

## Action of thyroid inhibitor propyl thiouracil on thyroid and interrenal axes in the freshwater tilapia *Oreochromis mossambicus* Peters

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### SUMMARY

The actions of propyl thiouracil (PTU), a thyrostatic drug, on thyroid hormones (THs) and cortisol production were examined in freshwater tilapia (*Oreochromis mossambicus*) to examine whether the hypothyroid state affects the function of interrenal axes in this fish. The plasma levels of triiodothyronine ( $T_3$ ), thyroxine ( $T_4$ ), and cortisol along with branchial and renal  $Na^+$ ,  $K^+$ -ATPase activities were measured after feeding varied doses (5-20  $\mu g g^{-1}$ ) of PTU for over 10 or 15 days. Feeding low dose (5  $\mu g g^{-1}$ ) of PTU produced significant rise in plasma  $T_4$  level but both  $T_3$  and  $T_4$  levels were decreased significantly after feeding a high dose (20  $\mu g g^{-1}$ ) of PTU for 15 days. All doses of PTU failed to alter the plasma cortisol. No significant correlation was found between the plasma levels of  $T_3$ ,  $T_4$  and cortisol after varied doses of PTU. Treatment of  $T_3$  or  $T_4$  (40 ng  $g^{-1}$ ) to the high dose PTU-fed tilapia produced a significant rise in plasma cortisol, suggesting a link to the thyroid and interrenal axes in this fish. The rise in  $T_3$  or  $T_4$  level in the PTU-treated tilapia after TH injections correlated with the branchial and renal  $Na^+$ ,  $K^+$ -ATPase activities, which imply an effective Na handling, though a tight regulation of  $Na^+$  and  $K^+$  transport was maintained in the plasma of these fish. The data show that high dose of PTU inactivates the thyroid axis resulting in the decreased production of  $T_3$  and  $T_4$ , though a low dose of PTU activates the thyroid axis. Our results provide evidence that exogenous  $T_3$  activates interrenal axis to produce cortisol and both exogenous  $T_3$  and  $T_4$  promote branchial and renal Na handling in PTU-induced hypothyroid tilapia.

**Key words:** Cortisol, fish, propyl thiouracil, *Oreochromis mossambicus*, osmoregulation, thyroid hormones, tilapia, stress.

### Introduction

Triiodothyronine ( $T_3$ ) and thyroxine ( $T_4$ ), the principal thyroid hormones (THs) of the hypothalamo-pituitary-thyroid (HPT) axis, are involved in development and growth of fishes (Leatherland, 1994; Power et al., 2001, Peter, 2007; Geven et al., 2009). Evidences for an involvement of THs in hydromineral regulation are available, though there is some ambiguity on its role in hydromineral balance (Leatherland, 1994; Shrimpton and McCormick, 1998; Schreiber and Specker 1999, 2000; Mancera and McCormick, 1999). Our earlier studies on freshwater tilapia provided evidence that physiological concentrations of both  $T_3$  and  $T_4$  enhance branchial sodium pump activity and chloride cell functions (Peter et al., 2000). On the contrary, there are studies that report a lack of THs effect on branchial  $Na^+$ ,  $K^+$ -ATPase activity in fishes including tilapia (Dange, 1986).

Cortisol, the product of hypothalamo-pituitary-interrenal (HPI) axis, controls many physiological functions in teleosts including hydromineral regulation (Wendelaar Bonga, 1997; Bowers et al., 2000; Dang et al., 2001). This major corticosteroid regulates chloride cell function (Li et al., 1997; Wendelaar Bonga, 1997; Perry, 1997) and stimulates branchial  $Na^+$ ,  $K^+$ -ATPase activity in many species of fishes (Mommsen et al., 1999; Mancera and McCormick, 1999). However, cortisol replacement in hypophysectomized coho salmon failed to restore completely  $Na^+$ ,  $K^+$ -ATPase activity, highlighting the involvement of other hormones in hydromineral regulation (Bjornsson et al., 1987, Richman et al., 1987). Cortisol regulates plasma  $Na^+$  levels in both seawater and freshwater fish and suppresses the rise of plasma  $Na^+$  during seawater acclimation of Coho salmon (Redding et al., 1991).

