



Review Article

Paracoccus marginatus, an invasive mealybug of papaya and its biological control – An overview

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ABSTRACT: The papaya mealybug, *Paracoccus marginatus* Williams and Granara de Willink (Hemiptera: Pseudococcidae) is native of Mexico and /or Central and North America. Since its description in 1992, it has invaded several Carbbean Islands, Florida, Sri Lanka, Indonesia, Taiwan, India, Bangladesh, West Africa, and it is likely to spread many other countries. It is highly polyphagus attacking more than 70 plant species causing severe loss. Insecticides failed to give adequate control of *P. marginatus*. On the other hand, natural enemies particularly, *Acerophagus papayae* Noyes and Schauff were highly useful to suppress the papaya mealybug. Its origin, distribution, taxonomy, biology, ecology, host range, natural enemies and methods of control particularly classical biological control are reviewed here.

KEY WORDS: *Paracoccus marginatus*, invasive mealybug, *Acerophagus papayae*, biology, distribution, host range, natural enemies, taxonomy, classical biological control.

(Article chronicle: Received: 30-05-2012 Revised: 16-06-2012 Accepted: 18-06-2012)

INTRODUCTION

Mealybugs throughout the world cause various economic problems. *Paracoccus marginatus* Williams and Granara de Willink, popularly known as papaya mealybug (PMB) has invaded several countries and damaged many economically important crop plants (Muniappan *et al.*, 2008; Shylesha *et al.*, 2011a). It is 'hard to kill pest' with conventional insecticides because of cryptic habit and waxy coating over the body. This review provides information on the biosystematics, origin, distribution, biology, host range, ecology, natural enemies, management options with particular emphasis on biological control which will be ultimately useful for the successful management of papaya mealybug.

ORIGIN AND DISTRIBUTION

Paracoccus marginatus is native to Mexico and/or Central and North America (Miller *et al.*, 1999; Watson and Chandler, 1999). Since its first description in 1992 from new tropical region, *P. marginatus* has spread to several Caribbean islands and central and south America (Miller *et al.*, 1999; Matile-Ferro *et al.*, 2000; Kauffman *et al.*, 2001a; Watson and Chandler, 1999; Miller and Miller,1999), Mexico (Williams & Granara de Willink, 1992); U.S. Virgin Islands (CABI/EPPO, 2000); The Dominican Republic (CABI/EPPO, 2000) and Grenada in 1994, Antigua and Barbuda (CABI/EPPO, 2000), Saint Martin (Pollard, 1999) and The British Virgin Islands in 1996 (CABI/EPPO, 2000); USA (Florida) (Pollard, 1999; Miller and Miller, 2002; Walker et al., 2006), Haiti, St. Kitts and Nevis (CABI/EPPO, 2000); St Barthélemy (Ben Dov, 2008) Guatemala (Ben Dov, 2008), Haiti (CABI/EPPO, 2000) and Guadaloupe (Ben Dov, 2008) in 1998; French Guyana (Ben Dov, 2008), Guiana (Ben Dov, 2008), Guadeloupe (Matile-Ferrero and Etienne, 1998), Cuba (CABI/EPPO, 2000), and Puerto Rico in 1999 (CABI/EPPO, 2000); Barbados (CABI/EPPO, 2000); Belize (Ben Dov, 2010), the Cayman Islands (CABI/EPPO, 2000), Costa Rica (Ben Dov, 2010), Cayman and Montserrat in 2000 (CABI/EPPO, 2000), Netherlands Antilles (CABI/EPPO, 2000), the Bahamas and Guam in 2002-03 ((Meyerdirk et al., 2004); Palau in 2003 (Anonymous, 2003; Muniappan et al., 2006) and neighboring islands in the Pacific (Meyerdirk et al., 2004).; Hawaii-Maui and Oahu in 2004 (Heu and Fukada, 2005; Heu et al., 2007), the Northern Marianas (Tinian) in 2005 and the Northern Marianas (Tinian) in 2005.

In Africa, it was reported in Ghana in 2009 (Cham *et al.*, 2011). In South and South East Asian region it was noticed in during 2008-09. In May 2008, it was recorded in Java, Indonesia and later spread to Bali and Sulawesi

Islands (Muniappan *et al.*, 2008 & 2009). It was also reported in July 2008 in Colombo and Gampaha districts in Sri Lanka (Galanihe *et al.*, 2010), Joyedpur in Bangladesh; Phnom Penh in Cambodia in 2010, Manila in Philippines in 2008; Thailand in 2010 (Muniappan *et al.*, 2009). In Malaysia, *P. marginatus* was first reported from Negeri Sembilan and Selangor in February, 2009 (Muniappan *et al.*, 2008; Mastoi *et al.*, 2011) and Taiwan in 2010 (Chen *et al.*, 2011) and Maldives very recently.

In India, it was first reported from Coimbatore in Tamil Nadu in July, 2008 (Muniappan *et al.*, 2008 & 2009; Regupathy and Ayyasamy, 2009; Suresh *et al.*, 2010). Since July 2008, it has spread subsequently to neighboring states such as Karnataka, Andhra Pradesh, Maharashtra, Kerala and to distant states like Tripura, Assam and Orissa in India (Shylesha *et al.*, 2011c; Rabindra, 2010; Krishanamurthy and Mani 2011; Sajeev, 2011; Jacob Mathew, 2011; Mani Chellappan, 2011a; Chandele *et al.*, 2011; Lyla and Philip, 2010; Krishnakumar and Rajan, 2009; Mahalingam *et al.*, 2010; Suresh *et al.*, 2010).

