



Cocos nucifera Linn.: A Promising Candidate for Drug Development

Bijesh Vatakkeel^{1*}, R. Mythreyi¹ and E. N. Siju²

¹Chettinad School of Pharmaceutical Sciences, Chettinad Hospital and Research Institute, Chettinad Academy of Research and Education, Kelambakkam, Chennai - 603103, Tamil Nadu, India; vadakkeel@gmail.com ²Department of Pharmacology, College of Pharmaceutical Science, Government Medical College, Kannur - 670503, Kerala, India

Abstract

Cocos nucifera Linn. (Arecaceae) is a common palm found in different parts of the world and is consumed for various purposes, especially food and oil. It is widely used as a nutraceutical globally. *Cocos nucifera* has a wide range of conventional medical applications in inflammation, protozoal and microbial infection, cancer, hepatic illness, diabetes, mellitus, pain, epilepsy, fungal infection, and depression. Different parts of *C. nucifera* like leaves, flowers, roots, oil, and cotyledon were explored for scientific authentication in various illnesses. The presence of phytochemicals like amino acids (lauric acid, arginine, caprylic acid, linoleic acid and palmitic acid), vitamin C, saponins, phenols, terpenoids, phenolics, and tannins were reported in *C. nucifera* and this knowledge further hint for hidden pharmacological activities. This paper presents a review of the phytopharmacological activities of *C. nucifera* so that it may be used for developing a promising herbal candidate for the drug discovery process.

Keywords: Antidiabetic, Antioxidant, Cocos nucifera, DPPH, MTT

1. Introduction

Cocos nucifera Linn. (Arecaceae) is mainly cultivated in tropical areas like India, Africa, and America for edible coconut fruit¹. The *C. nucifera* is also known as the "tree of life" or "tree of heaven" and nature's greatest gift to man as every part of the plant has value for the community's consumption².

The various parts of the *Cocos nucifera* plant like leaves, flowers, oil, endocarp, and roots were used traditionally in folkloric medicine by the diverse communities of the world. In traditional medicine, the fruits of the *C. nucifera* tree have long been used as painkillers, antibacterial, parasiticides, antineoplastic, and cardiotonic³. The tender coconut water and coconut kernel have a variety of medicinal characteristics, including antibiotic, fungicidal, antiviral, anthelmintic, antidermatophytic, antioxidant, hypoglycemic, hepatoprotective, and immunomodulatory⁴. Coconut milk is used for the treatment of illnesses like hematemesis, gallstones, and urinary issues. Coconut oil has been used as a burn wound remedy⁵. Additionally, coconut oil supports the immunological system in people. They are found to diminish the proinflammatory cytokines^{6,7}.

Numerous scientific studies were carried out in the different parts of *Cocos nucifera* to standardize and validate the claims of folkloric knowledge and these documented research outputs are to be compiled in an updated manner so that further studies can be promoted and a promising new chemical entity can be developed.

1.1 Taxonomical Classification

Domain: Eukaryota Kingdom: Plantae Phylum: Spermatophyta Subphylum: Angiospermae Class: Monocotyledonae Order: Arecales

^{*}Author for correspondence

Family: Arecaceae Genus: *Cocos* Species: *Cocos nucifera*

1.2 Synonyms

English (Coconut); Hindi (*Nariyal*) Bengali (*Nariel*); French (*Coco, Noix de coco*); German (*kokospalme*); Italian (*cocco*); Malayalam (*keram*); Lao (*Sino-Tibetan, Phaawz*); Tamil (*tennai-maram*), Malay (*Kelapa*); Mandinka (*Tubabsibo,coc*); Spanish (*cocotero,coco de agua, coco*); Swahili (*Mnazi*); Thai (*ma phrao*); Trade name (coconut); Manipuri (*Yubi*); Marathi (*Naral*).

1.3 Microscopy

The microscopical analysis of the roots of *C. nucifera* root demonstrated the presence of a large inner and outer cortex, as well as a solid round stele. The surrounding compact parenchymatous zone of the cortex is differentiated. In the center, the stele is arranged in a circular shape. The air chambers, metaxylem component, external cortex, pith, phloem, and partition filaments are characteristic of a thin root. A bundle of xylem elements and fibres with black inclusions, xylem elements with annular thickenings, and xylem components with spiral and scalariform thickening are powder characteristics of the root⁸.

1.4 Botanical Description

Cocos nucifera is a slender palm tree distributed in the tropical region. It has a cylindrically shaped trunk with a brown or brownish-grey-coloured surface. The surface of the trunk has circular leaf scars. The leaves of *C. nucifera* are pinnate and feather-shaped with a length of 6-8 meters. The roots are fasciculate, smooth surfaced and capped with a dark brown root cap. The flowers appear as branched clusters located at the base of the leaves. The spadix of the inflorescence is stout and erect, enclosed by the tough spathe. The axis is branched, with the branches bearing sessile flowers, both staminate and pistillate. The fruits are drupe and ovoid shaped. The fruits consist of brown fibrous bark and hard shell. The central cavity consists of coconut water.

