 (RBP4) is secreted by adipocytes, hepatocytes, and macrophages. A study revealed that women with PCOS in Asia have elevated RBP4 levels compared with controls⁸⁷. Visfatin is primarily expressed in human visceral fat, bone marrow, the liver, and muscles. There are conflicting studies on visfatin levels and expression in women with PCOS. Most studies have revealed higher visfatin levels in the ovary⁸⁸. Visceral adipose tissue-derived serpin (Vaspin) is a member of the serine protease inhibitor family and is secreted mainly by visceral and subcutaneous adipose tissues. Vaspin expression is strongly associated with lipid metabolism and IR. Numerous studies have demonstrated that women with PCOS have higher vaspin levels which contributes to IR⁸⁸. Lipocalin 2 is a novel adipokine majorly expressed in white adipose tissue. There are conflicting studies on lipocalin 2 levels in women with PCOS. Some studies have demonstrated that women have higher lipocalin 2 levels, some suggest that lipocalin 2 levels are lower in women with PCOS than in controls, while others have shown no significant difference between women with PCOS and controls⁸⁸.

Several studies have revealed a strong association between adipose tissue dysfunction, altered adipokine secretion, and PCOS in the Indian population. A study

conducted by Nambiar *et al.* in a south Indian population demonstrated that women with PCOS have lower adiponectin levels and higher resistin levels than controls. They also suggested that serum adiponectin levels might be a potential indicator of abdominal fat in PCOS⁸⁹. IL-6 regulates ovarian steroid synthesis, follicular maturation, fertilization, and implantation. The human IL-6 gene is situated at the 7p21-24 locus and has an upstream promoter of 303 bp. The IL-6 promoter's transcription rate is influenced by frequent G/C Single Nucleotide Polymorphisms (SNP) at np -174. A study conducted by Tumu *et al.*, for the first time, revealed a strong association between IL-6-174G/C SNP and PCOS in South Indian women. Compared to controls, there was a substantial increase in the G/G genotype frequency, indicating that the "G" allele may be regarded as a risk factor for PCOS. Thus, the IL-6 gene can be considered a potential candidate for PCOS treatment⁹⁰. As mentioned earlier, Ram *et al.* concluded that leptin, TNF- α , IL-6, and IL-8 levels are elevated in South Indian PCOS subjects compared with controls⁸².

6. Novel Therapies Targeting Adipocytokines for PCOS and Related Disorders

New drugs with therapeutic potential have been developed to target adipokines for PCOS treatment. Recombinant proteins, peptides, and therapeutic antibodies have been used in adipokine-targeting protein therapies. With encouraging results in murine models, anti-TNF- α therapy has been proposed as a potential treatment for PCOS⁹¹. Osmotin is an antifungal stress-responsive protein that is similar to adiponectin in terms of structure and functionality. Osmotin was identified as a mammalian adiponectin homolog by a variety of *in vitro* and *in vivo* studies. In ob/ob and db/db mice, osmotin prevents Non-Alcoholic Fatty Liver Disease (NAFLD)⁹². Apelin agonists and antagonists both have a positive impact, and further research is needed to fully understand the potential of recently developed therapeutic biologics that target apelin⁹². The anti-ChemR23 nanobodies CA4910 and CA5183, have been developed as new functional antibodies that target the chemerin receptor. CA4910 and CA5183 may represent novel strategies for investigating the involvement of the chemerin/ChemR23 system in the emergence of PCOS

and related disorders⁹². Although only a few monoclonal antibody medications that target adipokines have been investigated in PCOS preclinical models, they have already demonstrated tentative therapeutic results in a number of diseases⁹². Adipokine receptor agonists are mostly used to treat diabetes-associated conditions. PCOS is often associated with diabetes. Therefore, these agonists may help with PCOS symptoms. An oral drug called AdipoRon stimulates AdipoR1 and AdipoR2 expression in skeletal muscle and the liver, and the effects of AdipoRon are highly similar to those of adiponectin. AdipoRon treatment reduced IR and glucose intolerance in wild-type mice fed a high-fat diet. Additionally, in mice treated with corticosterone, AdipoRon reverses abdominal adiposity, hyperleptinemia, and hyperinsulinemia⁹². Adipo anti-inflammation agonist (AdipoAI) is a newly discovered and powerful AdipoR agonist. AdipoAI is structurally similar to AdipoRon and has potent anti-inflammatory effects in animal models of diet-induced obesity and Lipopolysaccharide (LPS)-induced septic shock⁹². Novel drugs that target adipokines may open new therapeutic possibilities for PCOS.

