

Pathogenicity of Some Fungi to *Parthenium*, an Obnoxious Weed in Madhya Pradesh

A.K. PANDEY, R.S. LUKA, S.K. HASIJA and R.C. RAJAK

Department of Biological Science, Rani Durgavati University

Jabalpur - 482 001

ABSTRACT

Pathogenicity of nineteen fungi isolated earlier from living and diseased parts of exotic weed *Parthenium hysterophorus* L., in Madhya Pradesh was tested. *Colletotrichum gloeosporioides*, *Alternaria alternata*, *A. dianthi*, *A. macrosporus*, *Myrothecium roridum*, *Fusarium oxysporum*, *F. moniliforme*, *Phoma herbarum* and *Bipolaris* sp. were found to cause considerable damage to the weed under laboratory conditions.

KEY WORDS : *Parthenium hysterophorus*, fungi, control potential

The exotic weed *Parthenium hysterophorus* L. is a serious environmental, agricultural and medical hazard in various states of India (Kanchan, 1975; Towers *et al.*, 1977; Subba Rao *et al.*, 1978; Joshi, 1987). Considerable sum of money and human and fossil energy are spent for mechanical and chemical control. Plant pathogens, especially fungi have been successfully used in the past to control unwanted weeds (Templeton, 1990; Templeton and Heiny, 1990 and Van Dyke, 1991). Phytopathogenic fungi being highly virulent, host specific, genetically stable, environmentally safe and having a wide range of parasitism, offer great potential as mortality agents for many trouble-some weeds including *Parthenium* in India.

During a survey of microbial diseases of *Parthenium*, nineteen fungi were isolated from its living as well dead parts (Rajak *et al.*, 1990). In the present work, the pathogenic potentiality of these fungi was investigated.

MATERIALS AND METHODS

Standard mycopathological techniques (Conway, 1979; Agarwal and Hasija, 1986) were followed. Plants were raised in earthen pots. Aqueous suspension of conidia prepared in sterile distilled water with 0.05% (W/V) oxysorbic (20 POE) polyoxyethylene

sorbitan mono-oleate surfactant (Tween 80) at concentration of $2-2.3 \times 10^1$ conidia/ml were applied by wetting the foliage of the seedlings in the four to five leaf stage. Control plants were sprayed with sterile distilled water. Immediately following inoculation, plants were placed in Environmental Test Chamber (Remi CH-6 A) at $28^\circ\text{C} \pm 1^\circ\text{C}$ and RH 95% for 48 h and then removed to green house benches and evaluated 14 days after inoculation for disease potentiality.

Pathogenicity of wilt inducing fungi was determined following the methods of Scheffer and Walker (1953) and Sati and Grewal (1981). The effect of spore suspensions on seed germination and seedling growth were tested as per Mortensen and Hsiao (1987). The treatments were replicated thrice and the experiment repeated twice. For data analysis, methods of Mortensen (1985) were followed.

RESULTS AND DISCUSSION

Results presented in Table-1 show that the maximum reduction in the height as compared to control was observed in the plants treated with *Collectotrichum gloeosporioides* followed by *Alternaria alternata*, *Myrothecium roridum*, *Fusarium moniliforme*, *F. oxysporum*, *Phoma herbarum*, *Bipolaris* sp. and *Alternaria*

Table 1. Response of *P. hysterophorus* to spores of various fungi in green house experiments (average of 30 plants after 14 days of inoculation, at $28 \pm 1^\circ\text{C}$, 95% RH)

Name of the fungi tested	Dose Spores/ml $\times 10^2$	Reduction in height of treated plant (in cm)	% disease damage in leaf	% plant killed	% inhibition of seed germination (a)	% seedling mortality	Disease Index
<i>Alternaria alternata</i>	2.1	7.6	95	95	98	100	A
<i>A. dianthi</i>	2.2	2.5	90	91	90	90	A
<i>A. macrosporus</i>	2.1	5.5	92	93	85	80	A
<i>Acremonium alternatum</i>	2.0	1.0	15	5	80	70	B
<i>Aspergillus niger</i>	2.3	1.0	5	2	100	-	C
<i>Chaetomium globosum</i>	2.1	1.0	5	1	100	-	C
<i>Cladosporium gladosporioides</i>	2.2	2.2	20	3	60	20	D
<i>Colletotrichum gloeosporioides</i>	2.2	8.0	98	100	98	100	A
<i>Curvularia lunata</i>	2.1	1.5	40	5	80	75	B
<i>C. senegalensis</i>	2.2	2.0	35	10	85	66	B
<i>Dreschlera indica</i>	2.2	2.5	30	15	75	48	B
<i>Fusarium oxysporum</i>	2.3	6.5	100*	100	90	100	A
<i>F. moniliforme</i>	2.1	7.3	100*	100	98	100	A
<i>Bipolaris</i> sp.	2.3	5.5	80	85	75	50	C
<i>Myrothecium roridum</i>	2.2	7.5	98	100	95	100	A
<i>Pestalotia</i> sp.	2.0	2.5	25	20	40	50	C
<i>Paecilomyces varioti</i>	2.2	2.1	15	5	80	60	C
<i>Phoma herbarum</i>	2.3	5.5	90	95	90	100	A
<i>Rhizopus stolonifer</i>	2.2	1.0	5	2	100	-	C

