Phytochemical Characterization, *In-Vitro* Cytotoxic and Antibacterial Activity of *Cotula cinerea* (Delile) Vis Essential Oil

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Abstract

Cotula cinerea is a traditional Algerian medicinal plant that has many biological properties, such as analgesic, antiseptic and antibacterial activities. Essential oil of Cotula cinerea was subjected to chromatographical and spectroscopical studies to determine the chemical composition. The analysis highlighted that the oil contains trans thujone (51.86%), santalina triéne (10.6%), α - pinéne (2.02%), sabinene (6.17%), cineole <1.8> (5.34%), δ - terpinene (1.57%), camphor (2.63%), β - terpineol (1.39 %) and terpin-4-ol (1.73%) as the major constituents. The essential oil was tested for antibacterial activity. The oil exhibited substantial antibacterial activity against both the tested Gram negative and Gram positive bacteria. In search of better anticancer agent the essential oil of Cotula cinerea was also subjected for *in-vitro* cytotoxic activity on two cancerous human cell lines; colon (HCT116) and hepatic (HePG2) cancer cell lines. The results indicates that the essential oil of Cotula cinerea has a significant cytotoxic activity on the tested colon cancer cell lines with a 66.9% inhibition and a minimal inhibition (33.9%) on liver cancer cell line.

Keywords: Essential Oil Composition, GC/MS, HCT116, HePG2

1. Introduction

For better knowing the phytochemical and microbiological properties of the plants of "El-Oued" region, *Cotula cinerea* [Syn. *Brocchia cinerea* (Del) Vis], locally known as (Shihia/Shihit El Ebel)¹, belonging to the family "Asteraceae" was chosen; as it is among the plants mostly used by the local population for TS medicinal properties. *Cotula cinerea* is used against insolation, colic, cough and bronchopulmonary cooling. This species is widely used in traditional Moroccan medicine for its biological properties such as anti-inflammatory, analgesic, antiseptic, antibacterial, antipyretic activities². It is also used as an infusion to facilitate digestion³. The species *Cotula cinerea* is one of three species belonging to the genus *Cotula* (Asteraceae) existing in South-Algeria³. It is a woolly

whitish plant, with thick leaf divided in their upper part to a 3-5 obtuse teeth, stems are 10–40 cm, slept then raised; capitulum from 6 to 10 mm in diameter, woolly involucres with a tubular flower, and brown buds which would become golden yellow^{4,5}. Its stems are diffuse or erect. The leaves and whitish-green stems are covered with tiny hairs thick⁶, velvety small leaves, whole are cut into three to seven teeth or "fingers", and the shaft of high branch yellow inflorescences⁸. The terpenes in the essential oils present are responsible for the characteristic odor⁸. The literature studies highlights that the leaf extracts of *Cotula cinerea* Del are effective against pathogenic fungi, and also have insecticidal activity on the larvae⁹. It finds importance in the management of stomach pain, fever, headache, migraine, cough and joint inflammation¹⁰.

In the present investigation, the chemical components of *Cotula cinerea* Del essential oil was analyzed and screened for antibacterial activity against some pathogenic and food-borne bacteria. The essential oil was also evaluated for anticancer activity against HePG2 (Hepatic) and HCT116 (Colon) human cancerous cell lines.

2. Materials and Methods

2.1 Plant Material

Aerial parts of *Cotula cinerea* Del., were collected from the Hassi Khalifa City in the wilaya of Eloued, north east of Algeria during the period of December 2015 to January 2016 for the investigation. Voucher specimens were deposited to the Herbarium of the Chemistry Laboratory, University of El-Oued.

2.2 Isolation of the Essential Oils

The aerial parts (100 g) were washed, sorted and dried for a month at room temperature, in the shade and then were finely pulverized by using a mill blade. Clevenger-type apparatus was used for extraction, and hydrodistillation was performed for 4 h¹¹. The distilled essential oil was dried over anhydrous sodium sulfate, filtered and stored at 4°C.

2.3 Gas Chromatography - Mass Spectrometry (GC/MS) Analysis Conditions

The oil was analyzed by GC on a Hewlett Packard GC-MS system, Model 6890 equipped with Flame Ionization Detector (FID), HP-5, $30m \times 0.25mm$ ID, 0.25mm film thickness, fused capillary column. The carrier gas was nitrogen (0.8mL/min). Temperature programming was done from $60-250\,^{\circ}$ C at $4\,^{\circ}$ C /min with initial and final hold time of 8 minutes. The injection volume was 0.4 μL neat and Split ratio was 1:20. The percentage of the constituents was calculated by electronic integration of FID peak areas without the use of response factor correction and the sample indices were calculated following Van den Dool and Kratz¹².