7. Summary

Adipose tissue dysfunction is a crucial factor in PCOS, and both lean and obese women with PCOS have abnormal adipose tissue morphology. In PCOS, adipokine, cytokine, and chemoattractant protein secretion are enhanced in a pro-inflammatory manner, resulting in lower insulin sensitivity via altered glucose transporters, and thus decreased glucose absorption. Considering the intricate metabolic environment associated with PCOS, including hyperandrogenism, hyperinsulinemia, central adiposity, and chronic low-grade inflammation, it is challenging to identify the factors (s) that cause adipose tissue dysfunction. Adiponectin and adipocyte size are the strongest predictors of insulin resistance in women with PCOS. Understanding whether high LH levels cause metabolic defects and adipose tissue dysfunction in PCOS would help diagnose or treat the metabolic complications associated with this reproductive disease.

8. Abbreviations

ACRP30, Adipocyte Complement-Related Protein 30 kDa; BMI, Body mass index; CD36, Cluster of differentiation 36/ Fatty acid translocase; CVD, Cardiovascular diseases;

FF, Follicular fluid; GLUT 4, Glucose transporter type 4; GSK3 β , Glycogen synthase kinase-3 β ; HDL, High-density lipoprotein; HRR, Heart rate recovery; HSL, Hormone-sensitive lipase; IL-6, Interleukin 6; IGT, Impaired glucose tolerance; IR, Insulin resistance; LH, Luteinizing hormone; LPL, Lipoprotein lipase; MNC, Mononuclear cell; NADPH, Nicotinamide adenine dinucleotide phosphate; NAFLD, Nonalcoholic fatty liver disease; ROS, Reactive oxygen species; SHBG, Sex hormone binding globulin; SNP, Single nucleotide polymorphism; T2DM; Type 2 diabetes mellitus; TG, Triglycerides; TNF α , Tumor necrosis factor α ; VLDL, Very-low-density lipoprotein.

