J Endocrinol Reprod 8(1) (2): 83-94 (2004) JER 50

MELATONIN AND THE THYROID GLAND - AN OVERVIEW

LEWINSKI A^{1,2} and KARBOWNIK M^{1,2}

¹ Department of Endocrinology and Isotope Therapy, Medical University of Lodz, ² Polish Mother's Memorial Hospital - Research Institute, 281/289, Rzgowska St., 93-338 Lodz, Poland, e-mail: alewin@csk.am.lodz.pl

SUMMARY

In this review, published data on the relationship occurring between melatonin the main pineal hormone - and the thyroid gland, are briefly summarized. Numerous experimental data suggest an inhibitory influence of melatonin on thyroid growth and secretion. These effects have been presented in different experimental models *in vivo* and *in vitro*, the former including short-term and/or chronic melatonin administration to various animal species, in pinealectomized animals, also following light restriction which is known to increase the activity of the pineal. On the other hand, stimulatory effects of thyroid hormones on the pineal growth and function have been demonstrated. Oxidative stress is involved in physiological and pathological processes in the thyroid gland. Recent results of experimental studies suggest that melatonin can protect against oxidative damage in the thyroid. A confirmation of the mutual relationship between the pineal gland and the thyroid is - in clinical studies, in humans - rather difficult, as both human beings and animal species used in experimental studies, live far away from their natural/original habitats. It makes almost impossible to compare the results of studies on the pineal-thyroid interrelationship, obtained in particular experiments in different animal species.

Key words: Cell proliferation; Growth processes; Melatonin; Pineal gland; Thyroid gland; Thyroid hormones.

INTRODUCTION

Melatonin (N-acetyl-5-methoxytryptamine) is the main secretory product of the pineal gland. The molecule performs several functions in living organisms. It is known for its role in seasonal reproductive physiology, circadian rhythmicity and sleep processes and for its ability to reduce the "jet lag" symptoms in humans (1). Additionally, melatonin has been shown to modulate immune functions, growth processes, oxidative processes and, to some extent, the complex process of cancerogenesis (2-10).

The existing relationship between the pineal and the thyroid gland has been evidenced by results of numerous experimental studies. Several questions, however, have still remained unanswered, namely: 1) to what extent is the relationship in question direct? 2) are there any intermediate substances or factors involved in this regulation? 3) are there any cells in the body capable of producing both thyroid hormones and melatonin? 4) is there a local (paracrine?) regulation of thyroid hormone secretion by melatonin in the thyroid gland? 5) does melatonin participate in the regulation of the peripheral metabolism of thyroid hormones (T_4 -monodeiodination)? 6) does melatonin control the expression of certain thyroid gland-related genes, i.e, NIS, TPO, thyroglobulin, pendrin, TSH, TSH receptor (TSHR), etc.? 8) is T_3 involved in the regulation of the expression level of genes, encoding for key enzymes, which participate in melatonin synthesis (hydroxy-indole-O-methyltransferase - HIOMT; N-acetyltransferase - NAT)?

Corresponding author : Prof. A. Lewinski, M.D., Ph.D.

Numerous evidence has been gathered that, beside the pineal gland, other organs, tissues or cells (11) are sites of melatonin production, with exclusively high amounts found in the bone marrow (12, 13) and in the gut (14). The presence of this hormone has been observed in APUD cells of airway epithelium, along the border between the cortex and medulla of adrenals, beneath the hepatic capsule, in kidney cortex, paraganglia, gall bladder, ovary, endometrium, placenta, and the inner ear (15). Colocalizations of melatonin and calcitonin in thyroid C-cells, melatonin and histamine in mast cells, melatonin, somatostatin and beta-endorphins in natural killer cells, and melatonin and prostaglandin F_2 in thymic reticulo-epithelial cells have been observed (16, 17). No studies have yet been performed to reveal a possible presence of melatonin in thyroid follicular cells.

As far as the regulatory mechanisms between melatonin and thyroid hormones are concerned, typical endocrine (due to the fact that melatonin is a hormone), paracrine (due to the presence of melatonin in thyroid C cells), and - possibly - autocrine (if melatonin is present in thyroid follicular cells) regulation should be considered.

The inhibitory influence of melatonin on growth processes in the thyroid gland

Numerous data, from our and other laboratories, indicate a suppressive effect of melatonin on thyroid growth processes. Already in the sixties of XX century, it has been shown that removal of the pineal gland, being the main source of melatonin, resulted in an increased thyroid weight in rats (18) and mice (19). Later on, the inhibitory effects of melatonin on thyroid growth processes were confirmed, using different indices of growth processes and different experimental models.

It is well known that light decreases the activity of the pineal gland; suppression of the thyroid growth was found in male mice under conditions of light restriction (20). Melatonin, administered to mice in late-afternoon s.c injections for 10 days, inhibited the basal and thyrotropin (TSH)-stimulated mitotic activity of the thyroid follicular cells; a similar effect was observed in thyroid organ culture (21). The indoleamine prevented the pinealectomy-induced increase of mitotic indices in the rat thyroid gland (22). The effect of short-term s.c. administration of melatonin (5 days) on ³H-thymidine incorporation into DNA of rat thyroid lobes, transferred, after collecting, into incubation *in vitro*, was dose-dependent; melatonin, in dose of 25 μ g/daily, effectively reduced ³H-thymidine incorporation, when used in dose of 50 μ g/daily - melatonin produced no effect, however, the indoleamine, applied in the highest dose - 100 μ g/daily, brought about an increase of ³H-thymidine uptake (23). However, under *in vitro* conditions, melatonin reduced ³H-thymidine incorporation into DNA of rat thyroid lobes only when used in the concentration of 10⁻⁹ M (24).