1.5 Phytochemistry of Cocos nucifera Linn.

The literature review on the phytochemistry (Figure 1) of different parts of *C. nucifera* is given below:

1.5.1 Coconut Cotyledons

The methanolic extract contains carbohydrate and chloroform extract contain lipids as primary metabolites. The secondary metabolites present in cotyledons are cardiac glycosides, phenolics, and tannins⁹.

1.5.2 Endosperm

The endosperm of *C. nucifera* contains carotenoids, tannins, flavonoids, alkaloids, steroids, phenols, saponins, and trace amounts of glycosides¹⁰.

1.5.3 Inflorescence

The phytochemical analysis of *C. nucifera* flowers indicated that steroids, alkaloids, flavonoids, phenolic substances, and tannins were present¹.

1.5.4 Coconut Shell

Alkaloids, steroids, flavonoids, glycosides, saponins, tannins, and phenols were discovered in the crude extracts¹¹.

1.5.5 Coconut Oil

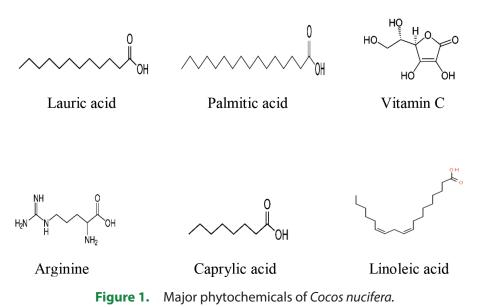
The triglycerides present in coconut oil are exclusively short-chain and medium-chain saturated fatty acids as well as unsaturated fatty acids. The main mediumchain fatty acid in coconut oil is lauric acid. Caprylic acid, capric acid, caproic acid, and myristic acid are additional medium-chain saturated fatty acids that are present. Palmitic acid and stearic acid are two examples of long-chain fatty acids. Linoleic acid, linolenic acid, arachidonic acid, and eicosanoic acid are among the unsaturated fatty acids present¹².

1.5.6 Leaf

The coconut leaf methanolic extract underwent phytochemical analysis, which revealed that it contained considerable amounts of flavonoids, phenols, saponins, terpenoids, and triterpenes. A small amount of tannins, alkaloids, and glycosides are also found¹³.

1.5.7 Liquid Albumen

Vitamin C, nicotinic acid, pantothenic acid, biotin, riboflavin, folic acid, and trace amounts of vitamins B1, B6, and C are also present. Other nutrients include amino acids, L-arginine, plant hormones (auxin, 1,3-diphenylurea, and cytokinin), and enzymes



(acid phosphatase, catalase, dehydrogenase, diastase, peroxidase¹.

1.5.8 Coconut Shell

Aqueous distillate from the shell powder of *C. nucifera* Linn. was examined, and several compounds were found, including dodecanoic acid, tetradecanoic acid, pentadecanoic acid, hexadecanoic acid, and squalene.

2. Pharmacological Studies of *Cocos nucifera* Linn.

The researchers subjected various parts of the *C. nucifera* to confirm the traditional medicinal uses. They can be summarized as below.

2.1 Antioxidant and Antimicrobial Activity

The antioxidant activity of the ethanolic extract, dry distilled extract, and aqueous extracts of the *C. nucifera* endocarp was investigated by Rajeev KS *et al.*, using a DPPH radical scavenging assay. They used the Agar disc diffusion method to examine the antibacterial and antifungal properties of extracts. The research revealed *C. nucifera*'s promising anti-inflammatory, anti-bacterial, and antifungal properties. All of the *Trichophyton* species - *Trichophyton mentagrophytes*, *Trichophyton tonsurans*, *Trichophyton rubrum*, and Microsporum species like *Microsporum canis*, *M. audouinii* and *T. violaceum* are susceptible to the antifungal properties of the alcohol extract¹⁴. Jose M

et al., evaluated the antimicrobial properties of *C. nucifera* (coconut) huskagainst common oral pathogens. The antibacterial activity of the alcohol-based *C. nucifera* extract was significantly concentration-dependent. The inhibitory effect was more severe with Candida and the majority of cariogenic organisms, with a zone of inhibition spanning from 4.6mm to 16.3mm¹⁵.