9. References

1. Kaur S, Ahamad I, Gouri Devi M, Singh R. Effect of body mass index on the biochemical outcome after fresh embryo transfer in women with and without Polycystic Ovary Syndrome (PCOS). *J Pharmaceutical Negative Results (JPNR)*. 2021; 12(1):74-9.
2. Kaur S, Archer KJ, Devi MG, Kriplani A, Strauss JF 3rd, Singh R. Differential gene expression in granulosa cells from Polycystic Ovary Syndrome patients with and without insulin resistance: identification of susceptibility gene sets through network analysis. *JCEM*. 2012; 97:E2016-21. <https://doi.org/10.1210/jc.2011-3441>
3. Rotterdam ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to Polycystic Ovary Syndrome. *Fertil Steril*. 2004; 81(1):19-25. <https://doi.org/10.1016/j.fertnstert.2003.10.004>
4. Adashi EY, Cibula D, Peterson M, Azziz R. The polycystic ovary syndrome: the first 150 years of study. *F S Rep*. 2022; 4(1):2-18. <https://doi.org/10.1016/j.xfre.2022.12.002>
5. Singh R, Kaur S, Yadav S, Bhatia S. Gonadotropins as pharmacological agents in assisted reproductive technology and Polycystic Ovary Syndrome. *Trends Endocrinol Metab*. 2023; 34(4):194-215. <https://doi.org/10.1016/j.tem.2023.02.002>
6. Blank SK, McCartney CR, Helm KD, Marshall JC. Neuroendocrine effects of androgens in adult Polycystic Ovary Syndrome and female puberty. *Semin Reprod Med*. 2007; 25(5):352-9. <https://doi.org/10.1055/s-2007-984741>
7. Dumesic DA, Lobo RA. Cancer risk and PCOS. *Steroids*. 2013; 78(8):782-5. <https://doi.org/10.1016/j.steroids.2013.04.004>
8. Bedaiwy MA, Abdel-Rahman MY, Tan J, *et al*. Clinical, hormonal, and metabolic parameters in women with subclinical hypothyroidism and Polycystic Ovary Syndrome: A cross-sectional study. *J Womens Health (Larchmt)*. 2018; 27(5):659-64. <https://doi.org/10.1089/jwh.2017.6584>
9. Escobar-Morreale HF, San Millán JL. Abdominal adiposity and the Polycystic Ovary Syndrome. *Trends Endocrinol Metab*. 2007; 18(7):266-72. <https://doi.org/10.1016/j.tem.2007.07.003>
10. Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the Polycystic Ovary Syndrome revisited: an update on mechanisms and implications. *Endocr Rev*. 2012; 33(6):981-1030. <https://doi.org/10.1210/er.2011-1034>
11. Fedeli V, Catizone A, Querqui A, Unfer V, Bizzarri M. The Role of Inositols in the hyperandrogenic phenotypes of PCOS: A re-reading of Lerner's results. *Int J Mol Sci*. 2023; 24(7):6296. <https://doi.org/10.3390/ijms24076296>
12. Urbanek M. The genetics of the Polycystic Ovary Syndrome. *Nat Clin Pract Endocrinol Metab*. 2007; 3(2):103-11. <https://doi.org/10.1038/ncpendmet0400>
13. Fahs D, Salloum D, Nasrallah M, Ghazeeri G. Polycystic Ovary Syndrome: pathophysiology and controversies in diagnosis. *Diagnostics (Basel)*. 2023; 13(9):1559. <https://doi.org/10.3390/diagnostics13091559>
14. Mannerås-Holm L, Leonhardt H, Kullberg J, *et al*. Adipose tissue has aberrant morphology and function in PCOS: enlarged adipocytes and low serum adiponectin, but not circulating sex steroids, are strongly associated with insulin resistance. *J Clin Endocrinol Metab*. 2011; 96(2):E304-11. <https://doi.org/10.1210/jc.2010-1290>
15. Omabe M, Elom S, Omabe KN. Emerging metabolomics biomarkers of Polycystic Ovarian Syndrome; targeting the master metabolic disrupters for diagnosis and treatment. *Endocr Metab Immune Disord Drug Targets*. 2018; 18(3):221-9. <https://doi.org/10.2174/1871530318666180122165415>
16. Legro RS, Kunselman AR, Dunaif A. Prevalence and predictors of dyslipidemia in women with Polycystic Ovary Syndrome. *Am J Med*. 2001; 111(8):607-13. [https://doi.org/10.1016/S0002-9343\(01\)00948-2](https://doi.org/10.1016/S0002-9343(01)00948-2)
17. Yilmaz M, Biri A, Bukan N, *et al*. Levels of lipoprotein and homocysteine in non-obese and obese patients with Polycystic Ovary Syndrome. *Gynecol Endocrinol*. 2005; 20(5):258-63. <https://doi.org/10.1080/09513590400027265>
18. Chen MJ, Ho HN. Hepatic manifestations of women with Polycystic Ovary Syndrome. *Best Pract Res Clin Obstet Gynaecol*. 2016; 37:119-28. <https://doi.org/10.1016/j.bpobgyn.2016.03.003>
19. Chang E, Varghese M, Singer K. Gender and sex differences in adipose tissue. *Curr Diab Rep*. 2018; 18(9):69. <https://doi.org/10.1007/s11892-018-1031-3>
20. Hausman DB, DiGirolamo M, Bartness TJ, Hausman GJ, Martin RJ. The biology of white adipocyte proliferation. *Obes Rev*. 2001; 2(4):239-54. <https://doi.org/10.1046/j.1467-789X.2001.00042.x>
21. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with