The following observation is of special attention : melatonin, when released from s.c. pellets, prevented the inhibitory effect of late-afternoon melatonin injections on growth processes in rat thyroid (25, 26). Those results suggest a counter-antithyroid action of melatonin released from pellets on the growth-inhibiting response of the gland, following melatonin injection. This would suggest that melatonin is able to regulate its own action.

Melatonin and, to a lesser extent, another indoleamine - 5-methoxytryptamine, decreased the mean nuclear volume of thyroid follicular cells in Syrian hamsters (27). Melatonin and its precursor - N-acetylserotonin (NAS), administered to male rats, decreased the mitotic activity in the thyroid gland (28). The inhibitory effect of short photoperiod on the thyroid growth processes was shown in mice (20) and in Indian palm squirrels *Funambulus pennanti* (29). It is to be stressed that the pineal gland involvement in the photoperiodic response of the thyroid cannot be excluded, since parallel changes of melatonin concentrations were observed, following a short photoperiod exposure (29). Moreover, much experimental evidence derived either from our (21, 30) or from other (31,

32) laboratories, speak in favour of direct melatonin influence on thyroid follicular cells. This hypothesis is further confirmed by the finding that the pituitary is not necessary to demonstrate the increase in thyroid weight after pinealectomy in mice (19, 33).

The activities of the following enzymes, treated as indices of growth processes, have been measured at our laboratory: thymidine kinase, thymidine phosphorylase and adenosine kinase. Additionally, we have examined the effect of indoleamines on cyclic AMP generation in rat thyroids *in vitro*. Thymidine kinase (TK: thymidine 5'-phosphotransferase, EC 2.7.1.21) is an enzyme responsible for catalyzing the phosphorylation of thymidine, functioning as a part of the pyrimidine salvage pathway involved in DNA synthesis and being closely correlated with ³H-thymidine incorporation and mitosis. Adenosine kinase (AK; EC 2.7.1.20) is an enzyme which catalyses the phosphorylation of adenosine (Ado) and deoxyadenosine (dAdo) to adenosine monophosphate (AMP) and deoxyadenosine monophosphate (dAMP), respectively. Adenosine kinase functions as a part of the purine metabolic pathway involved in DNA synthesis and is the key enzyme regulating the Ado content. Thymidine phosphorylase (dThdPase, EC 2.4.2.4) is an enzyme catalyzing the reversible phosphorolysis of thymidine, deoxyuridine and their analogues to the respective bases and to 2-deoxyribose-1-phosphate. This enzyme has been proved to be identical with the platelet-derived endothelial cell growth factor (PD-ECGF), which is involved in the process of angiogenesis.

Melatonin and NAS have been shown to decrease the concentration of cyclic AMP (34) and to reduce the activity of TK (35) in rat thyroid lobes incubated *in vitro*. Whereas melatonin under conditions *in vitro*, decreased TK activity in thyroids collected from older rats (35), the indoleamine, added to the incubation medium containing thyroids collected from much younger - intact, shamoperated and hemithyroidectomized animals - increased TK activity (36). Thus, may be in young animals, melatonin serves as a hormone helping organs in growth and differentiation, whereas in adult animals the indoleamine prevents enhanced growth processes.

In another study, increased dThdPase activity in the remaining thyroid lobe was found following hemithyroidectomy. Melatonin, applied *in vitro*, decreased the dThdPase activity in thyroid lobes collected from intact animals, sham-operated animals and hemithyroidectomized rats (37). The results suggest an involvement of melatonin in the regulation of thyroid growth, probably by inhibition of the process of angiogenesis.

Concerning AK, a decreased activity of the enzyme was found in the remaining thyroid lobe, following hemithyroidectomy; melatonin, used *in vitro*, increased AK activity in thyroid lobes collected from intact and sham-operated rats, but it did not change AK activity in the remaining thyroid lobes after hemithyroidectomy (36). The results suggest a certain role of AK in the regulation of (patho)physiological processes in the thyroid gland after hemithyroidectomy.

Karyometry is the method used to assess the activity rate - mainly - of secretion but also of growth processes in various tissues and organs. An increased volume of cell nuclei may either result from enhanced DNA synthesis or emerge from a stimulated functional activity (increased protein synthesis). At our laboratory, we examined the influence of melatonin and TSH on karyometric parameters of rat thyrocytes. We found that a short-photoperiod exposure, associated with a stimulation of the pineal gland, resulted in a decrease of the mean volume of thyrocyte nuclei in male gerbils (38). Additionally, we observed that melatonin, administered in late-afternoon injections, decreased the mean nuclear volume of thyrocytes in male Syrian hamsters (27), and, when used *in vitro*, the indole significantly decreased the mean nuclear volume and the nuclear intersection area of thyrocytes (39).

The protective effects of melatonin against cancer is a subject of an intensive research (2-4,40). Because of the potential role of ionizing radiation in the pathogenesis of thyroid cancer, the studies on protective effects of melatonin against radiation-induced oxidative stress and cancer of the thyroid gland seem to be of special value. However, the published data concerning this issue, are rather scarce. It has been found that histoenzymological changes in rat thyroid gland, caused by an exposure to γ -radiation (8 Gy), were partially reversed by pretreatment with melatonin (41). In another study, when using morphometric parameters, melatonin was shown to decrease the height of thyroid follicular cells and the nuclear volume of the cells from rats exposed to 8 Gy-radiation (42). The potential protective effect of melatonin against thyroid cancer will unquestionably be a subject of future studies.

The influence of melatonin on thyroid secretion

Most studies, related to the effects of melatonin on thyroid function, revealed that this influence is an inhibitory one. Late afternoon s.c. injections of melatonin decreased circulating thyroid hormone concentrations in adult Syrian hamsters of both sexes (melatonin - 25 µg/daily) (43) and in male Wistar rats (melatonin - 50 µg/daily) (44).