The analysis of the volatile constituent was run on a Hewlett-Packard GC-MS system Hewlet Packard 5973 A, equipped with an non polar capillary column HP5MS, $30~m\times0.25$ mm, phase thickness $0.25~\mu m$. The detection of elutent was done using an electron ionization system with ionization current: 70eV. Inert helium gas (99.999%) was used as a carrier gas at a constant flow rate of 0.7 mL/min. The injector temperature was 60 and 280°C, respectively. The temperature programmed for oven was from 2°C/min then held at isothermal condition for 8 min and finally raised to 280°C at 2°C/min, split ratio of 1:20, and volume injected was 0.1 to 0.2 μl of the isolated essential oil 13 .

2.4 Identification of Compounds

The identification of the essential oil constituents was based on a comparison of their retention times to n-alkenes function (C_9 – C_{28})¹⁴. The identification of the oil components were based on MS search Data Library (Wiley & Nist) or with authentic compounds or with the data published in the literature, and Retention Indices (RI) (Adams, 2010)¹⁵. The chromatographic conditions were identical to those used for GC-MS analysis¹⁶.

2.5 Antibacterial Activity

The Minimal Inhibitory Concentration (MIC) of essential oil values were evaluated in the Mueller Hinton Broth (MHB) by dilution method^{15,16} against Bacillus subtilis (ATCC6663), Micrococcus luteus (ATTC9314), Listeria monocytogene (CIP82110), Escherichia coli (CIP54.8), Klebsiella pneumonia (CIP82.91), Pseudomonas aeruginosa (CIPA22), Agrobacterium tumefaciens (N°2410), Salmonella enteric (CIP 81.3). An aliquot (1 mL) of this suspension was transferred to a sterile tubes of MHB containing various concentrations of oils $(0.1-15\mu L)^{18}$ and the volume was adjusted to 10 mL with ethanol (5%, w/v) to obtain with 10 µl bacterial inoculums adjusted a concentration of 106 CFU/ mL^{17,18}. They were incubated under shaking conditions (100-120 rpm) for 24 h at 37 °C^{19,20}.

2.6 In-Vitro Cytotoxic Activity

Cytotoxic activity on human cell line (HePG2–HCT116) was assessed by the mitochondrial dependent reduction of yellow MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) to purple formazan²¹.

3. Results

3.1 Chemical Composition of the Essential Oil

The yield of essential oil isolated by hydrodistillation of *Cotula cinerea* was 0.74% (w/w), based on the dry weight of the sample. Forty one constituents were identified by GC-MS (Figure 1), representing 88% of the oil (Table 1). The major compounds were trans thujone (51.86%), santalina triéne (10.6%), α - pinéne (2.02%), sabinene (6.17%), cineole <1.8> (5.34%), δ - terpinene (1.57%) and camphor (2.63%).

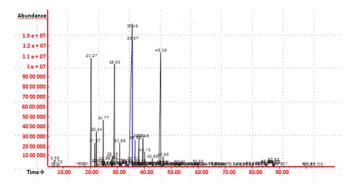


Figure 1. GC-MS of *Cotula cinerea* essential oil.

Table 1. Chemical composition, percentage composition of *Cotula cinerea* essential oil and Retention Indices (IR)

S. No.	IR _a	Compounds	IR _d	%
01	855	cis salven	-	-
02	908	santalina triéne (santolina triene)	914	10.6
03	931	α- thujen	935	0.88
04	939	α-pinene	943	2.02
05	953	camphene	956	0.85
06	976	sabinene	976	6.17
07	980	β- pinene	981	0.58
08	991	cineole dehydro 1.8	988	0.64
09	991	myrcene	993	0.07
10	999	mentha-1(7)-8dien < meta>	997	0.06
11	1018	α - terpinene	1017	0.83
12	1022	ortho cymen	1020	0.43
13	1026	para cymen	1029	0.6
14	1033	cineole <1.8>	1033	5.34
15	1062	δ - terpinene	1057	1.57
16	1068	sabinene hydrate (cis)	1069	0.46
17	1088	terpinolene	1088	0.36
18	1102	cis thujone	1100	0.52
19	1114	trans thujone	1117	51.86
20	1121	menth-2en-1-ol <cis para=""></cis>	-	-
21	1133	thujanol <iso-3-></iso-3->	-	-
22	1143	camphor	1140	2.63
23	1163	β- terpineol	1160	1.39
24	1177	terpin-4-ol	1178	1.73
25	1189	α - terpineol	1190	0.58
26	1193	cis piperitol	-	-
27	1194	myrtenol	1195	0.13
28	1217	trans carveol	-	