- macrophage accumulation in adipose tissue. *J Clin Invest.* 2003; 112(12):1796-808. <https://doi.org/10.1172/JCI200319246>
22. Kershaw EE, Flier JS. Adipose tissue is an endocrine organ. *J Clin Endocrinol Metab.* 2004; 89(6):2548-56. <https://doi.org/10.1210/jc.2004-0395>
 23. Virtue S, Vidal-Puig A. Adipose tissue expandability, lipotoxicity and the metabolic syndrome--an allostatic perspective. *Biochim Biophys Acta.* 2010; 1801(3):338-49. <https://doi.org/10.1016/j.bbali.2009.12.006>
 24. Rutkowski JM, Stern JH, Scherer PE. The cell biology of fat expansion. *J Cell Biol.* 2015; 208(5):501-12. <https://doi.org/10.1083/jcb.201409063>
 25. Benrick A, Chanclón B, *et al.* Adiponectin protects against the development of metabolic disturbances in a PCOS mouse model. *Proc Natl Acad Sci USA.* 2017; 114(34):E7187-96. <https://doi.org/10.1073/pnas.1708854114>
 26. Brennan KM, Kroener LL, *et al.* Polycystic Ovary Syndrome: Impact of lipotoxicity on metabolic and reproductive health. *Obstet Gynecol Surv.* 2019; 74(4):223-31. <https://doi.org/10.1097/OGX.0000000000000661>
 27. Dumesic DA, Phan JD, *et al.* Adipose insulin resistance in normal-weight women with Polycystic Ovary Syndrome. *J Clin Endocrinol Metab.* 2019; 104(6):2171-83. <https://doi.org/10.1210/jc.2018-02086>
 28. Scheja L, Heeren J. The endocrine function of adipose tissues in health and cardiometabolic disease. *Nat Rev Endocrinol.* 2019; 15(9):507-24. <https://doi.org/10.1038/s41574-019-0230-6>
 29. Würfel M, Blüher M, Stumvoll M, *et al.* Adipokines as clinically relevant therapeutic targets in obesity. *Biomedicines.* 2023; 11(5):1427. <https://doi.org/10.3390/biomedicines11051427>
 30. Steppan CM, Bailey ST, Bhat S, *et al.* The hormone resistin links obesity to diabetes. *Nature.* 2001; 409(6818):307-12. <https://doi.org/10.1038/35053000>
 31. Lee YH, Pratley RE. The evolving role of inflammation in obesity and the metabolic syndrome. *Curr Diab Rep.* 2005; 5(1):70-5. <https://doi.org/10.1007/s11892-005-0071-7>
 32. Haczeyni F, Bell-Anderson KS, Farrell GC. Causes and mechanisms of adipocyte enlargement and adipose expansion. *Obes Rev.* 2018; 19(3):406-20. <https://doi.org/10.1111/obr.12646>
 33. Fain JN, Bahouth SW, Madan AK. TNF α release by the nonfat cells of human adipose tissue. *Int J Obes Relat Metab Disord.* 2004; 28(4):616-22. <https://doi.org/10.1038/sj.ijo.0802594>
 34. Sayin NC, Gücer F, Balkanlı-Kaplan P, *et al.* Elevated serum TNF- α levels in normal-weight women with polycystic ovaries or Polycystic Ovary Syndrome. *J Reprod Med.* 2003; 48(3):165-70.
 35. Amato G, Conte M, Mazziotti G, *et al.* Serum and follicular fluid cytokines in Polycystic Ovary Syndrome during stimulated cycles. *Obstet Gynecol.* 2003; 101(6):1177-82. <https://doi.org/10.1097/00006250-200306000-00009>