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Na<sup>+</sup>, K<sup>+</sup>-ATPase activity provides the driving force for Na<sup>+</sup> transport in fishes. It is essential for reducing the urinary loss of Na<sup>+</sup> in the kidney tubules of freshwater fish (Madsen et al., 1995) and enhances the loss of Na<sup>+</sup> through the urine in seawater-adapted fish (Nolan et al., 1999). This enzyme system in the branchial chloride cells is involved in the absorption of Na<sup>+</sup> and Cl<sup>-</sup> in freshwater fish (Perry, 1997; Fenwick et al., 1999). In tilapia THs maintain hydromineral balance by regulating branchial and renal tissue Na<sup>+</sup>, K<sup>+</sup>-ATPase activities, which increased and decreased respectively after TH treatment (Peter et al., 2000; Peter, 2007).

The interrelationship between the thyroid and the interrenals in teleosts has been the subject of many studies (Leatherland, 1987; Redding et al., 1986; Brown et al., 1991; Vijayan et al., 1997; Larsen et al., 1998; Geven et al., 2006; De Groef et al., 2006). For example, an activation of T<sub>4</sub> release from the carp renal tissue has been reported recently (Geven et al., 2009). It is believed that some of the actions of THs on hydromineral regulation may be mediated through cortisol (Leatherland, 1994; De Groef et al., 2006), though concrete evidences for such an interaction is not available (Peter, 2007). For example, T<sub>4</sub> has been shown to potentiate the stimulatory action of cortisol on Na<sup>+</sup>, K<sup>+</sup>-ATPase activity (Dange, 1986). Shrimpton and McCormick (1998) demonstrated a positive interaction of T<sub>3</sub> with corticosteroid receptors in juvenile Atlantic salmon. Redding et al. (1986) provided evidence of a cortisol-stimulated increase in T<sub>3</sub> clearance and not T<sub>4</sub> clearance in eels. Conversely, no correlation between thyroid and cortisol has been observed in salmon (Madsen, 1990), rainbow trout (Gomez et al., 1997) and mummichog (Mancera and McCormick, 1999). The physiological interaction of THs and cortisol is still unclear in spite of certain studies that have shown an association between the HPT and HPI axes in fishes (e.g., Young et al., 1989; De Groef et al., 2006).

Lack of conclusive evidence for an interaction between thyroid and interrenal axes in fish (McCormick, 2001; Evans et al., 2005; Peter, 2007) prompted us to undertake this study to gather more information on the function of interrenal axis in a hypothyroid fish. Propylthiouracil (PTU), a thyroid inhibitor, was used in this study to induce hypothyroidism in the tilapia. PTU inhibits intrathyroidal synthesis of TH by interfering with thyroid peroxidase-mediated iodine utilization and the coupling of mono- and diiodotyrosines (MIT and DIT)

which are required for the synthesis of T<sub>3</sub> and T<sub>4</sub> (Cooper, 2005; van der Ven et al., 2006). Furthermore, it blocks the peripheral conversion of T<sub>4</sub> to T<sub>3</sub> (Cooper, 2005). PTU administration has been shown to reduce THs levels in peripheral tissues and feedback stimulation of the thyroid in zebrafish (van der Ven et al., 2006). Similar hypothyroidic function has been reported in climbing perch after thiouracil treatment (Peter and Oommen, 1989; Nair and Oommen, 1997).

## Materials and Methods

### *Fish and maintenance*

Adult mozambique tilapia *Oreochromis mossambicus* (body mass 50 ± 5 g) of both sexes in pre spawning phase were kept in separate 100 L glass aquaria (density 3 gm L<sup>-1</sup>) containing adequately aerated well water of 28 °C at 12h:12 h L:D cycle. Fish were fed daily with commercial fish feed at 1.5 % total body weight.

### *Experimental protocol*

Experimental hypothyroidism was tested by feeding varied doses (5, 10 and 20 µg g<sup>-1</sup>) of PTU (Sigma, St Louis, Missouri, USA) for 10 or 15 days. Fish were divided into groups of six each and kept in separate tanks. Two sets of experiment were conducted. Control feed at a ration of 1.5% body weight was fed to control fish daily for 10 or 15 days. Test groups of fish were fed with PTU meal (5 and 10 µg g<sup>-1</sup>) over 10 or 15 days. On day 11 fish groups containing both control-fed and PTU-fed were sampled. Similarly, the other fish groups including control fish were sampled on day 16. In the second set of experiment, fish groups were given high dose (20 µg g<sup>-1</sup>) of PTU for 15 days. On day 16 fish in these three groups were injected with saline, T<sub>3</sub> or T<sub>4</sub> (40 ng g<sup>-1</sup>) for 24 h. All fish were sampled at 24 h after feeding or injection. No mortality was observed in any fish groups during the experiments.