* Complete leaves become Yellow and fall down (a) = Average of three methods- (i) Blotter, (ii) seed + spore suspension + soil, (iii) soil + spore suspension + seed

** Height of control plants = 10.0 cm

Disease index (overall assessment)
 A = Most effective
 B = Slightly higher
 C = Moderate
 D = Less effective

macrosporus. At this stage, it was difficult to interpret the actual relationship of the treatment and reduction in the height of the plants. In addition, the above mentioned fungi also caused significant damage to the leaves and mortality of the plants. Most of the fungi tested also inflicted 75-100% inhibition in seed germination and seedling mortality. *Aspergillus niger* and *Rhizopus stolonifer* failed to cause sufficient damage to the leaves, but caused significant inhibition of seed germination, indicating their seed borne nature.

Culture filtrates of *F. oxysporum* and *F. moniliforme* caused drooping, yellowing and wilting of leaves and stem tips. Collapse of

vascular elements in proximal portion of the leaves was also seen. Wilted plants in incipient stage when placed in distilled water, failed to recover, indicating irreversible nature of wilting. The culture filtrate was also toxic to the seeds as it caused significant inhibition of seed germination. However, the chemical nature of the culture filtrate is yet to be determined.

The present findings reveal that some of the fungi tested possess considerable potentiality and needs thorough investigations to understand their disease cycle, specificity and safety towards non-target organisms which are underway in this laboratory.

ACKNOWLEDGEMENTS

The authors are grateful to Prof. G.P. Agarwal, Emeritus Scientist (CSIR), Department of Biological Science, R.D. University, Jabalpur for encouragement. Financial help provided by Madhya Pradesh Council of Science and Technology, Bhopal (B/56/89) is thankfully acknowledged.

REFERENCES

- AGARWAL, G.P. and HASIJA, S.K. 1986. "Micro-organisms in the Laboratory". A laboratory guide for mycology, microbiology and plant pathology". Print House (India), Lucknow, 155 pp.
- CONWAY, K.E. 1979. Evaluation of *Cercospora rodmanii* as a biological control agent of waterhyacinth. *Phytopathol.*, **66**, 914-917.
- JOSHI, N.C. 1987. "Manual of the weed control." Research Co. publications, New Delhi, 545 pp.
- KANCHAN, S.D. 1975. Growth inhibitors from *Parthenium hysterophorus* L. *Curr. Sci.*, **44**, 358.
- MORTENSEN, K. 1985. A proposal for a standardized scale of attack and its application to biocontrol agents of weeds in laboratory screening test. *Proc. VI Int. Symp. Contr. Weeds.* (Delfosse, E.S. ed.). 19-25, August 1984, Vancouver, Canada, *Agric. Can.* pp. 643-50.
- MORTENSEN, K. and HSIAO, A.I. 1987. Fungal infestation of seeds from seven populations of wild oats (*Avena fatua* L.) with different dormancy and viability characteristics. *Weed Res.*, **27**, 297-304.
- RAJAK, R.C. SUNITA FARAKYA, HASIJA, S.K. and PANDEY, A.K. 1990. Fungi associated with congress weed. *Proc. Nat. Acad. Sci. India*, **60**, 165-168.
- SATI, K.C. and GREWAL, J.S. 1981. A technique for seedling inoculation with *Fusaria* associated with Gram (*Cicer arietinum*). *Indian. J. Mycol. Pl. Pathol.*, **11**, 275-277.
- SCHEFFER, R.P. and WALKER, J.C. 1953. The physiology of *Fusarium* wilt of tomato. *Phytopathol.*, **43**, 116-125.
- SUBBA RAO, P.V., MANGALA, A., TOWERS, G.H.N. and RODRIQUEZ, E. 1978. Immunological activity of Parthenin and its diosteriommet in person sensitised by *Parthenium hysterophorus* L. *Contact Dermatitis*, **4**, 199-203.
- TEMPLETON, G.E. 1990. Weed control with Pathogens : Future need and directions. In "Microbes and Microbial Products as herbicides" (R.E. Hoagland ed.), ACS Symposium series 439, American Chemical Society, Washington, pp. 320-328.
- TEMPLETON, G.E. and HEINY, D.K. 1990. Mycoherbicides. In "New Direction in Biological Control : Alternatives for suppressing Agricultural pests and diseases" (R. Baker and P. Dunn, eds). A.R. Liss, New York, pp. 279-286.
- TOWERS, G.H.N., MITCHELL, J.C. RODRIQUEZ, E., BENNETT, F.D. and SUBBA RAO, P.V. 1977. Biology and Chemistry of *Parthenium hysterophorus* L. A problem weed in India. *J. Scient. Ind. Res.*, **36**, 672-684.
- VAN DYKE, C.G. 1991. Biological control of weeds with fungi. In "Handbook of Applied Mycology" Vol. I (D.K. Arora, Bharat Rai, K.G. Mukerjee and G.R. Knudson, eds). Marcel Dekker, Inc., New York, pp. 357-376.