 36. Arner E, Westermark PO, Spalding KL, *et al.* Adipocyte turnover: relevance to human adipose tissue morphology. *Diabetes.* 2010; 59(1):105-9. <https://doi.org/10.2337/db09-0942>
 37. Faulds G, Rydén M, Ek I, Wahrenberg H, Arner P. Mechanisms behind lipolytic catecholamine resistance of subcutaneous fat cells in the Polycystic Ovarian Syndrome. *J Clin Endocrinol Metab.* 2003; 88(5):2269-73. <https://doi.org/10.1210/jc.2002-021573>
 38. Herman R, Kravos NA, Jensterle M, Janež A, Dolžan V. Metformin and insulin resistance: A review of the underlying mechanisms behind changes in GLUT4-mediated glucose transport. *Int J Mol Sci.* 2022; 23(3):1264. <https://doi.org/10.3390/ijms23031264>
 39. Chang W, Goodarzi MO, Williams H, Magoffin DA, Pall M, Azziz R. Adipocytes from women with Polycystic Ovary Syndrome demonstrate altered phosphorylation and activity of glycogen synthase kinase 3. *Fertil Steril.* 2008; 90(6):2291-7. <https://doi.org/10.1016/j.fertnstert.2007.10.025>
 40. Corbould A, Dunaif A. The adipose cell lineage is not intrinsically insulin-resistant in Polycystic Ovary Syndrome. *Metabolism.* 2007; 56(5):716-22. <https://doi.org/10.1016/j.metabol.2006.12.021>
 41. Villa J, Pratley RE. Adipose tissue dysfunction in Polycystic Ovary Syndrome. *Curr Diab Rep.* 2011; 11(3):179-84. <https://doi.org/10.1007/s11892-011-0189-8>
 42. Weyer C, Foley JE, Bogardus C, Tataranni PA, Pratley RE. Enlarged subcutaneous abdominal adipocyte size, but not obesity itself, predicts type II diabetes independent of insulin resistance. *Diabetologia.* 2000; 43(12):1498-506. <https://doi.org/10.1007/s001250051560>
 43. Diamanti-Kandaraki E, Papavassiliou AG, Kandaraki SA, Chrousos GP. Pathophysiology and types of dyslipidemia in PCOS. *Trends Endocrinol Metab.* 2007; 18(7):280-5. <https://doi.org/10.1016/j.tem.2007.07.004>
 44. Seow KM, Tsai YL, Hwang JL, Hsu WY, Ho LT, Juan CC. Omental adipose tissue overexpression of fatty acid transporter CD36 and decreased expression of hormone-sensitive lipase in insulin-resistant women with Polycystic Ovary Syndrome. *Hum Reprod.* 2009; 24(8):1982-8. <https://doi.org/10.1093/humrep/dep122>
 45. Glinborg D, Højlund K, Andersen M, Henriksen JE, Beck-Nielsen H, Handberg A. Soluble CD36 and risk markers of insulin resistance and atherosclerosis are elevated in Polycystic Ovary Syndrome and significantly reduced during pioglitazone treatment. *Diabetes Care.* 2008; 31(2):328-34. <https://doi.org/10.2337/dc07-1424>
 46. Fernández-Sánchez A, Madrigal-Santillán E, Bautista M, *et al.* Inflammation, oxidative stress, and obesity. *Int J Mol Sci.* 2011; 12(5):3117-32. <https://doi.org/10.3390/ijms12053117>