### *Sampling procedure*

Fish in each set of experiment were netted and anaesthetized in a 2-phenoxyethanol solution (0.1% SRL, Mumbai) and blood samples were collected by caudal puncture using heparinised syringe fitted with 23 gauge needle. Plasma was separated by centrifugation (3 min, 5,000 xg) and stored at -20 °C. Fish were then killed by spinal transection and the gill arches and the kidney were excised and placed in 2 ml of ice cold SEI buffer (0.3 M sucrose, 20 mM Na<sub>2</sub>EDTA, 0.1 M imidazole, pH 7.4) and stored at -20 °C.

## Analyses

Plasma cortisol, and plasma total  $T_4$  and  $T_3$  were quantified by RIA (Peter, 2007). The specific activity of ouabain-sensitive  $Na^+$ ,  $K^+$ -ATPase was measured in homogenates prepared from branchial and renal tissues as described earlier (Peter et al., 2000). The protein concentration in homogenates was measured with a commercial Biuret protein-assay kit (Bio-Rad, Hercules, USA) using bovine serum albumin as standard. Phosphate release was quantified spectrophotometrically and the specific activity expressed as  $mmol Pi h^{-1} mg protein^{-1}$ . Plasma  $Na^+$  and  $K^+$  concentrations were measured with a flame-photometer (Systronics 129, New Delhi, India). The  $Cl^-$  concentration was determined spectrophotometrically in terms of formation of ferrothiocyanate complex. Plasma osmolality ( $mOsm Kg^{-1}$ ) was measured with a micro-osmometer (Gonotech 030, GmBH, Germany). Heparinized capillaries were filled with blood, centrifuged for 3 min at 5000  $xg$  and the haematocrit value was determined.