In 2021, Uche Chidiebere evaluated the antibacterial activity of the ethanolic extract of C. nucifera (coconut) husk ash against a variety of harmful bacteria, including Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli, as well as their capacity to treat wounds in experimental rats. On albino rats, an excision wound model was used to assess the healing capacity of the ointment made from ethanolic extract of C. nucifera husk. The antibacterial study was performed using the agar well diffusion method. The findings demonstrated that topical route administration of extract made from coconut husk ash blended into an ointment base accelerated wound healing, specifically increasing epithelialization in treatment groups¹⁶. The antioxidant activity of protein fractions was assessed by Li Y et al¹⁷. The protein fractions of coconut cake were examined for their characterisation, nutritional value, and antioxidant activity. Prolamin, glutelin-1, and glutelin-2 all possessed significant reducing power and good radical scavenging activity, and globulin and prolamin both exhibited significant ion chelating properties. Additionally, every fraction except glutelin-2 could successfully shield DNA from oxidative damage¹⁷. Udaya Prakash Nyayiru Kannaian *et al.*, studied the antioxidant activities of *C. nucifera* using DPPH, FRAP, FTC, TBA, and Nitric oxide radical scavenging assays. High content of secondary metabolites was extracted by methanolic extraction using cold percolation, and there was also noticeable antioxidant activity¹⁸. The antibacterial activity of the mesocarp of *C. nucifera* was assessed by Indira Bairy KS *et al*²⁶. The fresh mesocarp of an unripe coconut was processed by filtration and mixing to get the crude aqueous extract. Gram-positive and gram-negative bacteria were used. When tested, it had an antibacterial impact on all gram-positive cocci²⁶.

2.2 Hepatoprotective Activity

Manikantan et al., looked at the anti-hepatotoxic activities of the acetone extract from the inflorescence of coconut. The study's findings showed that the groups pre-treated with the extract exhibited a significant decrease in liver superoxide dismutase levels, reduced glutathione, glutathione-S-transferase, and glutathione peroxidase levels, and also inhibited the rise in serum transferase and alkaline phosphatase levels. Additionally, the extract reduced the increased level of malondialdehyde¹⁹. The preventive effects of alcoholic and aqueous extracts of endocarp against hepatotoxicity induced by paracetamol in rats were investigated by Nishant Singh Katiyar et al²⁰. Both extracts considerably decreased the raised blood levels of liver enzymes caused by paracetamol. Animals given the two extracts were partially or completely protected against histological alterations (steatosis, necrosis, etc.)²⁰.

2.3 Anticancer Activity

Cocos nucifera Linn.'s anticancer efficacy was evaluated by Nicole M Tayler *et al*²¹. This study involved extracting and testing *C. nucifera* against a line of breast cancer cells²¹. The anti-cancer properties of the ethanolic extract of *C. nucifera* flowers were assessed by VM Berlin Grace and Monisha M *et al*²². The MTT test was used to determine the extract's anti-cancer impact on a human lung cancer cell line, and an *in vitro* assay for the degradation of egg albumin was used to determine the extract's anti-inflammatory efficacy. Tannins and phytosterols are present, according to the biochemical assays. The gas chromatography-mass spectrometry examination identified a total of 152 phytochemicals, which also included a few fatty acids, polyphenols, and terpenes as well as various phytosterols. The extract had a 96.15% dose-dependent growth inhibitory effect on the cell line. The discovered secondary metabolites, including eugenol, catechol, stigmasterol, campesterol, and t-butyl hydroquinone, might have exerted these medicinal effects²². The proanthocyanidins soluble in ethyl acetate from the immature inflorescence of *C. nucifera.* were tested for their anticancer property. Compared to tamoxifen, EASPA also had higher cytotoxic action against Hela cells, but PC3 cells showed less cytotoxicity²³.

A brief communication on the anti-neoplastic properties of C. nucifera was written by PR Koscheket et al²⁴. He evaluated the aqueous husk fibre fractions from the different varieties of C. nucifera husks for their in vitro anti-tumour properties. Cytotoxicity against cancer cells was determined by the MTT assay²⁴. The anticancer properties of the ethanolic extract of C. nucifera's tender fruits were investigated by Shrabanti Dev et al²⁵. According to this study, tender fruit ethanolic extract has a significant amount of antioxidant activity. When compared to a carcinogenic control, the extract dramatically reduced the number, size, yield, and burden of the tumours in skin cancer model mice produced by DMBA and Croton oil. Natural antioxidants including GSH, SOD, and catalase were boosted by the extract²⁵.

2.4 Antiparasitic, Anti-malarial Activity

The effectiveness of the fruit of C. nucifera against intestinal parasites in sheep was examined by Oliveira LM et al²⁷. The studies were performed on the ethyl acetate extract that was produced from the liquid of green coconut husk fibre. The in vitro assay was done using Haemonchuscontortus for larval development and egg hatching test. At the highest studied concentrations, extract efficacy was 100% for egg hatching and 99.77% for larval development²⁷. Adebayo JO et al., assessed the antimalarial and toxicological potentials of husk fibre extracts of five Nigerian species of C. nucifera. The results demonstrated that alkaloids, tannins, and flavonoids are present in WAT (West African Tall) ethyl acetate extract fraction (WATEAEF), and were effective against Plasmodium falciparum and Plasmodium berghei²⁸.