An increase in serum thyroxine (T_4) concentration was found 10 weeks after pinealectomy performed in male Wistar rats, that process being prevented by melatonin administration, the concentrations of triiodothyronine (T_4) remaining unchanged in the pinealectomized rats (45).

However, an opposite effect was observed when melatonin was chronically released from s.c. pellets, implanted to male Wistar rats; the indoleamine increased both T_3 and T_4 levels after 10 days and also, however to a lesser degree, after 10 weeks; this effect may be called the "prothyroid" action of melatonin (44). On the other hand, the joint effect of late-afternoon melatonin injections and melatonin-implants caused no changes in thyroid hormone concentrations (44). Thus, this could be another piece of evidence, beside that observed in relation to growth processes, that melatonin is able to regulate its own action.

Not only may chronic melatonin availability but also a short-term treatment with the hormone result in a "prothyroid" action under certain conditions. Unexpectedly, when melatonin was injected in a dose of 25 µg/daily to rats for 5 consecutive days in the late light phase, it increased serum T₃ concentration, revealing a slight tendency towards rising serum T₄ (46). In the same study, also a 5-days treatment with NAS resulted in a "prothyroid" effect, concerning thyroid secretory processes.

Petterborg and Rudeen (47) have demonstrated that chronic afternoon melatonin administration in female hamsters results in a loss of estrous cyclicity, a significant gain in body weight and reduction of T_4 levels and T_3 uptake.

Wright *et al.* (48) were the first authors to have succeeded in showing an influence of melatonin on the amphibian thyroid gland. They found that melatonin directly antagonized the action of T_4 in promoting regression of tadpole tail tips *in vitro* (48). The *in vitro* secretion of T_4 from prometamorphic *Rana catesbeiana* tadpoles was significantly inhibited by melatonin in concentration of 10 mg/ml, and - especially - by that indoleamine in a higher concentration - 100 mg/ml, also in response to TSH; a complete suppression of the thyroid response to TSH was observed (49). In another *in vitro* experiment, thyroids from larval *Rana catesbeiana* or adult *Rana pipiens* were incubated in control or melatonin (0.01 to 100 mg/ml) media; melatonin directly inhibited T_4 secretion by thyroids from both tadpoles and frogs at all concentrations of melatonin used and at both prometamorphic and climax tadpole stages (50).

Lewinski and Karbownik

Melatonin administration to turtles decreased plasma T₄ levels, which was accompanied by reduced thyroid weight, reduced follicular epithelial cell height and a decreased activity of thyroid peroxidase (51). Effects of continuous light (CL; 24 L : 0 D), continuous darkness (CD; 24 D : 0 L) and pinealectomy (Px) were tested on the pineal-thyroid-gonadal axis of a tropical seasonally bred rodent, Funambulus pennanti, during its sexually active reproductive phase (February-March). CL had no effect on pineal, thyroid and ovarian functions, as demonstrated by thyroid and ovarian weight, plasma levels of T₄ and estradiol (E₂). However, CD reduced significantly thyroid and ovarian weight and plasma T₄ and E₂ concentrations (52). Pinealectomy resulted in stimulation of thyroid and ovarian functions under normal photoperiod, and under CL, Px significantly increased thyroid and ovarian weights with no observable changes in their hormonal levels. Under CD condition, Px prevented the reduction in thyroid and ovarian weights but the hormone levels were increased (52). These results suggest that CD may inhibit thyroid and ovarian functions by the stimulation of melatonin secretion (52). Furthermore, melatonin injection reduced T_4 concentration in control rats, and T_3 concentration in rats with transplanted anterior pituitary (53). The influence of melatonin on thyroid hormone secretion could either be direct or indirect. It has recently been shown that injections of melatonin caused a decrease in both blood TSH and thyroid hormone concentrations in rats (54). On the other hand, it has been shown that melatonin stimulates, whereas pinealectomy decreases TSH accumulation in the unique thyroid hormone-immunoreactive cells in rat pars tuberalis (55).

The influence of melatonin on the activities of monodeiodinases - enzymes participating in thyroid hormone metabolism in peripheral tissues - was also studied. It was previously shown that melatonin, released from s.c. pellets for 15 days, enhanced type II thyroxine 5'-monodeiodinase in brown adipose tissue of Syrian hamsters, without changing of serum thyroid hormone concentrations (56). Similarly, activation of cerebrocortical type II 5'-deiodinase activity in Syrian hamsters, kept under short photoperiod or subjected to the indoleamine liberated from s.c. pellets, was observed (57). It has been found in the more recent studies that treatment with melatonin results in an increased activity of type I 5'-monodeiodinase in the liver and kidney and of type II 5'-monodeiodinase in adipose tissue of newborn rabbits, the changes being accompanied by increased concentrations of serum T_3 and reverse T_3 , and - unexpectedly - by increased concentration was probably due to the stimulatory effect of melatonin on the secretory activity of the thyroid gland (58). Moreover, melatonin supplementation in hyperthyroid rats was found to suppress secretion of thyroid hormones and of testosterone (59).

Melatonin as antioxidant and the thyroid gland

Several thyroid disorders are accompanied by enhanced oxidative stress. In an experimental model of hyperthyroidism, a 2-week-treatment with L-T₄ (100 μ g/kg b. w. for 14 days) resulted, expectedly, in an increased concentration of free fractions of both thyroid hormones; a co-treatment with melatonin (5 mg/kg b. w. for 7 days) completely prevented the increase in the concentration of free T₂, being the most active form of thyroid hormones (60).

In another study, $L-T_4$ -treatment resulted in an increased level of Schiff's bases, malondialdehyde and conjugated dienes concentrations - all being parameters of oxidative stress - in lung, brain and kidney homogenates; these changes were reversed by melatonin (61, 62).