SYSTEMATICS

Paracoccus marginatus specimens were collected first in 1955 in Mexico, but it was described in 1992 from the specimen collected in neotropical region (Belize, Costa Rica, Guatemala and Mexico) by Williams & Granara de Willink (1992) and later redescribed by Miller and Miller (2002) and Angeles Martinez and De Los Suris (2005). Miller and Miller (2002) gave a complete description of all the stages of the papaya mealybug.

Adult male *P. marginatus* has a distinct aedeagus with ventral lobes that are broad and cylindrical in dorsal ventral view, lateral pore clusters, a heavily sclerotised thorax and head and well developed wings. Antenna is 10 segmented with bristle shaped and fleshy setae. Eight abdominal tergite usually without setae Adult males are approximately 1.0 mm long, with an elongate oval body and the thorax is 0.3 mm wide. 17 numbers of cerari, presence of oral rims on margins of coxae, a heavily sclerotized thorax and head, and well-developed wings (Watson and Chandler, 1999).

Adult female body is greenish yellow, dusted with mealywax not thick enough to hide body colour, without discrete bare areas on dorsum, with many short waxy filaments around the body margin. Caudal filaments are about one fourth of the body length. Adult females are wingless, 2-3 mm long; and 1.4mm wide. The species epithet is derived from a Latin word, meaning enclosed with a border and refers to the border of oral rim tubular ducts (Williams & Granara de Willink, 1992).

BIOLOGY

Details on the biology of *P. marginatus* were given by Amarasekare *et al.* (2008 a &b), Wallker *et al.* (2006)

Table 1. Development of Paracoccus marginatus at different temperature (Amarasekare et al., 2008a)

Temp		Life history stage							
$(\pm^{\circ} C)$	Eag	D '	Second		Third		Fourth	Cumulative	
	Egg	FIISt	Male	Female	Male	Female	Male	Male	Female
15	27.5±0.2ª								
18	23.1±0.2 ^b	25.3±0.5ª	21.1±1.6ª	13.5±1.3ª	$7.0{\pm}1.8^{a}$	13.2±0.9ª	11.7 ± 1.8^{a}	85.2 ± 1.8^{a}	74.4±1.4ª
20	14.4±0.2°	14.6±0.5 ^b	13.6±0.8 ^b	9.3±0.7 ^b	4.5 ± 0.7 ab	8.9±0.9 ^b	8.9 ± 0.7^{a}	53.4±0.7 ^b	45.9±0.9 ^b
25	8.7 ± 0.1^{d}	$6.5 \pm 0.1^{\circ}$	$6.6 \pm 0.5^{\circ}$	5.5±0.5°	2.4±0.5 ^b	5.2±0.2°	4.1 ± 0.5^{b}	28.5±0.3°	25.9±0.2°
30	7.3±0.2°	6.1± 0.2°	$6.3 \pm 0.4^{\circ}$	5.7±0.4°	$4.4 \pm 0.3^{\circ}$	4.4±0.3°	3.6 ± 0.4^{b}	24.9±0.6°	23.2± 0.3°
34	$5.9 \pm 0.1^{\text{f}}$	-	-	_	_	-	-	_	-
35	$5.5 \pm 0.1^{\text{f}}$	-	-	_	_	-	-	_	-
37	0.0	_	_	_	_	-	_	_	-
F	1922.10	100.59	57.41	17.09	5.35	15.31	15.66	725.42	521.23
df	6.212	3.132	3.97	3.104	3.94	3.95	3.91	3.84	3.90
р	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0020	< 0.0001	< 0.0001	< 0.0001	< 0.0001

Means within a column followed by the same letters are not significantly different at $\dot{a} = 0.05$ (Tukey's test n = 35)

and Thangamalar *et al.* (2010). There were differences in the developmental times of *P. marginatus* when reared on different host plants and also at different temperatures.

A single female lays about 230 eggs (Amarasekare *et al.*, 2008a) and 300-400 eggs (Mahalingam *et al.*, 2010) in an ovisac. The ovisac, developed ventrally, is three to four times the body length and entirely covered with white wax ((Pantoja *et al.*, 2007).

Females undergo only three instars and no sexual dimorphism between male and femal instars. However the duration of I, II and III instar nymph in mealybug was 6.5, 5.5 and 5.2 day respectively at 25° C (Amarasekare *et al.*, 2008a). The species is known to reproduce both sexually and parthenogenitically (CAB International, 2001). Adult female mealybug secretes large amounts of white wax. When specimens are placed in alcohol, they turned into blue black (Pantoja *et al.*, 2007).

Males have longer development time (27-30 days) than females (24-26 days) at $25\pm1^{\circ}$ C, $65\pm2\%$ RH and 12:12 (L:D) photoperiod (CAB International, 2001). Mean longevity of adult males and females was 2.3 and 21.2 days respectively (Amarasekare *et al.*, 2008a).

The ovipositional period is usually one to two weeks. Eggs are greenish yellow and egg hatching occurs in about 10 days.

Males undergo four instars, the first instar nymphs are called as crawlers, and the duration of I, II, III and IV instar male nymph was 6.5, 6.6, 2.4 and 41 days respectively at 25° C (Table 1). The fourth instar, referred as pupa says in the cocoon. Adult males are yellow in the first and second instar and later turn pink during the pre-pupal and pupal stages (Amarasekare *et al.*, 2008 a).