47. Cortón M, Botella-Carretero JJ, Benguría A, *et al.* Differential gene expression profile in omental adipose tissue in women with Polycystic Ovary Syndrome. *J Clin Endocrinol Metab.* 2007; 92(1):328-37. <https://doi.org/10.1210/jc.2006-1665>
48. Echiburú B, Pérez-Bravo F, Galgani JE, *et al.* Enlarged adipocytes in subcutaneous adipose tissue associated with hyperandrogenism and visceral adipose tissue volume in women with Polycystic Ovary Syndrome. *Steroids.* 2018; 130:15-21. <https://doi.org/10.1016/j.steroids.2017.12.009>
49. Nohara K, Liu S, Meyers MS, *et al.* Developmental androgen excess disrupts reproduction and energy homeostasis in adult male mice. *J Endocrinol.* 2013; 219(3):259-68. <https://doi.org/10.1530/JOE-13-0230>
50. Mannerås L, Cajander S, Holmäng A, *et al.* A new rat model exhibiting both ovarian and metabolic characteristics of Polycystic Ovary Syndrome. *Endocrinology.* 2007; 148(8):3781-91. <https://doi.org/10.1210/en.2007-0168>
51. Perello M, Castrogiovanni D, Giovambattista A, Gaillard RC, Spinedi E. Impairment in insulin sensitivity after early androgenization in the post-pubertal female rat. *Life Sci.* 2007; 80(19):1792-8. <https://doi.org/10.1016/j.lfs.2007.02.013>
52. Puttabyatappa M, Lu C, Martin JD, Chazenbalk G, Dumesic D, Padmanabhan V. Developmental programming: impact of prenatal testosterone excess on steroidal machinery and cell differentiation markers in visceral adipocytes of female sheep. *Reprod Sci.* 2018; 25(7):1010-23. <https://doi.org/10.1177/1933719117746767>
53. Keller E, Chazenbalk GD, Aguilera P, *et al.* Impaired preadipocyte differentiation into adipocytes in subcutaneous abdominal adipose of PCOS-like female rhesus monkeys. *Endocrinology.* 2014; 155(7):2696-703. <https://doi.org/10.1210/en.2014-1050>
54. Gupta V, Bhasin S, Guo W, *et al.* Effects of dihydrotestosterone on differentiation and proliferation of human mesenchymal stem cells and preadipocytes. *Mol Cell Endocrinol.* 2008; 296(1-2):32-40. <https://doi.org/10.1016/j.mce.2008.08.019>
55. Frayn KN, Karpe F, Fielding BA, Macdonald IA, Coppack SW. Integrative physiology of human adipose tissue. *Int J Obes Relat Metab Disord.* 2003; 27(8):875-88. <https://doi.org/10.1038/sj.ijo.0802326>
56. Luo L, Liu M. Adiponectin: friend or foe in obesity and inflammation. *Med Rev (Berl).* 2022; 2(4):349-62. <https://doi.org/10.1515/mr-2022-0002>
57. Atzmon G, Pollin TI, Crandall J, *et al.* Adiponectin levels and genotype: a potential regulator of life span in humans. *J Gerontol A Biol Sci Med Sci.* 2008; 63(5):447-53. <https://doi.org/10.1093/gerona/63.5.447>
58. Combs TP, Berg AH, Rajala MW, *et al.* Sexual differentiation, pregnancy, calorie restriction, and aging affect the adipocyte-specific secretory protein adiponectin. *Diabetes.* 2003; 52(2):268-76. <https://doi.org/10.2337/diabetes.52.2.268>
59. Vionnet N, Hani EH, Dupont S, *et al.* Genomewide search for type 2 diabetes-susceptibility genes in French whites: evidence for a novel susceptibility locus for early-onset diabetes on chromosome 3q27-qter and independent replication of a type 2-diabetes locus on chromosome 1q21-q24. *Am J Hum Genet.* 2000; 67(6):1470-80. <https://doi.org/10.1086/316887>
60. Kissebah AH, Sonnenberg GE, Myklebust J, *et al.* Quantitative trait loci on chromosomes 3 and 17 influence phenotypes of the metabolic syndrome. *Proc Natl Acad Sci U S A.* 2000; 97(26):14478-14483. <https://doi.org/10.1073/pnas.97.26.14478>
61. Fruebis J, Tsao TS, Javorschi S, *et al.* A proteolytic cleavage product of 30-kDa adipocyte complement-related protein increases fatty acid oxidation in muscle and causes weight loss in mice. *Proc Natl Acad Sci USA.* 2001; 98(4):2005-10. <https://doi.org/10.1073/pnas.98.4.2005>
62. Combs TP, Berg AH, Obici S, Scherer PE, Rossetti L. Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *J Clin Invest.* 2001; 108(12):1875-81. <https://doi.org/10.1172/JCI14120>
63. Yamamoto Y, Hirose H, Saito I, *et al.* Correlation of the adipocyte-derived protein adiponectin with insulin resistance index and serum high-density lipoprotein-cholesterol, independent of body mass index, in the Japanese population. *Clin Sci (Lond).* 2002; 103(2):137-42. <https://doi.org/10.1042/cs1030137>
64. Kubota N, Terauchi Y, Yamauchi T, *et al.* Disruption of adiponectin causes insulin resistance and neointimal formation. *J Biol Chem.* 2002; 277(29):25863-6. <https://doi.org/10.1074/jbc.C200251200>
65. Yamauchi T, Kamon J, Waki H, *et al.* The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nat Med.* 2001; 7(8):941-6. <https://doi.org/10.1038/90984>
66. Hotta K, Funahashi T, Arita Y, *et al.* Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol.* 2000; 20(6):1595-9. <https://doi.org/10.1161/01.ATV.20.6.1595>
67. Matsubara M, Maruoka S, Katayose S. Decreased plasma adiponectin concentrations in women with dyslipidemia. *J Clin Endocrinol Metab.* 2002; 87(6):2764-9. <https://doi.org/10.1210/jcem.87.6.8550>
68. Sun W, Liu G, Liu B. Association between circulating adiponectin and heart rate recovery in women with Polycystic Ovarian Syndrome. *Endocr Res.* 2022; 47(2):56-63. <https://doi.org/10.1080/07435800.2021.2011908>
69. Spritzer PM, Lecke SB, Satler F, Morsch DM. Adipose tissue dysfunction, adipokines, and low-grade chronic inflammation in Polycystic Ovary Syndrome. *Reproduction.* 2015; 149(5):R219-27. <https://doi.org/10.1530/REP-14-0435>