Under physiological conditions, numerous free radicals and reactive species are produced in the thyroid gland (63). However, the oxidative reactions undergoing in the thyroid gland, could be - under certain pathological conditions - the source of oxidative damage. Iron, which is present in the thyroid peroxidase (TPO) - the key enzyme catalyzing thyroid hormone synthesis - and H_2O_2 , which is essential for TPO activity, constitute substrates for Fenton reaction. Thus, using ferrous iron (Fe²⁺) and H_2O_2 , we have recently induced oxidative damage to lipids in

homogenates of porcine thyroid; Fenton reaction-induced lipid peroxidation [measured by the level of malondialdehyde + 4-hydroxyalkenals (MDA+4-HDA)] was prevented by melatonin in a concentration-dependent manner (64). Thus, we have shown for the first time that, under experimental conditions, it is possible to induce lipid peroxidation in the thyroid by using substrates of Fenton reaction and that this process could be prevented by melatonin.

Thyroid hormone-stimulation of pineal function or growth processes

The stimulatory effect of the thyroid hormones on the pineal gland is supported by many morphological, biochemical and clinical findings. Peschke (65-67) reported that T_4 significantly increased the surface area of cross sections of nuclei in rat pinealocytes *in vivo*. Thyroidectomy and/or methylthiouracil (an antithyroid drug) treatment caused a significant decrease of the surface area in question. Also the results of our studies speak in favour of pineal growth stimulation by thyroid hormones; thyroid hormones increased the mean nuclear volume of pinealocytes in organ culture, as well as slightly increased the mean mitotic activity rate of pinealocytes (68). In turn, Milcou *et al.* (69) found a significantly increased amount of DNA in rat pineals, following the administration of T_4 to culture medium."Our hypothesis was further supported by the observation that T_4 and T_3 increased melatonin concentration and norepinephrine-stimulated NAS content in cultured rat pineals (70).

The synthesis and release of melatonin were studied in pineal explants from 14-day-old (young) and 60-day-old ("maturing") male Long-Evans rats, in either the absence or the presence of T_3 being at or close to physiological levels and under light and dark conditions (71). Under light conditions, T_3 increased melatonin content in pineals and in medium of pineal cultures, the gland being collected from either young or maturing animals. Under dark conditions, T_3 decreased melatonin levels in the pineals of either age, but it did not affect melatonin levels in the medium (71). Since it is known from another study that 14-day-old rat pineal glands do not yet have a complete sympathetic innervation system, it seems evident that T_3 can modulate directly the pineal synthesis and the release of melatonin, while not depending upon a mature sympathetic innervation (71). Light - in the studied conditions - was permissive for the stimulatory action of T_3 on pineal synthesis and release of melatonin *in vitro* (71). The results confirm the pineal-thyroid feedback hypothesis, proposed originally by us (72-74).

With agreement with above observations is the finding that treatment with methimazole (resulted in hypothyroidism) caused a decrease in NAT activity in the Harderian gland of the male Syrian hamsters (75). In studies *in vivo*, treatment with T_4 resulted in an increased nocturnal peak of melatonin in rats (76).

Pineal-thyroid relationship in humans

The clinical data on the pineal-thyroid relationship are rather scarce. Whereas no changes were observed by some authors (77) in melatonin levels in either hypothyroidism or hyperthyroidism of human subjects, others investigators have found an increase in nocturnal melatonin concentrations in hypothyroid patients (78). Patients with hypothyroidism were found to have higher peak serum melatonin values, total nocturnal melatonin secretion and urinary excretion of melonin than normal individuals (78). Neither of those values differed significantly from normal in patients with thyrotoxicosis or obesity. Although thyrotoxic patients released normal amounts of melatonin during the night, their melatonin secretion peaks were phase-advanced (melatonin secretion peak appearing at 1.1 ± 0.5 h in thyrotoxic, and at 3.4 ± 0.5 h in normal participants). No such phasal shifts were seen in patients with obesity or hypothyroidism (78). Those findings imply that both hypothyroid and thyrotoxic patients have disturbed pineal function, which is not the case in patients with obesity (78). A decreased nocturnal melatonin blood concentration was observed in patients with recurrent non-toxic nodular goitre when compared to respective values in controls (79).

Blood concentrations of melatonin were evaluated in patients with very large non-toxic nodular goitre before and after thyroidectomy; unexpectedly, nocturnal melatonin concentrations

were significantly higher after than before the operation (80). The authors have drawn a conclusion that a very large goitre can possibly compress the superior cervical ganglia, and in consequence indirectly alter melatonin synthesis. Another explanation can be based on the assumption that melatonin may actively be taken up by enlarged thyroid with subsequent decrease in blood concentration of the indoleamine.

Melatonin treatment for 3-6 months in perimenopausal and menopausal women (aged 42-62 years), with initial low level of blood melatonin, resulted in a significant increase in thyroid hormone concentrations (81); thus, melatonin reveals a recovery effect of thyroid function towards a more juvenile pattern of regulation. It is not excluded that the described effect is a direct one because a similar treatment with melatonin in aging patients did not result in any changes in TSH concentration (82).

Concluding remarks

All the above mentioned results, while proving the suppression of thyroid growth and/or thyroid function by the pineal (83), as well as the reports on stimulation of the pineal gland activity and growth processes by the thyroid hormones (68, 70, 76, 84), have prompted us to formulate a hypothesis on the existence of a reciprocal relationship between the thyroid and the pineal gland (73, 74, 83) (Fig. 1).

The main conclusions from the up-to-date results, concerning thyroid-pineal relationship, are as follows: 1) melatonin influence on thyroid growth processes and thyroid hormone synthesis seems to be complex; it should be stressed once again that the evidence of the mutual relationship between the pineal gland and the thyroid is derived, almost exclusively, from studies performed in experimental animals; 2) the confirmation of these relations in clinical studies meets numerous difficulties and pitfalls, resulting, among others, from the fact that, nowadays, human beings, as well as animal species used in experimental studies, live far away from their natural and original habitats; 3) however, still much evidence indicates an undoubtful role of melatonin in physiological and pathological processes of the thyroid gland, providing "green light" for the future use of this indoleamine under certain clinical conditions.