S. No.	IR _a	Compounds	IR _d	%
29	1226	dihydro carveol <neo iso=""></neo>	1224	0.53
30	1239	cumin aldehyde	-	-
31	1246	carvotanacetone	1247	0.9
32	1262	chrystanthenyl acetate cis	1264	5.07
33	1273	iso pulegol acetate	1274	0.06
34	1287	cymen-7-ol- <para></para>	1285	0.08
35	1298	carvacrol	-	-
36	1314	decadienal <(E,E)-2,4->	-	-
37	1340	terpin-4-ol acetate	-	-
38	1365	neryl acetate	1367	-
39	1394	cis jasmone	1390	0.15
40	1458	farnesen <(E)- beta->	-	-
41	1480	germacrene D	1481	0.06

IR, Retention Index.

Table 2. MIC of essential oil from Cotula cinerea

Concentration (μL/mL)	0.1	0.2	0.3	0.4	0.5	-	7	m	4	5	10	15
	Gra	m-po:	sitive									
Bacillus subtilis (ATCC6663)	+	+	+	+	+	+	+	+	+	+	_	_
Micrococcus luteus (ATTC9314)	+	+	+	+	+	+	+	+	+	+	+	_
Listeria monocytogene (CIP82110)		+	+	+	+	+	+	+	+	+	_	_
	Grar	n–neg	jative									
Escherichia Coli (CIP54.8)	+	+	+	+	+	+	+	+	+	+	_	_
Klebsiella pneumonia (CIP82.91)	+	+	+	+	+	+	+	+	+	+	_	_
Pseudomonas aeruginosa (CIPA22)	+	+	+	+	+	+	+	+	+	+	_	_
Agrobacterium tumefaciens (N°2410)	+	+	+	+	+	+	+	+	+	+	+	_
Salmonella enterica (CIP 81.3)	+	+	+	+	+	+	+	+	+	+	+	_

Note: (–), total inhibition; (+), growing

3.2 Antibacterial Activity

The antibacterial activity of the *Cotula cinerea* essential oil was tested against three Gram positive and five Gram negative pathogenic bacterial strains. The results of the antibacterial activity are shown in Table 2.

3.3 In-Vitro Cytotoxic Activity

The cytotoxic activity of the *Cotula cinerea* essential oil was tested against two cell line HCT116 and HePG2. The results are as shown in Table 3.

Table 3. Cytotoxic activity of oil of *Cotula cinerea* against cultured different cell

Cell lines	100 μg/mL	50 μg/mL	25 μg/mL	12.5 μg/mL	LC ₉₀ (μg/mL)	LC ₅₀ (μg/mL)	Doxorubicin	DMSO at 100 ppm
HCT116	66.9 + 0.86	21.33 + 1.858	6.0 + 0.65	0	122.3	86.7	37.6	1%
HePG 2	33.9 + 0.56	11.1 + 0.12	0	0	-	21.6	21.6	1%

IR_d Experimentally Determined Retention Indices.

4. Discussion

Djellouli et al.,²² and Sieniawska et al.,²⁸ have reported that trans-thujone (41.4%), cis-verbenyl acetate (24.7%), 1,8-cineole (8.2%) and camphor (5.5%) as the major components of the plants collected in Morocco, while Atef et al.,⁹ have reported that santolina triene (18.58%), thujone (21.73%), 3-carene (30.99%) and camphor (6.21%) as major constituents²⁴. Current study has shown that the oil contains trans thujone (51.86%), santalina triéne (10.6%), α- pinéne (2.02%), sabinene (6.17%), cineole <1.8> (5.34%), δ - terpinene (1.57%), camphor (2.63%), β- terpineol (1.39%) and terpin-4-ol (1.73%) as the major constituents.

The four bacterial strains manifested the same sensitivity vis-à-vis to the *Cotula cinerea* essential oils. They were all inhibited at 15 μ L/mL concentration. As for mold, they were less sensitive than bacteria and their growth was stopped at 10 μ L/mL concentration. The current study is in agreement with study of Boussoula et al²⁵.

The essential oil showed potent cytotoxic activity on HcT116 colon cancer cell line with a 66.9 % inhibition at 100 μ g/mL concentration (LC50 86.7 μ g/mL and LC90 122.3 μ g/mL). The result is very promising compared with positive control doxorubicin 37.6 μ g/mL, while the results indicated week activity on HePG2 (33.9 %) inhibition.