The use of Cocos nucifera as an antimalarial treatment in Malaysian indigenous medicine was examined by Al Adhroey AH et al²⁹. The findings showed that the extract had some phytochemical components and was safe to take orally from a toxicological perspective. In all three in vivo assessment experiments, the extract considerably decreased the parasitemia. However, the extract did not substantially prolong the period of survival of the infected mice²⁹. The antiplasmodial efficacy of C. nucifera leaves was examined by Tayler N.M. et al., in mice infected with Plasmodium berghei. Comparing intramuscular administration of a C. nucifera leaf decoction to intraperitoneal, subcutaneous, and intragastric approaches, the results demonstrated that intramuscular administration to mice produced the best effects. The extract has a 48% inhibitory preventive impact, but no curative benefit, according to their findings. Finally, in a 4-day suppressive trial, they discovered that when delivered intramuscularly, the extract can limit the growth of the parasite by up to 54% at sub-toxic levels³⁰. Nicole M Tayler et al., analysed the antiparasitic activity of the C. nucifera L. The effectiveness of C. nucifera husk, leaves, pulp, and milk against Trypanosoma cruzi, Leishmania donovani, and Plasmodium falciparum parasites was examined in this study. The aqueous decoctions of leaves only showed antiplasmodial activity²¹.

2.5 Hypoglycemic Activity

The alpha-amylase inhibitory effects of Cocos nucifera husk were studied by Hamdalat Folake et al³¹. The pancreatic-amylase and lipid peroxidation inhibitory effects of methanolic extract and ethyl acetate fraction were identified. The total phenolic content and antioxidant capacity of the ethanolic fraction were significantly larger than those of the methanolic extract. The ethanolic fraction also demonstrated higher inhibitory effects on alpha-amylase activity and lipid peroxidation following intraperitoneal treatment of the C. nucifera extract to alloxan-induced hyperglycemic rats, decreasing blood glucose levels within 5 days³¹. A 2012 study by Victor Emojevwe examined the anti-diabetic properties of a coconut husk extract. On 21 diabetic rats induced with alloxan. the C. nucifera (coconut) husk was investigated. Blood glucose levels were dramatically lowered after extract consumption in diabetic rats under treatment. The rats that received the extracts (coconut husk tea) and those that received Daonil and Metformin both had the same regeneration, according to histopathological research. Therefore, it was determined that there were considerable hypoglycemic and anti-diabetic benefits of *C. nucifera* (coconut) husk tea in alloxan-induced diabetes. This result is similar to what is produced when Daonil and Metformin are combined, and it may be used as a useful adjunct in the treatment of type 2 diabetes³².

The anti-diabetic and antilipidemic effects of Cocos nut endocarp extract were assessed in a diabetic rat model by Farjana Akter *et al*³³. The endocarp of the *C*. nucifera was extracted using methanol and h-hexane, chloroform, and ethyl acetate fractions were prepared. Thirty diabetic rats induced by streptozotocin were employed in total, divided into six experimental groups. For a total of 14 days, all the animals received treatment. Acute hypoglycemia effects, lipid profiles, changes in body weight, and fasting blood sugar levels were all monitored. Additionally, phytochemical screening and anti-oxidant tests were carried out. The group given ethyl acetate fraction treatment saw a strong hypoglycemic impact³³. On streptozotocin (STZ)-induced diabetic rats, Naskar S et al., looked at the impact of the antidiabetic activity on lipid profiles and the cardioprotective effect of hydro-methanol extract of Cocos nucifera (HECN). In comparison to the diabetic control group, fasting blood glucose levels were significantly lower in the animals treated with HECN. Serum enzyme levels, lipid peroxidation, and antioxidant enzyme levels, as well as cholesterol and triglyceride levels in treated groups, were restored toward normal levels in comparison to diabetic control groups, and the values were significant and comparable with the standard group³⁴.

Saranya *et al.*, studied the biochemical changes in the antidiabetic activity of coconut flowers in diabetic rats. When given orally for 30 days, a *C. nucifera* flower extract reduced blood sugar, glycosylated haemoglobin, uric acid, and creatinine levels in diabetic rats. The abnormally high levels of serum aminotransferases and alkaline phosphatase returned to normal. The extract administration restored the plasma protein levels in the diabetic rats to levels that were close to normal. The extract increased the amount of glycogen present and improved glycogen synthase and glycogen phosphorylase activity.³⁵. Pinto IF *et al.*, evaluated the hypoglycemic activity of *C. nucifera* water in alloxaninduced diabetic rats. The findings demonstrated that rats in the coconut water and diabetic group had increased body weight and maintained blood glucose levels in comparison to the control group, as well as decreased HbA1c levels³⁶.