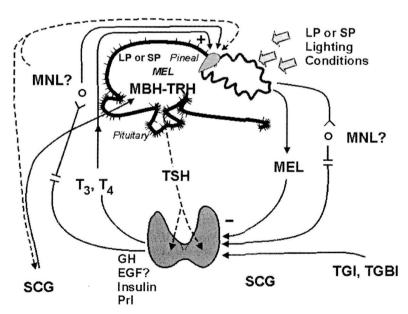


Fig. 1. A general model of possible reciprocal relationships between the pineal gland, the superior cervical ganglia and the thyroid, involved in the control of thyroid growth. EGF - epidermal growth factor, GH - growth hormone, IGFs - insulin-like growth factors, LP - long photoperiod, MBH - medio-basal hypothalamus, MEL - melatonin, MNL multineuronal link, Prl - prolactin, SCG - superior cervical ganglion, SP - short photoperiod, T₃ - triiodothyronine, T₄ - thyroxine, TGBI - thyroid growth-blocking immunoglobulins, TGI -thyroid growth-stimulating immunoglobulins, TRH - thyrotropin releasing hormone, TSH -thyrotropin, VIP - vasoactive intestinal peptide.

REFERENCES

- 1 Reiter RJ (1993). The melatonin rhythm: Both a clock and a calendar. *Experientia* 49: 654-664.
- 2 Karasek M and Pawlikowski M (1999). Pineal gland, melatonin and cancer. *Neuroendocrinol Lett* 20: 139-144.
- 3 Karbownik M and Reiter RJ (2000). Antioxidative effects of melatonin in protection against cellular damage caused by ionizing radiation. *Proc Soc Exp Biol Med* **225:** 9-22.
- 4 Karbownik M, Lewinski A and Reiter RJ (2001). Anticarcinogenic actions of melatonin which involve antioxidative processes: comparison with other antioxidants. *Int J Biochem Cell Biol* **33**: 735-753.
- 5 Lewinski A, Wajs E, Klencki M, Karbownik M, Gesing A, Sewerynek E, Slowinska-Klencka D, Skowronska-Jozwiak E, Bilinski P and Krotewicz M (1997). Pineal-thyroid interrelationships update: 1996. In: Webb SM, Puig- Domingo M, Moller M and E Pevet (eds.), *Pineal Update from Molecular Mechanisms to Clinical Implications*, PJD Publications Limited, Westbury, pp. 173-181.
- 6 Lewinski A, Wajs E, Klencki M, Krotewicz M and Bilinski P (1996). Melatonin-induced inhibition of thyroid growth processes and possible mechanisms of that phenomenon. In: Haldar C (ed.), *Recent Researches in Biology: The Pineal Gland: Its Molecular Signals*, Hindustan Publishing Corporation, New Delhi, Vol I, pp. 27-32.
- 7 Lewinski A, Sewerynek E and Karbownik M (2002). Melatonin from the past in the future- our own experience. In : Halder C, Singaravel M and Maitra SK (eds.), Enfield (NH), USA and Plymouth, UK: Treatise on Pineal Gland and Melatonin, Science Publishers Inc, USA, pp.157-175.
- 8 Lewinski A and Karbownik M (2002). Melatonin and the thyroid gland. *Neuroendocrinol Lett* **23** (suppl 1): 73-78.
- 9 Reiter RJ, Calvo JR, Karbownik M, Qi W and Tan DX (2000). Melatonin and its relation to the immune system and inflammation. *Ann N Y Acad Sci* **917**: 376-386.
- 10 Reiter RJ, Tan DX, Qi W, Manchester LC, Karbownik M and Calvo JR (2000). Pharmacology and physiology of melatonin in the reduction of oxidative stress *in vivo*. *Biol Signals Recept* **9**: 160-171.
- 11 Kvetnoy IM (1999). Extrapineal melatonin: location and role within diffuse neuroendocrine system. *Histochem J* **31**: 1-12.
- 12 Conti A, Conconi S, Hertens E, Skwarlo-Sonta K, Markowska M and Maestroni GJM (2000). Evidence for melatonin synthesis in mouse and human bone marrow cells. *J Pineal Res* 28: 193-202.
- 13 Tan DX, Manchester LC, Reiter RJ, Qi WB, Zhang M, Weintraub ST, Cabrera J, Sainz RM, Mayo JC (1999). Identification of highly elevated levels of melatonin in bone marrow : its origin and significance. *Biochim Biophys Acta* **1472**: 206-214.
- 14 Bubenik GA (2002). Gastrointestinal melatonin: localization, function, and clinical relevance. *Dig Dis Sci* **47:** 2336-2348.
- 15 Raikhlin NT and Kvetnoy IM (1994). The APUD system (diffuse endocrine system) in normal and pathological states. *Physiol Gen Biol Rev* 8: 1-44.
- 16 Kvetnoy IM and Yuzhakov VV (1993). Extrapineal melatonin: advances in microscopical identification of hormones in endocrine and nonendocrine cells. *Microsc Anal* **21**: 27-29.
- 17 Kvetnoy IM and Yuzhakov VV(1994). Extrapineal melatonin: non-traditional localization and possible significance for oncology. In: Maestroni GJM, Conti A and Reiter RJ (eds.), Advances in Pineal Research, Vol 7, John Libbey and Company, London, pp. 199-212.
- 18 Miline R (1963). La part du noyau paraventriculaire dans 1-histopathologie correlative de la glans et de la glande pineale. *Ann Endocrinol* **24:** 255-269.