Cotula cinerea has also been reported for anticandidal activity²⁶. Medicinal plants rich with volatile oil represent an important source of antioxidant and anticancer drugs²⁷. The use of essential oil in combination with cancer therapy decreases the side effect of drugs²⁸. Essential oil of Ricinus communis containing thujone and 1.8-cinole has been reported for antiproliferative activity²⁹. Cotula cinerea showed potent cytotoxic activity especially for colon cancer and this may also be corresponding to the chemical composition of essential oil which is rich in trans thujone 50% and santolina triene, as they are reported for anticancer activity.

5. Conclusion

The present study reveals antibacterial potency of essential oil of *Cotula cinerea* of eastern Algeria. *C. cinerea* can also be an inexpensive source of natural antibacterial substance for use in pathogenic systems to prevent the growth of bacteria and extend the shelf life of

the processed foods. *C. cinerea* showed better cytotoxic activity against colon cancer cell line (HCT116) than liver cancer cell line (HePG2).

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7. References

- 1. Abdenbi A, Abdelwahed D, Bouaaza M, Touati B. Screening Phytochimique et Activite Antibacterienne De L'Huile Essentielle De Cotula Cinerea (Gartoufa) Dans La Region De Bechar. Int J Res Eng Technol. 2014; 2(2): 49–54.
- Markouk M, Redwane A, Lazrek HB, Jana M. Antibacterial activity of Cotula cinerea extracts. Fitoterapia. 1999; 70: 314–6. https://doi.org/10.1016/S0367-326X(99)00042-8
- Redwane A, Markouk M, Lazrek HB, Amarouch H, Jana M. Laboratory evaluation of molluscicidal activity of extracts from Cotula cinerea (L) and Quercus lusitania var. infectoria galls (Oliv). Ann Pharm Fr. 1998; 56(6): 274–6. PMid:9872015
- 4. Dendougui H, Seghir S, Jay M, Benayache F, Benayache S. Flavonoids from Cotula cinerea Del. Int J Med Arom Plants. 2012; 2(4): 589–95.
- 5. Bensizerara D, Menasria T, Melouka M, Cheriet L, Chenchouni H. Antimicrobial activity of xerophytic plant (Cotula cinerea Delile) extracts against some pathogenic bacteria and fungi. Asian Pac J Trop Biomed. 2012: 1–5.
- Markouk M, Redwane A, Lazrek HB, Jana M. Antibacterial activity of Cotula cinerea extracts. Fitoterapia. 1999; 70(3): 314–6. https://doi.org/10.1016/S0367-326X(99)00042-8
- 7. Review T. Environmental Science. 2014; Jul: 302-16.
- Metwally MA, El-Dahmy S, Jakupovic J, Bohlmann F, Dawidar AM, Metwally SA. Glaucolide-like sesquiterpene lactones from Cotula cinerea. Phytochemistry. 1985; 25(1): 255–7. https://doi.org/10.1016/S0031-9422(00)94543-6
- Atef C, Boualem M, Cherif M, Youcef H, Azzedine C. Chemical composition and antimicrobial activity of essential oils in xerophytic plant cotula cinerea Del (Asteraceae) during two stages of development: flowering and fruiting. J Appl Pharm Sci. 2015; 5(3): 029–34. https://doi.org/10.7324/JAPS.2015.50306