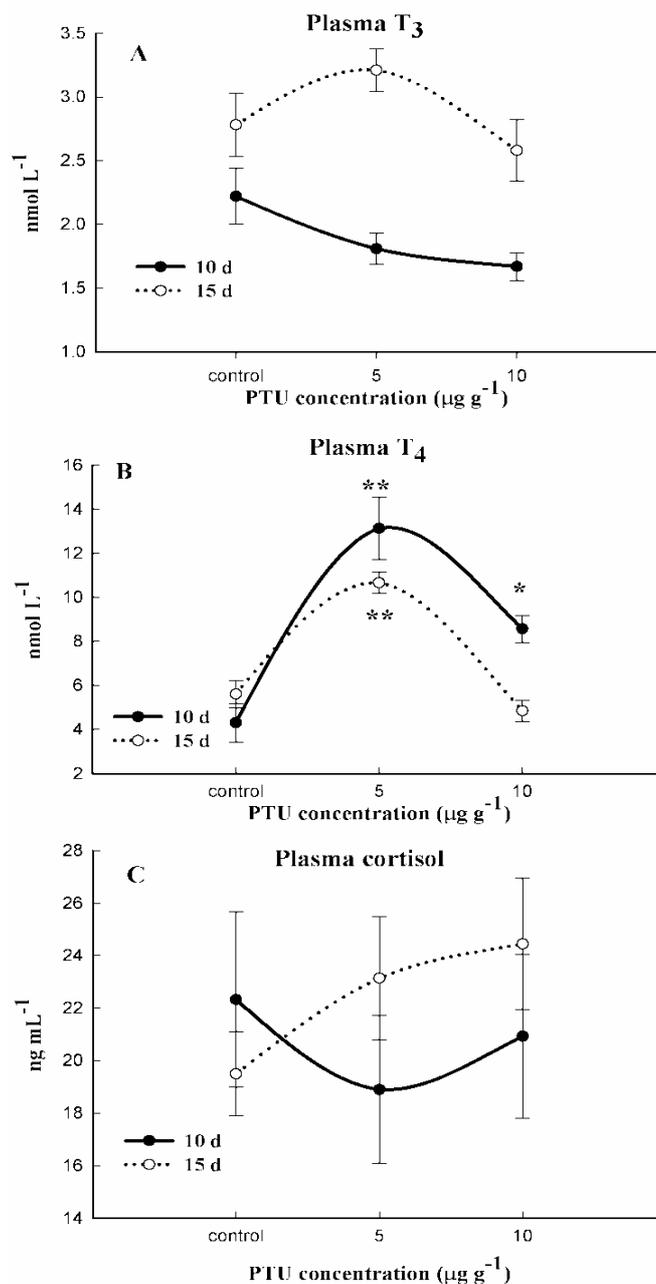
## Statistical analyses

Data were presented as means  $\pm$  S.E.M. for six fish and checked for normal distribution and variance homogeneity. One-way analysis of variance followed by Student-Newman-Keuls (SNK) multiple range tests (Instat-3, Graphpad Software Inc., San Diego, USA) were used to assess the significance between treatments and significance was accepted if  $P < 0.05$ .

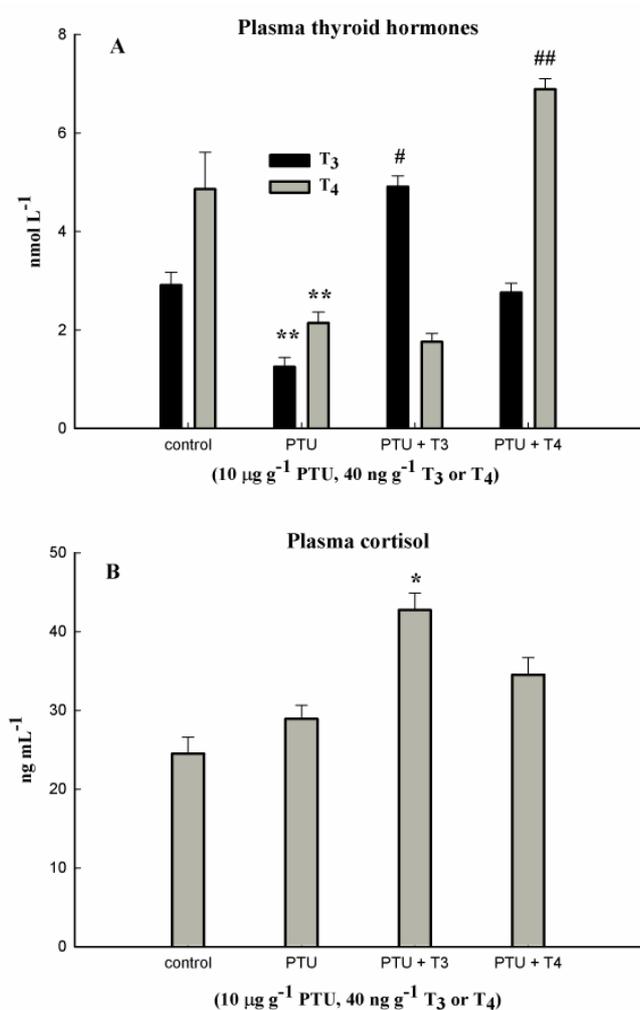
## Results

### Plasma $T_3$ , $T_4$ and cortisol

Feeding tilapia with low doses of PTU ( $5 \mu g g^{-1}$ ) for 10 and 15 days resulted in significant increase ( $P < 0.01$ ) in plasma  $T_4$  without affecting plasma  $T_3$ , compared with the control fish (Fig. 1A and B). However, feeding fish with  $20 \mu g g^{-1}$  of PTU resulted in significant decrease ( $P < 0.01$ ) of plasma  $T_3$  and  $T_4$  levels (Fig. 1A). Replacement of  $T_3$  ( $40 ng g^{-1}$ ) brought about significant increase of plasma  $T_3$  ( $P < 0.01$ ) level after 24 h without any change in  $T_4$  level (Fig. 2A). Treatment of  $T_4$  brought about significant increase in the plasma  $T_4$  ( $P < 0.01$ ) level in the PTU-fed tilapia (Fig. 2A).



**Fig. 1.** Plasma  $T_3$  (A),  $T_4$  (B) and cortisol (C) levels in freshwater tilapia after low doses ( $5$  and  $10 \mu g g^{-1}$ ) of PTU feeding for 10 and fifteen days. Each point is mean  $\pm$  SEM. The data were tested for statistical significance between treatments with the SNK test. \* ( $P < 0.05$ ) shows significance between control and PTU fed tilapia.