90

- 19 Houssay AB, Pazo JH and Epper CE (1966). Effects of the pineal gland-upon the hair cycles in mice. J Invest Dermatol 47: 230-234.
- 20 Lewinski A, Vaughan MK, Champney TH, Reiter RJ and Smith NKR (1984). Dark exposure inhibits the mitotic activity of thyroid follicular cells in male mice with intact pineal. *Experientia* **40**:1284-1285.
- 21 Lewinski A and Sewerynek E (1986). Melatonin inhibits the basal and TSH- stimulated mitotic activity of thyroid follicular cells *in vivo* and in organ culture. *J Pineal Res* **3**: 291-299.
- 22 Wajs E, Krotewicz M, Fryczak J, Kulak J, Sewerynek E, Szkudlinski M and Lewinski A (1989). Melatonin suppresses the pinealectomy-induced increase of mitotic incidence in the rat thyroid gland. *Med Sci Res* 17: 61-62.
- 23 Wajs E, Lewinski A, Krotewicz M and Kunert-Radek J (1992). [3H]-thymidine incorporation into DNA of thyroid lobes incubated *in vitro*, following pretreatment of animals with melatonin and thyrotropin. *Neuroendocrinol Lett* **14**: 75-81.
- 24 Wajs E and Lewinski A (1988). Melatonin and N-acetylserotonin two pineal indoleamines inhibiting the proliferation of jejunal epithelium cells in rats. *Med Sci Res* 16: 1125-1126.
- 25 Wajs E and Lewinski A (1992). Inhibitory influence of late-afternoon, melatonin injections and the counter-inhibitory effect of melatonin pellets on thyroid growth processes in male Wistar rats; comparison with effects of other indole substances. J Pineal Res 13: 158-166.
- 26 Lewinski A, Wajs E and Krotewicz M (1993). Melatonin and other indolic substances: their influence on thyroid growth and secretion. In : Touitou Y, Arendt J and Pevet P (eds.), *Melatonin and the Pineal Gland - From Basic Science to Clinical Application*, Excerpta Medica, Amsterdam, pp. 265-268.
- 27 Lewinski A, Webb SM, Sewerynek E, Champney TH, Vaughan MK and Reiter RJ (1986). Influence of melatonin and 5-methoxytryptamine on the nuclear volume of thyroid follicular cells in the Syrian hamster (*Mesocricetus auratus*). *Neuroendocrinol Lett* 8: 63-68.
- 28 Sewerynek E, Lewinski A, Szkudlinski M and Zerek-Melen G (1988). The effect of melatonin and N-acetylserotonin on mitotic activity of thyroid gland and adrenal cortex in the rat. *Endokrynol Pol* **39**: 269-275.
- 29 Haldar C, Shavali SS and Singh S (1992). Photoperiodic response of pineal- thyroid axis of the female Indian palm squirrel, *Funambulus pennanti. Neural Transm* **90:** 45-52.
- 30 Wajs E and Lewinski A (1991). Effects of melatonin on [³H]-thymidine incorporation into DNA of rat thyroid lobes *in vitro*. *Biochem Biophys Res Commun* **181**: 1187-1191.
- 31 Singh AK and Prasad GC (1981). *In vivo* and *in vitro* studies on the pineal thyroid relationship. *Indian J Med Res* **74:** 420-427.
- 32 Haldar C and Shavali SS (1992). Influence of melatonin on thyroxine (T₄) release from thyroid glands of female *Funambulus pennanti* : An *in vitro* study. *Neuroendocrinol Lett* **14**: 411-416.
- 33 Houssay AB and Pazo JH (1968). Role of pituitary in the thyroid hypertrophy of pinealectomized rats. *Experientia* 24: 813-814.
- 34 Lewinski A, Sewerynek E, Zerek-Melen G, Kunert-Radek J, Pawlikowski M and Karasek E (1989). Influence of melatonin and N-acetylserotonin on the cyclic AMP concentration in the rat thyroid lobes incubated *in vitro*. J Pineal Res 7: 55-61.
- 35 Lewinski A, Wajs E, Modrzejewska H, Klencki M, Karbownik M and Greger J (1994). Inhibitory influence of melatonin on thymidine kinase activity in the rat thyroid lobes incubated *in vitro*. *Neuroendocrinol Lett* 16: 221-226.