2.6 Anthelmintic Activity

The liquid extracted from the green coconut's bark (Liquid from Bark of Green Coconut (LBGC)) and its butanol extract were tested for their anthelmintic effects on mouse intestinal nematodes by Costa CT *et al*³⁷. Phytochemical analyses were used to ascertain the chemical makeup of the LBGC and its butanol extract. The effectiveness of a 1000 mg/kg dosage of butanol extract in lowering the mouse worm load was 90.7%. Triterpenes, saponins and condensed tannins were detected by phytochemical testing in the LBGC and butanol extracts³⁷.

2.7 Antiepileptic Activity

The antiepileptic effectiveness of *Cocos nucifera* flowers against experimentally induced convulsions in rats was assessed by Archana B *et al*³⁸. Pentylene Tetrazole (PTZ) and Maximal Electroshocks (MES) induced models were used for the investigation. Pretreatment of animals with *C. nucifera* flower extract against MES and Pentylenetetrazol-induced convulsion models showed considerable anticonvulsant efficacy³⁸.

2.8 Antidepressant Activity

Lima EBC *et al.*, studied the antidepressant activity of *Cocos nucifera*. One hour following the final dose of medicine, the mice underwent the Forced Swimming Test (FST), Tail Suspension Test (TST) and Open Field Test (OFT). Immediately following the behavioural tests, the brains of the animals were taken for neurochemical analysis. The findings revealed that hydroalcoholic extract had an antidepressant-like effect by reducing the immobility duration during the FST and TST examinations. The extract treatment caused levels of malondialdehyde to drop. Additionally, nitrite levels in all doses were reduced by the administration³⁹.

2.9 Analgesic Activity

Alviano *et al.*, showed that *Cocos nucifera* aqueous husk fibre extract had analgesic properties. The acetic

acid-induced writhing response in mice was prevented when *C. nucifera* aqueous extract was taken orally. Treatment of rats with this plant extract (dose of 200 mg/kg) caused attenuation in response to the heat stimulation, as shown by tail flick and hot plate assays⁴⁰.

2.10 Peptic Ulcer

Nneli *et al.*, examined the antiulcerogenic properties of coconut extract in rats. The outcomes demonstrated that via macroscopic inspection, coconut milk exhibited protective effects on the inflamed stomach mucosa comparable to those of conventional sucralfate⁴¹.

2.11 Anti-inflammatory Activity

The anti-inflammatory and antinociceptive properties of *Cocos nucifera* Linn. were examined by Sebastian Rinaldi *et al.*, utilising the acetic acid-induced abdominal writhing, Eddy's hot plate, and Rat paw oedema. The findings supported the widespread use of *C. nucifera* in treating several inflammatory illnesses by showing that both the plant and its fractions possess antinociceptive and anti-inflammatory properties⁴². The anti-inflammatory ethanolic extract of *C. nucifera* flowers was assessed by VM Berlin Grace and Monisha M *et al*²². An *in vitro* egg albumin degradation assay was used to assess the extract's anti-inflammatory effect. The extract at 1000g/ml and the 4000g/ml treatment exhibited an effective inhibition of protein breakdown (>50%) as a sign of anti-inflammatory activity²².

2.12 Anti-hypertensive Activity

The anti-hypertensive potential of coconut (*Cocos nucifera* L.) water was assessed by AI Airaodion *et al.*, in Wistar rats. When comparing animals fed a diet without coconut water treatment to those who were fed a diet and treated with coconut water, or to the control group, respectively, in response to heat stimulation, a substantial rise in blood pressure was seen⁴³.

2.13 Nephroprotective Activity

Famurewa *et al.*, investigated the nephroprotective activity of virgin coconut oil on nephrotoxicity produced by diclofenac in rats. In the study, diclofenac elevated serum urea and creatinine, renal tumour necrosis factor- α and malondialdehyde levels markedly increased and decreased the renal glutathione peroxidase, catalase, and superoxide dismutase. Histopathological alterations were also seen. Virgin coconut oil pretreatment attenuated histological changes of renal damage and restored antioxidant enzyme activity and TNF- α levels in the kidney⁴⁴.

3. Conclusion

It is concluded that researchers from around the world conducted scientific pharmacological evaluations of *Cocos nucifera* to confirm the traditional medicinal uses such as anti-oxidant, anti-inflammatory, anti-protozoal, anti-microbial, anti-cancer, hepatoprotective, antidiabetic, analgesic, anti-epileptic, anti-fungal, and antidepressant, and others. These documented scientific studies may aid in the development of safer herbal chemical moiety in drug discovery.