70. Toulis KA, Goulis DG, Farmakiotis D, *et al.* Adiponectin levels in women with Polycystic Ovary Syndrome: a systematic review and a meta-analysis. *Hum Reprod Update.* 2009; 15(3):297-307. <https://doi.org/10.1093/humupd/dmp006>
71. Zhang N, Shi YH, Hao CF, *et al.* Association of +45G15G(T/G) and +276(G/T) polymorphisms in the ADIPOQ gene with Polycystic Ovary Syndrome among Han Chinese women. *Eur J Endocrinol.* 2008; 158(2):255-60. <https://doi.org/10.1530/EJE-07-0576>
72. Glintborg D, Frystyk J, Højlund K, *et al.* Total and High Molecular Weight (HMW) adiponectin levels and measures of glucose and lipid metabolism following pioglitazone treatment in a randomized placebo-controlled study in Polycystic Ovary Syndrome. *Clin Endocrinol (Oxf).* 2008; 68(2):165-74. <https://doi.org/10.1111/j.1365-2265.2007.03015.x>
73. Glintborg D. Endocrine and metabolic characteristics in Polycystic Ovary Syndrome. *Dan Med J.* 2016; 63(4):B5232.
74. Vatannejad A, Kheirollahi A. Adiponectin/leptin and HOMA/adiponectin ratios in Iranian women with Polycystic Ovary Syndrome. *Ir J Med Sci.* 2023; 192(4):1793-9. <https://doi.org/10.1007/s11845-023-03408-4>
75. Mishra P, Mittal P, Rani A, Bharti R, Agarwal V, Suri J. Adiponectin to leptin ratio and its association with insulin resistance in women with Polycystic Ovarian Syndrome. *Indian J Endocrinol Metab.* 2022; 26(3):239-44. https://doi.org/10.4103/ijem.ijem_137_22
76. Nishizawa H, Shimomura I, Kishida K, *et al.* Androgens decrease plasma adiponectin, an insulin-sensitizing adipocyte-derived protein. *Diabetes.* 2002; 51(9):2734-41. <https://doi.org/10.2337/diabetes.51.9.2734>
77. Panidis D, Kourtis A, Farmakiotis D, Mouslech T, Rousso D, Koliakos G. Serum adiponectin levels in women with polycystic ovary syndrome. *Hum Reprod.* 2003; 18(9):1790-6. <https://doi.org/10.1093/humrep/deg353>
78. Dos Santos E, Dieudonné MN, Leneuve MC, Pecquery R, Serazin V, Giudicelli Y. *In vitro* effects of chorionic gonadotropin hormone on human adipose development. *J Endocrinol.* 2007; 194(2):313-25. <https://doi.org/10.1677/JOE-06-0101>
79. Mannerås-Holm L, Benrick A, Stener-Victorin E. Gene expression in subcutaneous adipose tissue differs in women with Polycystic Ovary Syndrome and controls matched pair-wise for age, body weight, and body mass index. *Adipocyte.* 2014; 3(3):190-6. <https://doi.org/10.4161/adip.28731>
80. Asterholm IW, Scherer PE. Enhanced metabolic flexibility is associated with elevated adiponectin levels. *Am J Pathol.* 2010; 176(3):1364-76. <https://doi.org/10.2353/ajpath.2010.090647>