DAMAGE

Papaya mealybug infestations are typically observed as clusters of cotton-like masses on the above-ground portion of plants. *Paracoccus marginatus* damages various parts of the host plant including the leaves, stems, flowers and fruits. *P. marginatus* may show very similar symptoms to pink hibiscus mealybug, *Maconellicoccus hirsutus* (Green) (Pollard, 1999). The insect sucks the sap by inserting its stylets into the epidermis of the leaf, fruit and stem. While feeding, it injects a toxic substance into the leaves resulting in curling, crinkling, rosetting, twisting and general leaf distortion (Miller *et al.*, 1999; Walker *et al.*, 2003; Heu and Fukada, 2005; Pantoja *et al.*, 2007). Heavy mealybug infestations render fruit inedible. Due to the build-up of thick white waxy coating and sooty mould development on the honeydew excreted by mealybug, infested fruits get reduced market value. Fruits may fail to develop normally and may be unusually small. Such fruits eventually shrivel and drop (Tanwar *et al.*, 2010; Heu *et al.*, 2007). Some economically important crops such as papaya, mulberry, cotton, cassava, citrus, sweet potato, peas and beans, okra, eggplant, guava and ornamentals such as hibiscus, *Jathropha, Allamanda, Acalypha* McComie, 2000b; Meyerdirk *et al.*, 2000; Shylesha *et al.*, 2011b).

ECOLOGY

The incidence of *P. marginatus* was noticed throughout the year, but active in warm dry weather. Prolonged drought with scanty rain fall and less number of rainy days favour the faster multiplication (Ayyasamy and Regupahy, 2010). During the rainy season, population of *P. marginatus* especially the crawlers decreased drastically because heavy rain washed the insects off the plants. However, mealybugs sheltered within unopened leaves and other hiding places survived and built up their numbers again during the warm, dry weather. The climatic preferences of *P. marginatus* have been documented well, but its occurrence in countries located 30°C from the Equator suggest that probably does not tolerate cold conditions (CAB International, 2001).

HOST PLANTS

It is a highly polyphagus insect pest that can damage large number of tropical and subtropical fruits, vegetables and ornamental plants (Miller and Miller, 2002). According to Muniyappan *et al.* (2008), it was known to infest plants belonging to 22 families from Asia. Galanihe *et al.*, (2010) recorded more than 40 plant species in Sri Lanka compared to 55 plants species recorded in Florida (Walker *et al.*, 2003). *P. aracoccus marginatus* attacks over 60 species of plants including field crops, fruit trees ornamentals, weed and scrub vegetation in India (Shylesha *et al.*, 2011b).

NATURAL ENEMIES

It has never gained the status as a pest in the native home of Mexico, Central and North America probably, due to presence of endemic natural enemy complex (Walker *et al.*, 2003). The *P. marginatus* papaya mealybug became pest when it invaded the Caribbean region maily due to the absence of natural enemies. *Spalgius epeius* Westwood was the predominant natural

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enemy on *P. marginatus* on papaya mealybug damaging several host plants in South India (Thangamalar *et al.*, 2010). *Cryptolaemus montrouzieri* Mulsant, a general predator of mealybug was also recorded occasionally on *P. marginatus* papaya mealybug in India and elsewhere. Parasitoids of *P. marginatus* from Mexico and Caribbean are listed by Schauff (2000). Four species of chalcidoid parasitoids and two predators were found attacking *P. marginatus* in Malaysia (Mastoi *et al.*, 2011).

A total of 22 natural enemies occurring either naturally /introduced were reported on papaya mealybug in different countries (Table 2).

 Table 2. List of host plants for papaya mealybug, Paracoccus marginatus

Family and Species	Country	References	
Caricaceae			
Carica Carica papaya	Ghana, India Florida Sri Lanka Malaysia Puerto Rico Indonesia Hawaii	Cham <i>et al.</i> (2011) Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010) Mani Chellappan (2011a) Muniappan <i>et al.</i> (2008) Jacob Mathew (2011) Walker <i>et al.</i> (2003) Miller and Miller (2002) Galanihe <i>et al.</i> (2010) Mastoi <i>et al.</i> (2011) Pantoja <i>et al.</i> (2007) Muniappan <i>et al.</i> (2007)	Euph hirta Euph hetero Hura crepiu Phyll nirur Acaly Conv
	Palau	Muniappan et al. (2006)	aceae Ipome
Anacar- diaceae Mangifera indica	Ghana Florida Sri Lanka Palau India	Cham et al. (2011) Walker et al. (2003) Galanihe et al. (2010) Anonymous (2003) Muniappan et al. (2008) Jacob Mathew (2011) Shylesha et al. (2011d)	batata Legun /Faba <i>Cajar</i>
Anogunogogo	Indonesia Hawaii	Muniappan <i>et al.</i> (2008) Ronald <i>et al.</i> (2007)	Glyri sepiu
Plumeria alba	Ghana	Cham et al. (2011)	Phase
Plumeria rubra	Ghana Palau Florida India	Cham <i>et al.</i> (2011) Muniappan <i>et al.</i> (2006) Miller and Miller (2002) Muniappan <i>et al.</i> (2008)	vulga
		Mani Chellappan, (2011a) Jacob Mathew (2011) Shylesha <i>et al.</i> (2011d)	Vigna ungui