- Benhammou N, Ghambaza N, Benabdelkader S, Atik-Bekkara F, Kadifkova Panovska T. Phytochemicals and antioxidant properties of extracts from the root and stems of Anabasis articulata. Int Food Res J. 2013; 20(5): 2057–63.
- Yá-ez X, Pinzón ML, Solano F, Sánchez LR. Chemical composition of the essential oil of Psidium caudatum McVaugh. Molecules. 2002; 7(9): 712–6. https://doi. org/10.3390/70900712 PMCid:PMC6146449
- Vandendool H, Kratz PD. A generalization of the retention index system including linear temperature programmed gas-liquid partition chromatography. J Chromatogr. 1963; 11(3): 463–71. https://doi.org/10.1016/S0021-9673(01)80947-X
- 13. Al Abbasy DW, Pathare N, Al-Sabahi JN, Khan SA. Chemical composition and antibacterial activity of essential oil isolated from Omani basil (Ocimum basilicum Linn.). Asian Pacific J Trop Dis. 2015; 5(8): 645–9. https://doi.org/10.1016/S2222-1808(15)60905-7
- 14. Chang LP, Sheng LS, Yang MZ, An DK. Retention index of essential oil in temperature-programmed capillary column gas chromatography. Acta Pharm Sin. 1989; 24(11): 847–52.
- May J, Chan CH, King A, Williams L, French GL. Timekill studies of tea tree oils on clinical isolates. The Journal of antimicrobial chemotherapy. 2000; 45: 639–43. https:// doi.org/10.1093/jac/45.5.639 PMid:10797086
- 16. Sparkman OD. Identification of essential oil components by gas chromatography/quadrupole mass spectroscopy Robert P. Adams. J Am Soc Mass Spectrom. 2005; 16(11): 1902–3. https://doi.org/10.1016/j.jasms.2005.07.008
- 17. Ferreira A, Proenea C, Serralheiro MLM, Araejo MEM. The in vitro screening for acetylcholinesterase inhibition and antioxidant activity of medicinal plants from Portugal. J Ethnopharmacol. 2006; 108(1):31–7. https://doi.org/10.1016/j.jep.2006.04.010 PMid:16737790
- Aligiannis N, Kalpoutzakis E, Mitaku S, Chinou IB. Composition and antimicrobial activity of the essential oils of two Origanum species. J Agric Food Chem. 2001; 49(9): 4168–70. https://doi.org/10.1021/jf001494m PMid: 11559104
- 19. Saadana D, Mahjoub MA, Boussaada O, Chriaa J, Charaif I, Daami M, et al. Chemical composition and antimicrobial activity of volatile compounds of Tamarix boveana (Tamaricaceae). Microbiol Res. 2008; 163(4): 445–55. https://doi.org/10.1016/j.micres.2006.07.009 PMid:17223327
- 20. Hammer KA, Carson CF, Riley T V. Antimicrobial activity of essential oils and other plant extracts. J Appl Microbiol.

- 1999; 86(6): 985–90. https://doi.org/10.1046/j.1365-2672.1999.00780.x PMid:10438227
- 21. El-Sharkawy ER, Matloub AA, Atta EM (2103). Cytotoxicity of new flavonoid compound isolated from Farsetia aegyptia. Int J Pharm Sci Invent. 2013; 2(1): 22–7.
- 22. Djellouli M, Benmehdi H, Mammeri S, Moussaoui A, Ziane L, Hamidi N. Chemical constituents in the essential oil of the endemic plant Cotula cinerea (Del.) from the southwest of Algeria. Asian Pac J Trop Biomed. 2015; 5(10): 870–3. https://doi.org/10.1016/j.apjtb.2015.06.007
- 23. Kasrati A, Alaoui Jamali C, Bekkouche K, Wohlmuth H, Leach D, Abbad A. Comparative evaluation of antioxidant and insecticidal properties of essential oils from five Moroccan aromatic herbs. J Food Sci Technol. 2015; 52(4): 2312–9. https://doi.org/10.1007/s13197-014-1284-z PMid:25829614 PMCid:PMC4375224
- 24. Bensizerara D, Menasria T, Melouka M, Cheriet L, Chenchouni H. Antimicrobial Activity of Xerophytic Plant (Cotula cinerea Delile) extracts against some pathogenic bacteria and fungi. Jordan Journal of Biological Sciences. 2013; 6(4): 266–71. https://doi.org/10.12816/0001624
- 25. Boussoula E, Ghanmi M, Satrani B, Alaoui MB, Rhafouri R, Farah A, Nadine A, Abdelaziz C. Chemical quality, antibacterial and antifungal activities of Cotula cinerea essential oil from South Morocco. ESAIJ. 2016; 12(5): 209–16.
- 26. El-Bouzidi L, Abbad A, Fattarsi K, Hassani L, Leach D, Markouk M, et al. Chemical composition and anticandidal properties of the essential oil isolated from aerial parts of Cotula cinerea: A rare and threatened medicinal plant in Morocco. Nat Prod Comm. 2011; 6(10): 1491–4. PMid:22164791
- 27. Elsharkawy ER, Hyaa G, Donia A. Comparison between Thuja orientalis growing in Egypt and Saudi Arabia. British Journal of Pharmaceutical Research. 2017; 15(5): 1–9. https://doi.org/10.9734/BJPR/2017/32387
- Sieniawska E, Świątek Ł, Rajtar B, Koziol E, Polz-Dacewicz M, Skalicka-Wózniak K. Carrot seed essential oil-source of carotol and cytotoxicity study. Industrial Crops and Products. 2016; 92: 109–115. Crossref
- Zarai Z, Ben Chobba I, Ben Mansour R, Békir A, Gharsallah N, Kadri A. Essential oil of the leaves of Ricinus communis L.: In vitro cytotoxicity and antimicrobial properties. Lipids in health and disease. 2012; 11: 102–108. https://doi.org/10.1186/1476-511X-11-102 1, 2.