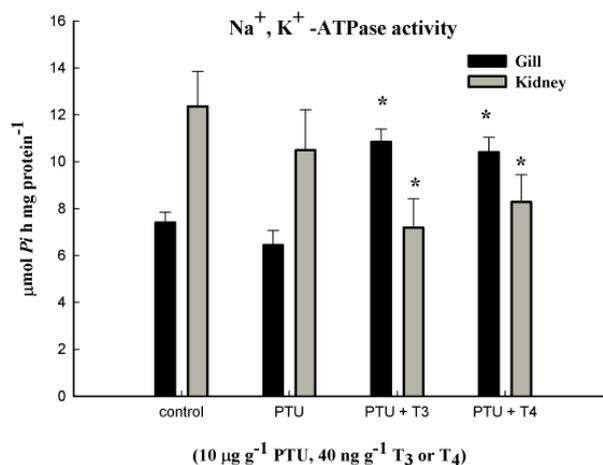
**Fig. 2.** Plasma T<sub>3</sub>, T<sub>4</sub> and cortisol levels in freshwater tilapia after PTU feeding (20  $\mu\text{g g}^{-1}$ ) for 15 days with or without a single injection of T<sub>3</sub> (40  $\text{ng g}^{-1}$ ) or T<sub>4</sub> (40  $\text{ng g}^{-1}$ ) for 24 h. Each column, mean  $\pm$  SEM. The data were tested for statistical significance between treatments with the SNK test. \* ( $P < 0.05$ ) shows significance between control and PTU fed tilapia # ( $P < 0.05$ ) and ## ( $P < 0.01$ ) show significance between PTU fed tilapia and PTU fed + TH treated fish.

The plasma cortisol level in the fish fed 5 and 10  $\mu\text{g g}^{-1}$  of PTU failed to show any change (Fig. 1C). However, a significant increase in plasma cortisol ( $P < 0.01$ ) occurred in the 20  $\mu\text{g g}^{-1}$  PTU-treated tilapia after a single T<sub>3</sub> injection (Fig. 2B).

#### Na<sup>+</sup>, K<sup>+</sup>-ATPase activity

The branchial and renal Na<sup>+</sup>, K<sup>+</sup>-ATPase activities remained unchanged following feeding of low doses of PTU for either 10 or 15 days (data not shown). Feeding PTU (20  $\mu\text{g g}^{-1}$ ) for 15 days altered neither branchial nor renal Na<sup>+</sup>, K<sup>+</sup>-ATPase activity (Fig. 3). However,

treatment of T<sub>3</sub> or T<sub>4</sub> to PTU-treated fish brought about significant ( $P < 0.01$ ) increase in the branchial Na<sup>+</sup>, K<sup>+</sup>-ATPase activity ( $P < 0.05$ ) and decrease in the renal Na<sup>+</sup>, K<sup>+</sup>-ATPase activity (Fig. 3).



**Fig. 3.** Branchial and renal Na<sup>+</sup>, K<sup>+</sup>-ATPase activities in freshwater tilapia after PTU feeding (20  $\mu\text{g g}^{-1}$ ) for 15 days with or without a single injection of T<sub>3</sub> (40  $\text{ng g}^{-1}$ ) or T<sub>4</sub> (40  $\text{ng g}^{-1}$ ) for 24 h. Each column, mean  $\pm$  SEM. The data were tested for statistical significance between treatments with the SNK test. \* ( $P < 0.05$ ) shows significance between PTU fed fish and PTU fed and TH treated fish.

#### Plasma parameters

Neither PTU-treatment nor treatment of T<sub>3</sub> or T<sub>4</sub> in PTU-fed tilapia altered the plasma Na<sup>+</sup> and K<sup>+</sup> contents (Table 1). Similarly, plasma osmolality and haematocrit were not different from the control groups and were considered normal (Table 2).

#### Discussion

Data to the effect that PTU-feeding brings about hypothyroidism in tilapia but produces no effect on interrenal activity, are presented. An activation of interrenal axis occurred in the hypothyroid tilapia supplemented with T<sub>3</sub>. We also demonstrate that replacement of THs stimulate hydromineral capacity of the PTU-induced hypothyroid tilapia.