Family and Species	Country	References
	Indonesia Hawaii Guam	Muniappan <i>et al.</i> (2008) Ronald <i>et al.</i> (2007) Meyerdirka <i>et al.</i> (2004)
Euphorbiaceae Manihot esculenta	Ghana India Sri Lanka Palau Puerto Rico Florida Indonesia Malaysia	Cham et al. (2011) Mani Chellappan (2011a) Jacob Mathew (2011) Muniappan et al. (2008) Tanwar et al. (2010) Shylesha et al. (2010) Galanihe et al. (2010) Muniappan et al.(2006) Pantoja et al. (2007) Miller and Miller (2002) Muniappan et al. (2008) Mastoi et al. (2011)
Euphorbia hirta	Ghana India	Cham <i>et al.</i> (2011) Tanwar <i>et al.</i> (2010) Shylesha <i>et al.</i> (2011d)
Euphorbia heterophylla	Ghana	Cham et al. (2011)
Hura crepitans Phyllanthus	Ghana	Cham et al. (2011)
niruri Acalypha sp	India Florida	Tanwar <i>et al.</i> (2010) Miller and Miller (2002)
Convolvul- aceae Ipomoea batatas	Ghana Puerto Rico India Indonesia	Cham <i>et al.</i> (2011) Pantoja <i>et al.</i> (2007) Muniappan <i>et al.</i> (2008) Muniappan <i>et al.</i> (2008)
Leguminaceae /Fabaceae <i>Cajanus cajan</i>	Hawan Ghana India	Konaid <i>et al.</i> (2007) Cham <i>et al.</i> (2011) Tanwar <i>et al.</i> (2010) Shylesha <i>et al.</i> (2011d) Muniappan <i>et al.</i> (2008)
Glyricidia sepium	Ghana	Cham <i>et al.</i> (2011)
Phaseolus vulgaris	Ghana Florida Sri Lanka Palau Hawaii	Cham <i>et al.</i> (2011) Walker <i>et al.</i> (2003) Galanihe <i>et al.</i> (2010) Muniappan <i>et al.</i> (2006) Ronald <i>et al.</i> (2007)
Vigna unguiculata	Ghana India	Cham <i>et al.</i> (2011) Mani Chellappan (2011a) Shylesha <i>et al.</i> (2011d)

Family and Species	Country	References	Family and Species	Country	References
Arachis hypogaea	Ghana	Cham <i>et al.</i> (2011)	Lycopersicon esculentum	Ghana	Cham <i>et al.</i> (2011) Topwar <i>et al.</i> (2010)
Phaseolus atropurpureus	Ghana	Cham et al. (2011)		Florida	Mani Chellappan (2011a) Walker <i>et al.</i> (2003)
Gliricidia sepium	Sri Lanka	Galanihe <i>et al.</i> (2010)		Palau	Galanihe <i>et al.</i> (2010) Muniappan <i>et al.</i> (2006)
n Mimosa nigra	Florida	Miller and Miller (2002)		Hawaii	Ronald et al.(2007)
Acacia	Florida	Miller and Miller (2002)	Solanum melongena	Ghana	Cham <i>et al.</i> (2011)
Malvaceae Abelmoschus	Fiorida	while and while (2002)	0	India Florida	Tanwar <i>et al.</i> (2010) Mani Chellappan (2011a) Walker <i>et al.</i> (2003)
esculentus	Ghana	Cham et al. (2011)		1101144	Miller and Miller (2002)
Abutilon	Ghana	Cham et al. (2011)		Sri Lanka	Galanihe <i>et al.</i> (2010)
indicum Sida sp	India Ghana Florida	Tanwar <i>et al.</i> (2010) Cham <i>et al.</i> (2011) Miller and Miller (2002)		Hawaii Malaysia	Ronald <i>et al.</i> (2007) Mastoi <i>et al.</i> (2011)
Ceiba				Puerto Rico	Pantoja <i>et al.</i> (2007)
pentandra	Ghana	Cham et al. (2011)	Solanaum torvum	India	Tanwar <i>et al.</i> (2010)
Gossypium hirsutum	India Florida	Tanwar <i>et al.</i> (2010) Walker <i>et al.</i> (2003)	Compositae Tridax	Ghana	Cham <i>et al.</i> (2011)
Hibiscus			procumbens	India	Tanwar et al. (2010)
rosa sinensis	India	Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010)	Launaea taraxacifolia	Ghana	Cham et al. (2011)
		Muniappan, <i>et al.</i> (2008) Jacob Mathew (2011)	Cucurbitaceae Luffa cylindrical	Ghana	Cham <i>et al.</i> (2011)
	Florida	Walker <i>et al.</i> (2003) Miller and Miller (2002)	Curcubita sp	Ghana Palau	Cham et al. (2011) Muniappan et al. (2006)
	Sri Lanka	Galanihe <i>et al.</i> (2010)	Ronincasa	1 alau	Humappun et ut. (2000)
	Puerto Rico	Mastoi <i>et al.</i> (2011) Pantoja <i>et al.</i> (2007)	hispida	India	Mani Chellappan (2011a)
	Hawaii Palau Guam	Ronald <i>et al.</i> (2008) Muniappan <i>et al.</i> (2007) Meyerdirka <i>et al.</i> (2004)	Rutaceae <i>Citrus</i> sp.	Ghana Palau Florida Sri Lanka	Cham <i>et al.</i> (2011) Muniappan <i>et al.</i> (2006) Walker <i>et al.</i> (2003) Galanihe <i>et al.</i> (2010)
Solanaceae Capsicum			Murraya koenigii	India	Mani Chellannan (2011a)
annuum	Ghana Hawaii	Cham <i>et al.</i> (2011) Ronald <i>et al.</i> (2007	Sterculiaceae		enemppun (2011u)
	India Sri Lanka	Mani Chellappan (2011a) Galanihe <i>et al.</i> (2010)	Theobroma cacao	Ghana	Cham et al. (2011)
Cansicum			Lauraceae		
frutescens	Ghana Florida Palau	Cham <i>et al.</i> (2011) Walker <i>et al.</i> (2003) Muniappan <i>et al.</i> (2006)	Persea americana	Ghana Florida Puerto Rico Florida	Cham <i>et al.</i> (2011) Walker <i>et al.</i> (2003) Pantoja <i>et al.</i> (2007) Miller and Miller (2002)