4. References

- Lima EB, Sousa CN, Meneses LN, Ximenes NC, Santos Júnior MA, Vasconcelos GS, Lima NB, Patrocínio MC, Macedo D, Vasconcelos SM. *Cocos nucifera* (L.) (Arecaceae): A phytochemical and pharmacological review. Brazilian Journal of Medical and Biological Research. 2015; 48(11):953-64. https://doi.org/10.1590/1414-431x20154773 PMid:26292222 PMCid: PMC4671521
- 2. Dyana JP and Kanchana G. Preliminary phytochemical screening of *Cocos nucifera* L. flowers. International Journal of Current Pharmaceutical Research. 2012; 4 (3):62-3.
- 3. Akter F, Bhuiya NMM, Hasan N, Rokeya B, Raihan S, Al MZ. *Cocos nucifera* endocarp extract exhibits anti-diabetic and anti-lipidemic activities in a diabetic rat model. International Journal of Scientific Research in Dental and Medical Sciences. 2022; 4(1):8-15. https://doi.org/10.30485/ ijsrdms.2022.320342.1228
- Lima EBC, Sousa CNS, Meneses LN, Ximenes NC, Santos, Júnior MA, Vasconcelos G.S, Lima NBC, Patrocínio MCA, Macedo D, Vasconcelos SMM. *Cocos nucifera* (L.) (Arecaceae): A phytochemical and pharmacological review. Brazilian Journal of Medicine and Biological Research. 2015; 48(11):953-64. https://doi.org/10.1590/1414-431x20154773 PMid: 26292222 PMCid: PMC4671521
- 5. Chaichit C. Thai herbs and herbal products. Bangkok, Srimuang Printing Co. 2004.
- Wan JM, Grimble RF. Effect of dietary linoleate content on the metabolic response of rats to *Escherichia coli* endotoxin. Clinical Science. 1987; 72:383-85. https://doi.org/10.1042/ cs0720383 PMid:3545648

- Sadeghi S, Wallace FA, Calder PC. Dietary lipids modify the cytokine response to bacterial lipopolysaccharide in mice. Immunology. 1999; 96:404-10 https://doi. org/10.1046/j.1365-2567.1999.00701.x PMid:10233721 PMCid: PMC2326770
- Dhumansure R, Biradar SK, Nanjwade B, Patil MH, Sirse KK. Microscopical and phytochemical investigation of root *Cocos nucifera* Linn. International Research Journal of Pharmacy. 2012; 3(7):285-87.
- Nyayiru KUP, Edwin JB, Rajagopal V, Nannu Shankar S, Srinivasan B. Phytochemical composition and anti-oxidant activity of coconut cotyledon. Heliyon. 2020; 13:6(2): e03411. https://doi.org/10.1016/j.heliyon.2020.e03411 PMid: 32083218 PMCid: PMC7021540
- Offor CE, Uzuegbu UE, Opajobi AO, Ugwu Okechukwu PC, Uneke FC, Abara PN, and Orinya O. Proximate composition and phytochemical analysis of the endosperm of *Cocos nucifera* Linn. IDOSR Journal of Scientific Research. 2017; 2(1):83-90.
- Kendeson AC, Iloka SG, Abdulkadir AG, Ushie OA, Abdu Z, Jibril S, John ST. Phytochemical screening, antimicrobial and elemental analyses of crude extracts from *Cocos nucifera* (Coconut) shell. Dutse Journal of Pure and Applied Sciences. 2019; 5(1b):237-243.
- Obidoa, Onyechi, Joshua, Parker Elijah, and Eze, Nkechi J. Phytochemical analyses of *Cocos nucifera* Linn. Archives of Pharmaceutical Science. 2009; 1(1):87-96.
- MD Shahidul Islam. Phytochemical screening and *in vitro* anti-oxidant and anti-inflammatory potential evaluations of methanolic extracts of *Cocos nucifera* (L.) leaves. Scientific Journal of Environmental Sciences. 2019; (2):24-9. https:// europub.co.uk/articles/-A-574283
- Rajeev KS, Nitesh J, Varadaraj BG, Hitesh J. Anti-oxidant and anti-microbial activities of *Cocos nucifera* Linn. (Arecaceae) endocarp extracts. Indo Global Journal of Pharmaceutical Sciences. 2011; 1(4):354-61 https://doi. org/10.35652/IGJPS.2011.34
- Jose M, Cyriac MB, Pai V, Varghese I, Shantaram M. Antimicrobial properties of *Cocos nucifera* (coconut) husk: An extrapolation to oral health. Journal of Natural Science, Biology and Medicine. 2014; 5(2):359-64. https://doi. org/10.4103/0976-9668.136184 PMid:25097415 PMCid: PMC4121915
- Uche C. Evaluation of anti-bacterial and wound healing activity of *Cocos nucifera* (coconut) husk ash extract in mice. Journal of Food Processing and Technology. 2021; 12(4):1000878.
- Li Y, Zheng Y, Zhang Y, Xu J, Gao G. Anti-oxidant activity of coconut (*Cocos nucifera* L.) protein fractions. Molecules. 2018; 23(3):707. https://doi.org/10.3390/ molecules23030707 PMid:29558429 PMCid: PMC6017440