This study indicates a direct and specific effect of PTU on thyroid activity in tilapia as this goitrogen in its low dose activates thyroid axis by releasing T<sub>4</sub>, whereas in its high dose, it inactivates thyroid by inhibiting T<sub>3</sub> and T<sub>4</sub> production. It is likely that the low doses of PTU might have induced hyperplasia of thyroid follicles and releasing its stored T<sub>4</sub> into the blood. This view is consistent with

**Table 1** Plasma Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> levels in freshwater tilapia after 20 µg g<sup>-1</sup> of PTU feeding for fifteen days, and 40 ng g<sup>-1</sup> TH for 24 h. Each value is mean ± SEM. The data were tested for statistical significance between treatments with the SNK test.

	Na <sup>+</sup> (mmol L <sup>-1</sup> )	K <sup>+</sup> (mmol L <sup>-1</sup> )	Cl <sup>-</sup> (mmol L <sup>-1</sup> )
Control + saline	155 ± 2.15	4.91 ± 0.36	133.4 ± 1.18
PTU 20 µg g <sup>-1</sup> for 15 d + saline	151 ± 1.84	5.14 ± 0.41	132.6 ± 1.71
PTU 20 µg g <sup>-1</sup> for 15 d + 40 ng T <sub>3</sub>	153 ± 1.69	5.29 ± 0.51	134.8 ± 1.69
PTU 20 µg g <sup>-1</sup> for 15 d + 40 ng T <sub>4</sub>	152 ± 1.79	5.89 ± 0.34	131.4 ± 1.72

**Table 2** Plasma haematocrit and osmolality levels in freshwater tilapia after 20 µg g<sup>-1</sup> of PTU feeding for fifteen days, and 40 ng g<sup>-1</sup> TH for 24 h. Each value is mean ± SEM. The data were tested for statistical significance between treatments with the SNK test.

	Haematocrit value (%)	Osmolality (mOsmol Kg <sup>-1</sup> )
Control + saline	30.4 ± 1.21	312 ± 1.42
PTU 20 µg g <sup>-1</sup> for 15 d + saline	32.8 ± 1.32	318 ± 1.23
PTU 20 µg g <sup>-1</sup> for 15 d + 40 ng T <sub>3</sub>	29.6 ± 1.19	314 ± 1.51
PTU 20 µg g <sup>-1</sup> for 15 d + 40 ng T <sub>4</sub>	30.4 ± 1.32	315 ± 1.65

the report on the thyroid hypertrophy observed in zebrafish after PTU feeding (van der Ven et al., 2006). The unaffected plasma T<sub>3</sub> in the low dose PTU-treated fish, on the contrary, supports the notion that PTU would impair the T<sub>4</sub> conversion to T<sub>3</sub>. Such impaired TH synthesis was also found in zebrafish (van der Ven et al., 2006). High doses of PTU feeding in tilapia, on the contrary, substantially inhibited TH production and made these tilapia experimentally hyperthyroidic as demonstrated in zebrafish (van der Ven et al., 2006). This hyperthyroid tilapia, however, failed to show any impairment in their hydromineral effects, probably because of the involvement of other hormones in the absence of THs.

A close functional relationship exists between TH and corticosteroids in mammals, though little evidence for such an interaction is available in fish (Leatherland, 1994; Peter, 2007). There is no activation of interrenal axis in PTU-fed fish and it is likely that some other hormone(s) are involved in the maintenance of the hydromineral regulation in the PTU-induced hypothyroid tilapia. Surprisingly, an up-regulated interrenal axis, as indicated by high levels of plasma cortisol, occurred in hypothyroid