Family and Species	Country	References	Family and Species	Country	References
	India Indonesia	Muniappan <i>et al.</i> (2008) Muniappan <i>et al.</i> (2008)	Wedelia trilobata	Ghana Florida	Cham <i>et al.</i> (2011) Miller and Miller (2002)
Bombacaceae Adansonia digitata	Ghana	Cham et al. (2011)	Hevea brasiliensi	Sri Lanka India	Galanihe <i>et al.</i> (2010) Mani Chellappan (2011a) Jacob Mathew (2011)
Plumbagi- naceae <i>Plumbago</i>			Tagetes erecta	Sri Lanka	Galanihe et al. (2010)
<i>auriculata</i> Moraceae	Ghana	Cham et al. (2011)	Helianthus annuus	Sri Lanka	Galanihe et al. (2010)
Morus alba	India	Tanwar et al. (2010)	Aizoaceae Trianthema		
Psidium guaiava	India	Tanwar <i>et al.</i> (2010) Mani Chellappan (2011a)	portulacastrum	India	Tanwar <i>et al.</i> (2010)
Verbenaceae	Sri Lanka	Galanihe <i>et al.</i> 2010	Rubiaceae Canthium inerme	India	Tanwar et al. (2010)
Tectona grandis	India	Tanwar et al. (2010)	Moraceae Artocarpus		
Amarantha- ceae Achyranthus			integrifolia	Sri Lanka Malaysia India	Galanihe <i>et al.</i> (2010) Mastoi <i>et al.</i> (2011) Mani Chellannan (2011a)
aspera	India	Tanwar <i>et al.</i> (2010)	D '	Inuia	Main Chenappan (2011a)
Amaranthus cruentus	India	Mani Chellappan (2011a)	Punicaceae Punica granatum	Florida Sri Lanka	Walker <i>et al.</i> (2003) Galanihe <i>et al.</i> (2010)
Capridaceae Cleome viscosa	India	Tanwar <i>et al.</i> (2010)	Oleaceae Jasminum spp	Sri Lanka	Galanihe et al. (2010)
Commeli- naceae <i>Commelina</i>			Sapindaceae Nephelium lappecium	Sri Lanka Malaysia	Galanihe <i>et al.</i> (2010) Mastoi <i>et al.</i> (2011)
benghalensis Convolvu- laceae	India	Tanwar <i>et al.</i> (2010)	Musaceae Musa		
Convolvulus			sapientum	Sri Lanka	Galanihe et al. (2010)
arvensis	India	Tanwar <i>et al.</i> 2010	Arecaceae Cocos nucifera	Sri Lanka	Galanihe et al. (2010)
batatas	Florida	Walker <i>et al.</i> (2003)	Oxalidaceae	Malaysia	Mastoi et al. (2011)
Lamiaceae	Palau	Muniappan <i>et al.</i> (2006)	Annonaceae Annona spp	Florida	Miller and Miller (2002)
Leucas aspera	India	Tanwar et al. (2010)	Poaceae		
Ocimum	India	Tanwar <i>et al.</i> (2010)	Zea mays	Florida	Miller and Miller (2002)
sanctum Asteraceae Parthenium	Ghana	Cham <i>et al.</i> (2011)	Phyllantha- ceae Phyllanthus		
hysterophorus	India	Tanwar et al. (2010)	emblica	India	Mani Chellappan (2011a)

Papaya mealybug and biological control

MANAGEMENT

Mealybugs are difficult to control because they live in protected areas such as cracks, crevices and under the bark of their host plants. Most of the stages including eggs of mealybug are covered with waxy secretions that protect them. An integrated pest management (IPM) approach involving cultural practices, legal, chemical and biological control is advisable.

(1) Legal

Strict quarantine measures are needed to prevent the entry of mealybug infested planting materials/fruits/ flowers from other countries. Domestic quarantine measures are to be strengthened to prevent the movement from one state to other states within the country (Tanwar *et al.*, 2010).

(2) Cultural Control

Planting material free from mealybugs is to be used. In the initial stages of appearance of mealybug, collection and destruction of infested plant parts are to be carried out (Ayyasamy and Regupathy, 2010; Tanwar *et al.*, 2010).

(3) Chemical Control

Chemicals were used desperately when there was outbreak of mealybugs, and other methods were not available immediately. A number of insecticides like monocrotophos, methyl demeton, dimethoate, acephate, methomyl, fenthion, imidacloprid, thiomethoxam, dichlorovos, quinalphos, profenophos, fenitrothion, carbaryl, chlorpyriphos, diazinon, malathion, buprofezin were used against papaya mealybug (Tanwar et al., 2010; Regupathy and Ayyasamy, 2009; Mahalingam et al., 2010; Banu et al., 2010; Suresh et al., 2010). They give short-term control but chemical control is difficult and requires repeated application of the insecticides (Tanwar et al., 2010; Ayyasamy and Regupathy, 2010; Galanihe et al., 2010)). The chemicals were recommended for the control of the mealybug until the biological control agents could be introduced.