- Udaya PNK, Jasmine BE, Vidhya R, Sripriya NS, Bhuvaneswari S. Phytochemical composition and antioxidant activity of coconut cotyledon. Heliyon. 2020; 6(2): e03411. https://doi.org/10.1016/j.heliyon.2020.e03411. PMid:32083218 PMCid: PMC7021540
- Manikantan AC, Thadiyan PI, Harsha K, Rajeev KS, Kainoor KJ, Palpu P, Varughese G. *Cocos nucifera* L. inflorescence extract: An effective hepatoprotective agent. Indian Journal of Traditional Knowledge. 2020; 19(1):128-36 https://doi.org/10.56042/ijtk.v19i1.30852
- Nishant SK, Rajesh A. Protective effect of extracts of *Cocos nucifera* endocarp on paracetamol induced hepatotoxicity in rats. Asian Journal of Pharmacy and Pharmacology. 2019; 5(6):1162-66. https://doi.org/10.31024/ajpp.2019.5.6.12
- Nicole MT, Cristopher AB, Liuris H, Jamie M, Michelle NG, Laura P, Alejandro A, Sara R, Lorena MC, Ricardo C. Analysis of the anti-parasitic and anti-cancer activity of the coconut palm (*Cocos nucifera* L. Arecaceae) from the natural reserve of Punta Patiño, Darién. P One. 2019; 14(4):e0214193. https://doi.org/10.1371/journal. pone.0214193 PMid:30939131 PMCid: PMC6445518
- 22. Berlin Grace VM and Monisha M. *In vitro* evaluation of anti-inflammatory and anti-cancer activities of the *Cocos nucifera* flower extract and the phytochemical identification by gas chromatography/mass spectrometry analysis. Indian Journal of Pharmaceutical Science. 2022; 84(2):348-57. https://doi.org/10.36468/pharmaceutical-sciences.928
- Padumadasa C, Dharmadana D, Abeyskera A, Thammitiyagodage M. *In vitro* anti-oxidant, antiinflammatory and anti-cancer activities of ethyl acetate soluble proanthocyanidins of the inflorescence of *Cocos nucifera* L. BMC Complementary and Alternative Medicine. 2016; 16(1):345. https://doi.org/10.1186/s12906-016-1335-2. PMid:27595601 PMCid: PMC5011879
- 24. Koschek PR, Alviano DS, Alviano CS, Gattass CR. The husk fibre of *Cocos nucifera* L. (Palmae) is a source of anti-neoplastic activity. Brazilian Journal of Medicine and Biological Research. 2007; 1339-343. https://doi. org/10.1590/S0100-879X2006005000153 PMid:17713650
- 25. Shrabanti D, Anha AS, Archana M, Projit RG, Kaniz A, Md. Abdullah AB, Masum S, Asish KD. Evaluation of antioxidant and chemopreventive effect of *Cocos nucifera* Linn. Journal of Biological Science. 2017; 6(2):47-58. https://doi. org/10.3329/jujbs.v6i2.36590
- Bairy I, Bhat KS and Shivananda PG. Anti-bacterial activity of shell fibres of *Cocos nucifera* Linn. On oral microflora. Indian Drugs.1997; 34(3):143-45
- Oliveira LM, Bevilaqua CM, Costa CT, Macedo IT, Barros RS, Rodrigues AC, Camurça-Vasconcelos AL, Morais SM, Lima YC, Vieira LS, Navarro AM. Anthelmintic activity of *Cocos nucifera* L. against sheep gastrointestinal nematodes. Veterinary Parasitology. 2009; 159(1):55-9. https://doi. org/10.1016/j.vetpar.2008.10.018 PMid:19042088