tilapia following T<sub>3</sub> replacement. This positive correlation of exogenous T<sub>3</sub> and cortisol levels in hypothyroid tilapia is probably due to a direct action of T<sub>3</sub> on the interrenal axis which becomes apparent only in the PTU-induced hypothyroid fish. This interpretation is in agreement with a report on the stimulatory effects of THs on the interrenal axis of coho salmon (Young, 1988). It appears that the exogenous T<sub>3</sub> that stimulates the interrenal tissue to produce more cortisol might lead to an increased clearance of T<sub>3</sub>, as cortisol-stimulated increase in T<sub>3</sub> clearance has been reported in European eels (Redding et al., 1986) and coho salmon (Redding et al., 1984). The results of other studies also indicate a rapid clearance of T<sub>3</sub> after cortisol treatment. For instance, an increased hepatic conversion of T<sub>4</sub> to T<sub>3</sub> by cortisol has been found in brook trout (Vijayan et al., 1988). However, reduction in plasma T<sub>3</sub> level by enhancing its clearance, without changing the hepatic conversion of T<sub>4</sub> to T<sub>3</sub>, occurs in rainbow trout (Brown et al., 1991). It appears that a decreased TSH production might occur in the PTU fed tilapia due to the increase in plasma T<sub>3</sub> availability through a negative feedback. This might activate corticotrophin-releasing hormone (CRH), which is responsible for cortisol release

in these hypothyroid fish. It is likely that a sort of central interaction between thyroid and interrenal axis might occur in the tilapia. Besides these central regulations of interrenal and thyroid axes, peripheral interaction between TH and cortisol has been reported in common carp (Geven et al., 2009). The release of cortisol from head kidney fragments does not happen in carp tissues after  $T_4$  treatment (Geven et al., 2006). On the contrary, exposure of head kidney and kidney fragments to cortisol and ACTH stimulates the release of  $T_4$  from these carp tissues (Geven et al., 2009). Similar interaction of thyroid and interrenal axis at the central level has been found in the carp after  $T_4$  injection that decreased the plasma cortisol (Geven et al., 2006).

Our data clearly indicate that replacement of  $T_3$  in PTU-fed hypothyroid tilapia stimulates Na handling as it stimulated branchial  $Na^+$ ,  $K^+$ -ATPase activity and decreased renal  $Na^+$ ,  $K^+$ -ATPase activity. This is in line with our earlier report on the hydromineral regulatory action of THs in freshwater tilapia (Peter et al., 2000) and, thus, confirms the role of THs in sodium handling in tilapia. Madsen and Korsgaard (1989) have shown that long-term treatment of  $T_4$  increased branchial  $Na^+$ ,  $K^+$ -ATPase activity in juvenile Atlantic salmon. Injection of  $T_3$  or  $T_4$  for five days, on the contrary, failed to produce a stimulatory action on branchial  $Na^+$ ,  $K^+$ -ATPase activity (Peter, 2007). Similarly, branchial enzyme activity remained unresponsive to  $T_4$  treatment in freshwater acclimated rainbow trout (Madsen, 1990). Since  $Na^+$ ,  $K^+$ -ATPase activity is often considered as the biomarker of chloride cell function (Dang et al., 2000), our data further point to the effect of TH on chloride cell activity in tilapia. Thus, it may be argued that THs are involved in the uptake of ions in freshwater tilapia as the chloride cells in the branchial epithelium of freshwater fish have been implicated in the uptake of ions ( $Na^+$ ,  $Cl^-$ ,  $Ca^{2+}$ ) from the surrounding water (Flik et al., 1994; Perry, 1997). Induction of a hypothyroid state by PTU treatment in freshwater tilapia did not affect the hydromineral parameters measured. This observation stands against the prominent effect of THs reported earlier on hydromineral regulation of tilapia (Peter et al., 2000). It appears that compensatory adaptations mediated by other osmoregulatory hormones such as prolactin, growth hormone or cortisol might occur in these hypothyroid tilapia as evidences have suggested a synergistic action of THs with other hormones (Leatherland, 1994; Peter and Oommen, 1993). For example, growth hormone and cortisol, along with  $T_3$ , have been reported to promote the hypo-osmoregulatory ability of coho salmon during smoltification and seawater adaptation (Young et al., 1989). Similarly,  $T_4$  treatment in tilapia had been shown to exert

an additive effect on the stimulatory action of cortisol on  $Na^+$ ,  $K^+$ -ATPase activity, though  $T_4$  alone failed to elicit a response (Dange, 1986).

Overall, our results demonstrate the sensitivity of interrenal axis to TH availability, an indication of a positive correlation between the exogenous TH and interrenal activity in the PTU-fed fish. It was found that high dose of PTU inactivates the thyroid axis of tilapia though a low dose activates the thyroid axis. Further, stimulatory action of THs on  $Na^+$  handling is clearly evident in the PTU-induced hypothyroid tilapia.

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