(4) Biopesticides

Fish oil rosin soap, azadirachtin and white mineral oils were found partially effective against papaya mealybug. The three fungal pathogens *Verticillium lecanii* (Zimm.), *Beauria bessiana* (Bals.) and *Metarhium anisopliae* (Metsch.) were known to cause 40-50% mortality of *P. marginatus* (Banu *et al.*, 2010).

(5) Biological Control

Though several methods were available, excellent control of mealybug was obtained with use of biocolntrol agent thoughout the World (Myerdirk, 2000). In case of PMB also, outstanding control was achieved with use of parasitoids in several countries (Mani and Shivaraju, 2012; Shylesha *et al.*, 2011c).

Guam

P. marginatus was reported in April 2002; Survey of P. marginatus in Guam before the release of the parasitoids showed that there were no local parasitoids recorded on this mealybug. A few coccinellids such as C. montrouzieri and Chilocorus nigrita (Fabricius) were however, found feeding on it. They were not capable of suppressing the populations of P. marginatus. The parasitoids, Acerophagus papayae, Anagyrus loecki and Pseudleptomastix mexicana totaling 46, 200 individuals were introduced from Puerto Rico, and released in Guam from June to October, 2002. Establishment of the parasitoids was confirmed within a month of release at the sample sites and releases were continued at other geographical locations across the Island. A reduction of over 99% of PMB was observed about a year of introduction of these parasitoids. By August 2003, the population of PMB declined to a level which was hard to find in the field. Almost all papaya, Plumeria spp. and Hibiscus spp. plants recovered and no symptoms of damage were noted at that time (Meyerdirk et al., 2004).

Palau

The pest was reported in March 2003, and was causing serious damage to papaya plumeria, Hibiscus and many other plants. Very few C. montrouzieri larvae and adults were encountered on P. marginatus in the survey. The parasitoids A. loecki, P. mexicana and A. papayae totaling 24, 586 were imported from Puerto Rico, and released in Palau from August 2003 to June 2004. Establishment of parasitoids was confirmed within a month. A. loecki and A. papavae appeared to be promising biological control agents of PMB in Palau. No field recovery of P. mexicana was made in spite of several field releases. The reduction of the papaya mealybug population density levels below detectable levels was observed in a six-month period following the introduction of these exotic parasitoids. Following the successful implementation of a classical biological control program, the risk of this mealybug spreading to other islands in the Republic of Palau and to neighboring Micronesian Islands has been considerably reduced (Muniyappan et al., 2006).

Sri Lanka

The PMB was reported on a large number of plant species in Columbo and Gampha district in Sri Lanka for the first time in 2008. It has caused worst damage in papaya growing districts of Sri Lanka. A classical biological control work was initiated in 2009. Three parasitoids *A. loecki* (2000), *P. Mexicana* (3200) and *Acerophagus papayae* (4800) was released in October, 2009. After two to three months, *A. papayae* established in all the sites and subsequently PMB was controlled to level of 90-100 per cent by December, 2009 (Wahundenya *et al.* 2009 Personal Communication, 2009).

Mexico

Biological control appears to be the main factor keeping the species under control in Mexico. The most important natural enemies were the encyrtids, *Anagyrus* spp., *Acerophagus* spp. and *Apoanagyrus* spp. The general predators such as *Chrysopa* spp. and *Chilocorus* spp. were also encountered in low densities on PMB (Gonzalez *et al.*, 1999).

Puerto Rico and Dominican Republic

Paracoccus marginatus was first intercepted from Puerto Rico in 1995, and by 1998 it was found to be distributed throughout Puerto Rico with a higher density on the west side of the Island (Saez, 2000). During 2001-2002, severe infestation of papaya mealybug required several insecticides applications to control pest (Pantoja *et al.*, 2007).

USDA-APHIS found that the five parasitoid species, *Anagyrus loecki, Apoanagyrus californicus, Acerophagus* sp. and *Pseudophycus* sp and *Pseudleptomasix mexicana* brought about a 99.7% reduction in papaya mealybug populations in the Dominican Republic, and a 97% reduction in Puerto Rico, with parasitism levels of 35.5-58.3% (Kauffman *et al.*, 2001a; Meyerdirk and Kauffman, 2001). However, *Acerophagus* sp. emerged as the dominant parasitoid species in both Puerto Rico and the Dominican Republic (Meyerdirk and Kauffman, 2001; Ramirez and Saez, 2002; Walker *et al.*, 2003; Arnold, 2001; Kauffman *et al.*, 2001b).

Florida

Paracoccus marginatus was discovered in Florida 1998, the USDA Animal and Plant Health Inspection Service (APHIS) and USDA Agricultural Research Service (ARS) initiated a classical biological control programme for the papaya mealybug (Ngyen, 2000). Four genera of encyrtid endoparasitoid wasps specific to the mealybug were collected in Mexico by USDA and ARS researchers and Mexican cooperators as potential biological control agents: *Acerophagus papayae*, *Anagyrus loecki*, *Anagyrus californicus* and *Pseudaphycus* sp. (USDA 1999, 2000; Meyerdirk and Kauffman 2001). A fifth collected species was later reared and identified as *Pseudleptomastix mexicana* (Noyes and Schauff 2003). The first releases of these four parasitoids were made in Florida in October 2000 (Walker *et al.*, 2003) and again released in 2003 (Meyerdirk, 2003). Allthough it is belived that these parasitoids are established in the released areas, *Acerophagus papayae* had higher per cent parasitism than *A. loecki* and there is no recovery of *P. mexicana* (Kaushalya *et al.*, 2008).