- 36 Gesing A, Modrzejewska H, Karbownik M, Sewerynek E, Greger J and Lewinski A (2000). Thymidine kinase and adenosine kinase activities in homogenates of thyroid lobes in hemithyroidectomized rats; effects of melatonin *in vitro*. *Neuroendocrinol Lett* 21: 453-459.
- 37 Gesing A, Miszczak-Zaborska E, Karbownik M, Sewerynek E, Greger J and Lewinski A (1999). Effects of hemithyroidectomy on thymidine phosphorylase in homogenates of rat thyroid lobes incubated *in vitro* in the presence of melatonin. *Thyroidol Clin Exp* **11**: 19-24.
- 38 Lewinski A, Vaughan MK, Champney TH, Reiter RJ and Smith NKR (1984). Inhibitory action of the pineal gland on the volume of thyroid follicular cells in male gerbils (*Meriones unguiculatus*). *Exp Clin Endocrinol* 84: 239-244.
- 39 Klencki M, Slowinska-Klencka D, Kunert-Radek J and Lewinski A (1994). Melatonin- induced decrease of the size of thyrocytes nuclei in rat thyroids incubated in vitro. Cytobios 78: 159-162.
- 40 Karbownik M and Reiter RJ (2002). Melatonin protects against oxidative stress caused by delta-aminolevulinic acid: Implication for cancer reduction, *Cancer Invest* **20**: 276-286.
- 41 Kundurovic Z and Scepovic M (1989). Histoenzymological reactions of the thyroid gland in irradiated and previously melatonin-treated irradiated rats. *Acta Med Yugosl* **43**: 337-347.
- 42 Kundurovic Z and Mornjakovic Z (1992). Morphometric characteristics of thyroid cells in irradiation stressed rats treated with pinealectomy and melatonin [In Serbo-Croatian (Roman)]. Med Arh 46: 9-10.
- 43 Vaughan MK, Richardson BA, Petterborg LJ, Holtorf AP, Vaughan GM, Champney TH and Reiter RJ (1984). Effects of injection and/or chronic implants of melatonin and 5 methoxytryptamine on plasma thyroid hormone in male and female Syrian hamsters. *Neuroendocrinology* 39: 361-366.
- 44 Krotewicz M, Lewinski A and Waja E (1992). The inhibitory effect of late afternoon melatonin injections, but not of melatonin-containing subcutaneous implants, on thyroid hormone secretion in male Wistar rats. *Neuroendocrinol Lett* 14: 405-411.
- 45 Krotewicz M and Lewinski A (1994). Effects of pinealectomy and of late afternoon injections of pineal indole substances on thyroid hormone secretion in male Wistar rats. *Biochem Lett* **50**: 101-107.
- 46 Krotewicz M and Lewinski A (1994). Thyroid hormone secretion in male Wistar rats treated with melatonin and/or thyrotropin; dependence of effects on the used doses. *Neuroendocrinol Lett* **16:** 263-268.
- 47 Petterborg LJ and Rudeen PK (1989). Effects of daily afternoon melatonin administration on body weight and thyroid hormones in female hamsters. *J Pineal Res* **6**: 367-373.
- 48 Wright ML, Pikula A, Cykowski LJ and Kuliga K (1996). Effect of melatonin on the anuran thyroid gland: follicle cell proliferation, morphometry, and subsequent thyroid hormone secretion *in vitro* after melatonin treatment *in vivo*. *Genv Comp Endocrinol* **103**: 182-191.
- 49 Wright ML, Pikula A, Babski AM, Labieniec KE and Wolan RB (1997). Effect of melatonin on the response of the thyroid to thyrotropin stimulation *in vitro*. Gen Comp Endocrinol **108**: 298-305.
- 50 Wright ML, Cuthbert KL, Donohue MJ, Solano SD and Proctor KL (2000). Direct influence of melatonin on the thyroid and comparison with prolactin. *J Exp Zool* **286**: 625-631.
- 51 Sarkar S, Sarkar NK, Bjattacharyya S and Das P (1997). Melatonin action on thyroid activity in the soft-shelled turtle, *Lissemys punctata punctata*. *Folia Biol (Krakow)* **45**: 109-112.
- 52 Shavali SS and Haldar C (1998). Effects of continuous light, continuous darkness and pinealectomy on pineal-thyroid-gonadal axis of the female Indian palm squirrel, *Funambulus pennanti. J Neural Transm* **105**: 407-413.

Lewinski and Karbownik

- 53 Esquifino A, Agrasal C, Velazquez E, Villanua MA and Cardinali DP (1997). Effect of melatonin on serum cholesterol and phospholipid levels, and on prolactin, thyroid- stimulating hormone and thyroid hormone levels, in hyperprolactinemic rats. *Life Sci* **61**: 1051-1058.
- 54 Ozturk G, Coskun S, Erbas D and Hasanoglu E (2000). The effect of melatonin on liver superoxide dismutase activity, serum nitrate and thyroid hormone levels. *Jpn J Physiol* **50**: 149-153.
- 55 Sakamoto S, Nakamura K, Inoue K and Sakai T (2000). Melatonin stimulates thyroid-stimulating hormone accumulation in the thyrotropes of the rat pars tuberalis. *Histochem Cell Biol* **114**: 213-218.
- 56 Puig-Domingo M, Guerrero JM, Menendez-Pelaez A and Reiter RJ (1989). Melatonin specifically stimulates type-II 5'-deiodination in brown adipose tissue of Syrian hamsters. J Endocrinol 122: 553-556.
- 57 Puig-Domingo M, Guerrero JM, Vaughan MK, Little JC and Reiter RJ (1989). Activation of cerebrocortical type II 5'-deiodinase activity in Syrian hamsters kept under short photoperiod and reduced ambient temperature. *Brain Res Bull* 22: 975-979.
- 58 Brzezinska-Slebodzinska E, Slebodzinski AB and Styczynska E (1998). Stimulatory effect of melatonin on the 5'-monodeiodinase activity in the liver, kidney, and brown adipose tissue during the early neonatal period of the rabbit. J Pineal Res 24: 137-141.
- 59 Mogulkoc R and Baltaci AK (2003). The effect of intraperitoneal melatonin supplementation on the release of thyroid hormones and testosterone in rats with hyperthyroidism. *Neuroendocrinol Lett* **24**: 345-347.
- 60 Sewerynek E, Wiktorska J and Lewinski A (1999). Effects of melatonin on the oxidative stress induced by thyrotoxicosis in rats. *Neuroendocrinol Lett* **20**: 157-163.
- 61 Wiktorska J, Sewerynek E and Lewinski A (2000). Effects of melatonin and of other antioxidants on the Schiff bases induced by thyrotoxicosis in rats. 12th International Thyroid Congress; October 22-27, 2000; Kyoto, Japan. *Endocrine J* (suppl) 47: p.530 p.239.
- 62 Wiktorska J, Sewerynek E and Lewinski A (2000). Effects of different antioxidants on the oxidative damage induced by L-thyroxine injections in rats. 11th International Congress of Endocrinology; October 29-November 2, 2000; Sydney, Australia, Abstract book p. 797 p. 294.
- 63 Karbownik M and Lewinski A (2003). The role of oxidative stress in physiological and pathological processes in the thyroid gland; possible involvement in pineal-thyroid interactions. *Neuroendocrinol Lett* **24**: 293-303.
- 64 Karbownik M and Lewinski A (2003). Melatonin reduces Fenton reaction- induced lipid peroxidation in porcine thyroid tissue. *J Cell Biochem* **90:** 806-811.
- 65 Peschke E (1981). Morphologische, physiologische und statistische Untersuchungen an der maennlicher Wistar-Ratte zum Problem eines moeglichen funktionellen Connexus: Epiphysis cerebri-Schilddruese. Teil IV: Neurosekretorischer Hypothalamus und Epiphyse. Zool Jahr Anat 105: 147-176.
- 66 Peschke E (1981). Morphologische, physiologische und statistische Untersuchungen an der maennlicher Wistar-Ratte zum Problem eines moeglichen funktionellen Connexus: Epiphysis cerebri-Schilddruese. Teil V: Zusammenfassung der Befunde und Diskussion. Zool Jahr Anat 105: 297-319.
- 67 Peschke E (1981). Morphologische, physiologische und statistische Untersuchungen an der maennlicher Wistar-Ratte zum Problem eines moeglichen funktionellen Connexus: Epiphysis cerebri-Schilddruese. Teil VI: Ergebnisse der Untersuchungen und Literatur. Zool Jahr Anat 105: 320-340.
- 68 Lewinski A, Sewerynek E and Zerek-Melen G (1986). Thyroid hormone- induced activation of rat pinealocytes in organ culture. *Neurosci Lett* (*Suppl*) **26:** S 302.