- Adebayo JO, Balogun EA, Malomo SO, Soladoye AO, Olatunji LA, Kolawole OM, Oguntoye OS, Babatunde AS, Akinola OB, Aguiar AC, Andrade IM, Souza NB, Krettli AU. Anti-malarial activity of *Cocos nucifera* husk fibre: further studies. Evidence-Based Complementary Alternative Medicine. 2013; 742476. https://doi.org/10.1155/2013/742476 PMid:23983800 PMCid: PMC3745858
- 29. Al-Adhroey AH, Nor ZM, Al-Mekhlafi HM, Amran AA, Mahmud R. Evaluation of the use of *Cocos nucifera* as an anti-malarial remedy in Malaysian folk medicine. Journal of Ethnopharmacology. 2011; 134(3):988-91. https://doi. org/10.10 16/j.jep.2011.01.026 PMid:21277969
- Tayler NM, De Jesus R, Spadafora R. Anti-plasmodial activity of *Cocos nucifera* leaves in Plasmodium bergheiinfected mice. Journal of Parasitic Disease. 2020; 44(2):305-13. https://doi.org/10.1007/s12639-020-01207-7 PMid: 32499668 PMCid: PMC7244650
- 31. Hamdalat FM, Jubril OA, Sarah AA, Azeemat TA, Raliat AA, Abdulkabir BB. Anti-oxidant and alpha-amylase inhibitory potentials of *Cocos nucifera* husk. Food Science and Nutrition. 2018; 6:1676-83. https://doi.org/10.1002/ fsn3.741 PMid: 30258612 PMCid: PMC6145255
- Victor Emojevwe. Anti-diabetic effects Of the Cocos nucifera (Coconut) husk extract. Journal of Medical and Applied Biosciences. 2012; 4:16-19 https://doi.org/10.13140/ RG.2.2.14402.27844
- 33. Farjana Akter NM. Mahmudul AB, Nazmul H, Begum R, Sheikh Z R, Zobaer A M. *Cocos nucifera* endocarp extract exhibits anti-diabetic and anti-lipidemic activities in diabetic rat model. International Journal of Scientific Research in Dental and Medical Sciences. 2022; 4(1):8-15.
- Naskar S, Mazumder UK, Pramanik G, Gupta M, Kumar RB, Bala A, Islam A. Evaluation of anti-hyperglycemic activity of *Cocos nucifera* Linn. streptozotocin-induced type 2 diabetic rats. Journal of Ethnopharmacology. 2011; 138(3):769-73. https://doi.org/10.1016/j.jep.2011.10.021 PMid:22041106
- 35. Saranya S, Pradeepa S, Subrahmanian S. Biochemical evaluation of the anti-diabetic activity of *Cocos nucifera* flowers in STZ-induced diabetic rats. International Journal of Pharmaceutical Sciences Review and Research. 2014; 26(1):67-75.
- 36. Pinto IF, Silva RP, Chaves FAB, Dantas LS, Bispo VS, Matos IA, Otsuka FA, Santos AC, Matos HR. Study of antiglycation, hypoglycemic, and nephroprotective activities of the green dwarf variety coconut water (*Cocos nucifera* L.) in alloxan-induced diabetic rats. Journal of Medicinal Food. 2015; 18(7):802-9. https://doi.org/10.1089/jmf.2014.0046 PMid: 25651375
- Costa CT, Bevilaqua CM, Morais SM, Camurça-Vasconcelos AL, Maciel MV, Braga RR, Oliveira LM. Anthelmintic activity of *Cocos nucifera* L. on intestinal nematodes of mice. Research in Veterinary Science. 2010; 88(1): 101-3. https://doi.org/10.1016/j.rvsc.2009.05.008 PMid: 19505703

- Archana B, Ravi NM, Vinay E, Ravali N, Paka SK. Evaluation of antiepileptic activity of flowers of *Cocos nucifera* L. against experimentally induced convulsions in rats. Journal of Drug Delivery and Therapeutics. 2021; 11(6):159-66. https://doi.org/10.22270/jddt.v11i6.5097
- Lima EBC, De Sousa CNS, Vasconcelos GS. Antidepressant, antioxidant and neurotrophic properties of the standardized extract of *Cocos nucifera* husk fibre in mice. Journal of Natural Medicine. 2016; 70(3):510-21. https://doi.org/10.1007/ s11418-016-0970-8 PMid:26857134
- Alviano, Daniela. Antinociceptive and free radical scavenging activities of *Cocos nucifera* L. (Palmae) husk fibre aqueous extract. Journal of Ethnopharmacology. 2004; 92:269-73. https://doi.org/10.1016/j.jep.2004.03.013 PMid:15138011
- Nneli RO and Woyike OA. Antiulcerogenic effects of coconut (*Cocos nucifera*) extract in rats. Phytotherapy research. 2008; 22(7):970-72. https://doi.org/10.1002/ ptr.2318. PMid: 18521965

- Sebastian R, Davi OS, Fabiana B, Celuta S. Alviano, Daniela S. Alviano, Maria EM, Patricia DF. Characterization of the antinociceptive and anti-inflammatory activities from *Cocos nucifera* L. (Palmae), Journal of Ethnopharmacology. 2009; 122(3):541-46. https://doi.org/10.1016/j.jep.2009.01.024 PMid:19429325
- 43. Airaodion AI, Ekenjoku JA, Megwas AU, Ngwogu KO, and Ngwogu AC. The Antihypertensive potential of coconut (*Cocos nucifera* L.) water in Wistar rats. Asian Journal of Research in Cardiovascular Diseases. 2019; 1(1):1-8.
- 44. Famurewa AC, Akunna GG, Nwafor J, Chukwu OC, Ekeleme Egedigwe CA, Oluniran JN. The nephroprotective activity of virgin coconut oil on diclofenac-induced oxidative nephrotoxicity is associated with antioxidant and anti-inflammatory effects in rats. Avicenna J Phytomed. 2020; 10(3):316-24. PMID: 32523886; PMCID: PMC7256280.