81. Mahde A, Shaker M, Al-Mashhadani Z. Study of Omentin1 and other adipokines and hormones in PCOS patients. *Oman Med J.* 2009; 24(2):108-18.
82. Ravishankar Ram M, Sundararaman PG, Mahadevan S, Malathi R. Cytokines and leptin correlation in patients with Polycystic Ovary Syndrome: Biochemical evaluation in south Indian population. *Reprod Med Biol.* 2005; 4(4):247-54. <https://doi.org/10.1111/j.1447-0578.2005.00114.x>
83. Franik G, Sadlocha M, Madej P, *et al.* Circulating omentin-1 levels and inflammation in Polycystic Ovary Syndrome. *Ginekol Pol.* 2020; 91(6):308-12. <https://doi.org/10.5603/GP.2020.0057>
84. Lee MW, Lee M, Oh KJ. Adipose tissue-derived signatures for obesity and type 2 diabetes: adipokines, cytokines, and MicroRNAs. *J Clin Med.* 2019; 8(6):854. <https://doi.org/10.3390/jcm8060854>
85. Dravecká I, Figurová J, Lazúrová I. Is Apelin a new biomarker in patients with Polycystic Ovary Syndrome? *Physiol Res.* 2021; 70(4):S635-41. <https://doi.org/10.33549/physiolres.934708>
86. Gao Y, Xin C, Fan H, Sun X, Wang H. Circulating apelin and chemerin levels in patients with Polycystic Ovary Syndrome: A meta-analysis. *Front Endocrinol (Lausanne).* 2023; 13:1076951. <https://doi.org/10.3389/fendo.2022.1076951>
87. Jia J, Bai J, Liu Y, *et al.* Association between retinol-binding protein 4 and Polycystic Ovary Syndrome: A meta-analysis. *Endocr J.* 2014; 61(10):995-1002. <https://doi.org/10.1507/endocrj.EJ14-0186>
88. Chen P, Jia R, Liu Y, Cao M, Zhou L, Zhao Z. Progress of adipokines in the female reproductive system: A focus on Polycystic Ovary Syndrome. *Front Endocrinol (Lausanne).* 2022; 13:881684. <https://doi.org/10.3389/fendo.2022.881684>
89. Nambiar V, Vijesh VV, Lakshmanan P, Sukumaran S, Suganthi R. Association of adiponectin and resistin gene polymorphisms in South Indian women with Polycystic Ovary Syndrome. *Eur J Obstet Gynecol Reprod Biol.* 2016; 200:82-8. <https://doi.org/10.1016/j.ejogrb.2016.02.031>
90. Tumu VR, Govatati S, Guruvaiah P, Deenadayal M, Shivaji S, Bhanoori M. An interleukin-6 gene promoter polymorphism is associated with Polycystic Ovary Syndrome in South Indian women. *J Assist Reprod Genet.* 2013; 30(12):1541-6. <https://doi.org/10.1007/s10815-013-0111-1>
91. Bril F, Ezeh U, Amiri M, *et al.* Adipose tissue dysfunction in Polycystic Ovary Syndrome. *J Clin Endocrinol Metab.* 2023; dgad356. <https://doi.org/10.1210/clinem/dgad356>
92. Xu Y, Zhu H, Li W, *et al.* Targeting adipokines in Polycystic Ovary Syndrome and related metabolic disorders: from experimental insights to clinical studies. *Pharmacol Ther.* 2022; 240:108284. <https://doi.org/10.1016/j.pharmthera.2022.108284>