- 69 Milcou SM, Holban R, Tasca C, Ghinea E and Stanescu O (1968). *In vitro* study of thyroxine effects on enzymatic activity and cell differentiation in the pineal gland. *Rev Roum Endocrinol* 5: 203-207.
- 70 Nir I and Hirschmann N (1978). The effect of thyroid hormones on rat pineal indoleamine metabolism *in vitro*. J Neural Transm 42: 117-126.
- 71 Catala MD, Quay WB and Timirast (1988). Effects of thyroid hormone on light/dark melatonin synthesis and release by young and maturating rat pineal glands in vitro. Int J Dev Neurosci 6: 285-288.
- 72 Lewinski A (1986). Evidence for pineal gland inhibition of thyroid growth: contribution to the hypothesis of a negative feedback between the thyroid and the pineal. In: Reiter RJ and Lukaszyk A (eds.), *Advances in Pineal Research*, Vol I, John Libbey and Company Ltd., London, pp. 167-176.
- 73 Lewinski A, Webb SM and Reiter RJ (1984). Possible mechanisms of TSH- independent thyroid growth. *Med Hypotheses* **14:** 141-160.
- 74 Lewinski A, Webb SM and Reiter RJ (1987). Pineal inhibition of thyroid growth: its involvement in a possible negative feedback interaction between both glands. In: Reiter RJ (ed.), *Pineal Research Reviews*, Vol V, Alan R. Liss, Inc, New York, pp. 69-94.
- 75 Buzzell GR, Chen Z, Vaughan MK and Reiter RJ (1989). Effects of inhibition of thyroid function and of cold on melatonin synthesis and porphyrin content in the Harderian glands of male Syrian hamsters, *Mesocricetus auratus*. *Comp Biochem Physiol* **A94**: 427-429.
- 76 Bondarenko LA (1991). Effects of excess and deficiency of thyroid hormones in the body upon blood melatonin in pubertal male rats. *Bull Exp Biol Med* **111**: 590-591.
- 77 Soszynski P, Zgliczynski S and Pucilowska J (1988). The circadian rhythm of melatonin in hypothyroidism and hyperthyroidism. *Acta Endocrinol* (Copenh) **119**: 240-244.
- 78 Rojdmark S, Berg A, Rossner S and Wetterberg L (1991). Nocturnal melatonin secretion in thyroid disease and in obesity. *Clin Endocrinol* (Oxf) **35:** 61-65.
- 79 Kuzdak K (1995). The role of endocrine and autocrine growth factors after subtotal restrumectomy in non-toxic nodular goitre special reference to risk of goitre relapse. Endokrynol Pol *Polish J Endocrinol* **46 suppl. 2 to no. 2:** 59-68.
- 80 Karasek M, Stankiewicz A, Bandurska-Stankiewicz E, Zylinska K, Pawlikowski M and Kuzdak K (2000). Melatonin concentrations in patients with large goiter before and after surgery. *Neuroendocrinol Lett* **21**: 437-439.
- 81 Bellipanni G, Bianchi P, Pierpaoli W, Bulian D and Ilyia (2001). Effects of melatonim in perimenopausal and menopausal women: a randomized and placebo controlled study. *Exp Gerontol* **36**: 297-310
- 82 Siegrist C, Benedetti C, Orlando A, Beltran JM, Tuchscherr L, Noseda CM, Brusco LI and Cardinali DI (2001). Lack of changes in serum prolactin, FSH, TSH, and estradiol after melatonin treatment in doses that improve sleep and reduce benzodiazepine consumption in sleep-disturbed, middle-aged, and elderly patients. *J Pineal Res* **30**: 34-42.
- 83 Lewinski A (1990). Some aspects of the pineal-thyroid interrelationship and their possible involvement in the regulation of function and growth of these two glands. In: Reiter RJ and Lukaszyk A (eds.), *Advances in Pineal Research*, Vol IV, John Libbey and Company Ltd., London, pp. 175-188.
- 84 Rom-Boguslavskaia ES, Bondarenko LA and Sil'chenko TN (1991). Epiphyseal-thyroid interrelationship: effect of calcitonin on indole metabolism in health and in the presence of an excess of thyroid hormones. (In Russian with English Abstract) Probl Endokrinol (Moskwa) 37: 33-35.

94