India

Paracoccus marginatus invaded India in 2008 and has become severe on several agricultural and horticultural crops. The potential economic loss due to this pest ranges from 60-80% in papaya. The parasitoids *Acerophagus papayae*, *Pseudleptomastix mexicana* and *Anagyrus loecki* from USDA-APHIS Puerto Rico were shipped to India. A total of 3429 of *A. papayae*, 1485 of *P. mexicana* and 516 of *A. loecki* were received by National Bureau of Agriculturaly Important Insects, Bangalore during July- October, 2010. After ascertaining the safety in quarantine, these three parasitoids were distributed to different ststes in India.

During October 2010, *Acerophagus papayae* was released on jatropha and plumeria in Bangalore, on papaya in Jakkur, Mysore, Chamaraj nagar, and Nelamangala and on mulberry in Mandya district and. *A. papayae* was observed in few numbers on 20 days after release and the spread of the parasitoid was very good on 40 and 60th day of observation. After a span of 3 months, there was a reduction of 80-90% in the pest population, and new shoots developing were not found to harbor any mealybug (Shylesha *et al.*, 2010; Shylesha *et al.*, 2011; Qadri *et al.*, 2011).

Paracoccus marginatus was reported in Pune region of Western Maharashtra on July, 2010. It assumed the status of major pest of papaya in Pune, Nandurbar and Jalgaon districts of Maharashtra. The parasitoid *Acerophagus papayae* was found parasitizing for the first time in August, 2010. Subsequently a release rate of 1000-1500/ac was recommended for several papaya gardens in Maharastra. There was 85-92% declined in the mealybug population within three months (Pokharkar *et al.*, 2011; Mundale and Nakat, 2011; Chandale *et al.*, 2011; Nakat *et al.*, 2011). _

Family and	Country	References	Species		
Species			Apoanagyrus	Mexico	Meyerdirka and
Hymenoptera			californicus		Kauffman (2001)
Encyrtidae			Compere	Puerto Rico	Pantoja et al. (2007)
Acerophagus papayae Noyes and			Pseudlepto- mastrix mexicana	India	Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010) Ayyasamy and Ragupathy
Schauff	India	Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010) Jothi <i>et al.</i> (2011) Ayyasamy and Ragupathy (2010) Chandale <i>et al.</i> (2011) Qadri (2011)	Noyes and Schauff	Indonesia Sri Lanka	(2010) Chandale <i>et al.</i> (2011) Qadri (2011) Nakata <i>et al.</i> (2011) Muniappan, <i>et al.</i> (2008) Kalyanasundaram <i>et al.</i> (2011) Muniappan <i>et al.</i> (2008) Galanihe <i>et al.</i> (2010)
		Kalyanasundaram <i>et al.</i> (2011) Muniappan <i>et al.</i> (2008)		Malaysia Puerto Rico Florida	Mastoi <i>et al.</i> (2011) Pantoja <i>et al.</i> (2007) Miller and Miller (2002)
	To to one to	Jacob Mathew (2011)		Indonesia	Muniappan <i>et al.</i> (2008)
	Indonesia Sri Lanka	Galanihe <i>et al.</i> (2008)		Hawaii	Ronald <i>et al.</i> (2007)
	Malaysia	Mastoi <i>et al.</i> (2011)		Palau Florida	Muniappan <i>et al.</i> (2006) Kaushalva <i>et al.</i> (2010)
	Puerto Rico	Pantoja et al. (2007)		Mexico	Meyerdirka and Kauffman
	Indonesia	Muniappan et al. (2008)			(2001)
	Hawaii	Ronald et al. (2007)		Guam	Meyerdirka et al. (2004)
	Palau	Muniappan <i>et al.</i> (2006)	Pseudaphycus		
	Florida	Kausnalya <i>et al.</i> (2010) Miller and Miller (2002)	sp.	Mexico	Meyerdirka and Kauffman
	Mexico	Meverdirka and Kauffman		(2001)	
Anaovrus		(2011)	Lepidoptera: Lycaenidae		
loecki Noves	India	Shylesha <i>et al.</i> (2011d)	(Westwood)	India	Shylesha <i>et al.</i> (2011d)
1000000 110900		Tanwar $et al.$ (2010)	(11050110000)		Tanwar $et al.$ (2010)
		Jothi et al. (2011)			Jothi et al. (2011)
		Ayyasamy and Ragupathy			Jonathan <i>et al.</i> (2011)
		(2010)			Krishnamurthy and Mani
		Chandale <i>et al.</i> (2011)			(2011)
		Nakata <i>et al.</i> (2011)			Chandale et al. (2011)
		Kalyanasundaram <i>et al.</i> (2011)			Nakat et al. (2011)
		Muniappan, et al. (2008)	Coleoptera:		
		Jacob Mathew (2011)	Coccinellidae		
	Indonesia	Muniappan et al. (2008)	Cryptolaemus		
	Sri Lanka	Galanihe et al. (2010)	Mulsant	India	Shylesha <i>et al.</i> (2011d)
	Malaysia	Mastoi <i>et al.</i> (2011)			Tanwar $et al.$ (2010)
	Puerto Rico Pantoja <i>et al.</i> (2007)				Nakata et al. (2011)
	Hawaii	Ronald <i>et al.</i> (2007)			Jothi et al. (2011)
	Palau	Muniappan $et al.$ (2006)			Ayyasamy and Ragupathy
	Florida	Kaushalya <i>et al.</i> (2010)			(2010) Jonathan <i>et al.</i> (2011)
		Miller and Miller (2002)		Malaysia	Mastoi <i>et al.</i> (2011)
	Mexico	Meyerdirka and Kauffman (2001)		Palau Hawaii	Muniappan <i>et al.</i> (2008) Ronald <i>et al.</i> (2007)

Family and

Country

References

Table 3. List of natural enemies on Paracoccus marginatus

Family and Species	Country	References	Family and Species	Country	References
	Florida Guam British	Walker <i>et al.</i> (2011) Anon (2010)			Ayyasamy and Ragupathy (2010)
	Virgin Island	Meyerdirka et al. (2004) CAB International (2011)	Apertochrysa sp.	Malaysia	Mastoi et al. (2011)
Nephus bilucernarius (Mulsant)	Hawaii	Ronald et al. (2007)	Diptera: Syrphidae Ischiodon	India	Shylesha at al. $(2011d)$
taiwanus (Ohta)	India	Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010) Nakata <i>et al.</i> (2011) Chandele <i>et al.</i> (2011) Jonathan <i>et al.</i> (2011)	Entomopatho- genic fungi <i>Metarrhizium</i>	muia	Tanwar <i>et al.</i> (2011d) Jonathan <i>et al.</i> (2011)
Brumoides suturalis	Hawaii	Ronald <i>et al.</i> (2007)	anisopliae (Metsch.)	India	Shylesha <i>et al.</i> (2011d) Ayyasamy and
Hyperaspis silvestrii Weise	Hawaii Hawaii	Ronald <i>et al.</i> (2007) Ronald <i>et al.</i> (2007)	Verticillium lecanii (Zimm.)	India	Ragupathy (2010)
Curinus coeruleus Mulsant	Hawaii	Ronald et al. (2007)	(Ziiiiii.)	muta	Ayyasamy and Ragupathy (2010) Jonathan <i>et al.</i> (2011)
Cheilomenus sexmaculatus (F.)	India	Jonathan et al. (2011)	Paecilomyces	India	Mani Chellappan (2011b) Ayyasamy and
Coccinella transversalis Fabricius	India	Jonathan et al. (2011)	pictus Beauveria bassiana		Ragupathy (2010)
Neuroptera: Chrysopidae			(Bals.) Neozygytes	India India	Shylesha <i>et al.</i> (2011d) Shylesha <i>et al.</i> (2011d)
Chrysoperla carnea (Stephens)	India	Shylesha <i>et al.</i> (2011d) Tanwar <i>et al.</i> (2010)	Chilocorus nigrita Fab	Guam Mexico	Meayerdirk et al. (2004) Gonzalez et al. (1999)

In Tamil Nadu, *P. marginatus* was first observed during July 2008 in Coimbatore district and subsequently spread to neighboring districts of Tamil nadu. Initially several chemicals were used for the control of PMB. During the October 2010, the parasitoids, *Acerophagus papayae*, *Pseudleptomastix mexicana* and *Anagyrus loecki* were obtained from NBAII and reared initially. *A. papayae* was subsequently multiplied on large scale and released in the farmer's field of papaya in several districts in Tamil nadu. There was substantial reduction of PMB density to very low level with three month of

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after its introduction (Kalyanasundaram *et al.*, 2011; Jonathan *et al.*, 2011; Regupathy and Ayyasamy, 2011).

P. marginatus is a serious pest on rubber, several forest trees, vegetables, fruits, plantation crops, flower crops and other weed plants in Kerala. Biological control programme was initiated in 2011 with the release of *A. papayae* in Kerala (Mani Challappan 2011b; Jacob Methew, 2011; Sajeev, 2011).

In eastern state of Odisha, *P. marginatus* is a serious pest on several agriculture and horticultural crops in

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2011. Biological control programme was initiated in 2011 itself with the release of *A. papayae* (Shylesha *et al.*, 2011a). Similarly in north eastern state of Tripura, more than 60% of papaya plants were infested with *P. marginatus* during 2009-10 in four districts of Tripura. Biological control programme was initiated in 2011 with the release of *A. papayae* (Agarawala, 2011).

Caribbean Islands

As an exotic introduction to the Caribbean islands, there were good prospects for control of *P. marginatus* by hymenopteran parasitoids originating from its area of origin in Central America (Pollard, 1999).

Malaysia

Paracoccus marginatus was reported for the first time in Malaysia on papaya, cassava, eggplant, jatropha and hibiscus plants. Four species of chalcidoid parasitoids were observed parasitizing the PMB. *Acerophagus papayae* was the major parasitoid of PMB. Two common predators namely *Apertochrysa* sp. and *Cryptolaemus montouzieri* were also found feeding on PMB (Mastoi *et al.*, 2011).

Taiwan

P. marginatus was found damaging papaya in Taiwan for the first time in 2011. *A. papayae* was useful in controlling the papaya mealybug in Taiwan. Bio-control is to be initiated for the control of PMB (Chen Shu Pei *et al.*, 2011).

Indonesia

The papaya mealybug, *Paracoccus marginatus* was recorded in Indonesia (Java) in 2008. Introduction of parasitoid, *A. papayae* is to be carried out in controlling the papaya mealybug in Indonesia (Herlina, 2011).

CONCLUSION

The papaya mealybug *Paracoccus marginatus* is likely to invade several countries in the World. Several methods like cultural, chemical, legal and biological have been adopted to tackle the mealybug problem. However only chemical and biological control methods have been widely practices for the management of mealybugs. Only the host specific parasitoids have given control of *P. marginatus*. More specifically the hymenopteran parasitoid *Acerophagus papayae* played a predominant role in suppressing the papaya mealybug in several countries. It is advisable to obtain *A. papayae* from the other countries and release the same whenever the invasion of *P. marginatus* was observed in the countries.

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