

In vitro antagonism of three Trichoderma spp. against Sclerotium rolfsii Sacc., a collar-rot pathogen in elephant foot yam

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ABSTRACT: The present investigation focuses on the screening of potential biocontrol agents against the collar-rot pathogen, *Sclerotium rolfsii* Sacc. on elephant foot yam, an intercrop of coconut in Andhra Pradesh. Among the three antagonists screened, Trichoderma viride Pers. Fr., T. harzianum Rifai and T. hamatum (Bon.) Bain. isolated and screened, T. hamatum inhibited the radial growth of S. rolfsii to an extent of 52.22 per cent followed by T. viride (44.11%) and T. harzianum (38.87%). All the three Trichoderma spp. were very effective in producing volatile and non-volatile metabolites that were suppressive to S. rolfsii. Viability tests on sclerotia of S. rolfsii parasitized by Trichoderma spp. revealed complete colonization and replacement of sclerotial contents by 7th day of parasitisation in case of T. hamatum and by 10th day in case the of T. viride and T. harzianum.

KEY WORDS: Antagonism, elephant foot yam, Sclerotium rolfsii, Trichoderma

Elephant foot yam (Amorphophallus paeoniifolius (Dennst) Nicolson) is an important tuber crop grown as intercrop in coconut ecosystem. The crop is often affected by collar-rot or foot-rot disease caused by the soil-borne fungus, Sclerotium rolfsii Sacc. The symptoms include water soaked lesions on the stem near the collar region. In the advanced stages, the stem collapses due to rotting and a thick, white mycelial mat is seen. Biological control is an ecofriendly and easily adoptable technology to manage soil-borne diseases. Trichoderma spp. have been found antagonistic to S. rolfsii and Rhizoctonia solani Kuhn (Chet et al., 1979; Elad et al., 1983) and are successfully used for the control of these pathogens in several crops under greenhouse and field conditions (Wells et al., 1972; Mathur and Sarbhoy, 1978; Chet et al., 1979; Hadar et al., 1979 and Elad et al., 1980). Upadhyay and Mukhopadhyay (1983) reported that T. harzianum produced diffusible antibiotics at varying amounts under in vitro conditions, which was detrimental

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to the growth of S. rolfsii. Hence the present investigation was taken up to screen different native antagonistic fungi against the collar-rot pathogen S. rolfsii and the methods of antagonism by these antagonists in controlling the pathogen.

Isolation of Trichoderma spp.

Rhizosphere soil saples were collected from healthy plants surrounding the diseased plants of elephant-foot yam. The soil samples were then dried and later sieved to fine powder and serially diluted in sterilized distilled water (SDW) to 10^{-3} and 10^{-4} concentrations. Then, 500 µl of each dilution was spread on Petri-dishes containing *Trichoderma* specific medium.

Two plates were maintained for each dilution. The plates were incubated for 4 days at 28°c and typical *Trichoderma* spp. colonies were identified according to the identification key based on the branching of conidiophores, shape of the phialides, emergence of phialophores and phialospores (Rifai, 1969).

Dual culture studies

Dual cultures of the antagonists and the test pathogen S. rolfsii were prepared by inoculating PDA discs from the growing margins of fresh fungal cultures on to Petri-dishes containing PDA (Gams *et al.*, 1980) and incubating at 29°C. The dual cultures were observed for antibiosis and agar blocks from the regions where the colonies merged were observed for hyphal interaction under the light microscope.

Viability of sclerotial bodies

The sclerotial bodies of the test pathogen from dual-culture plates that were parasitised by the *Trichoderma* spp. are then subjected to viability studies (Morton & Stroube, 1955; Abd-El Moity and Shatla, 1981). The parasitized sclerotia were surface sterilized with mercuric chloride (0.1%), transferred onto sterilized wet towel paper and incubated at 30°C temperature for 72 hours and finally plated on PDA for isolation of the pathogen. Sclerotial bodies exposed to different time intervals were subjected to viability studies.

Inhibition by volatile metabolites

Production and inhibitory effect of volatile metabolites by the antagonists were tested against the test pathogen by using procedure given by Dennis and Webster (1971). The pathogen growth was measured after 4 days of incubation at 29°C.

Inhibition by non-volatile metabolites

The antagonists that had shown inhibition in dual-culture studies were grown on potato dextrose broth to test the effect of the culture filtrates (non-volatile antibiotics) on the test pathogen by food poisoning technique (Khara and Hadwan, 1990). The culture filtrates were purified by autoclaving and the sterilized filtrate was incorporated in the medium for observing fungal growth and inhibition at different concentrations (50% and 100%). The PDA mixed filtrate in all the cases were poured (20ml each) into sterilized Petridishes and the same were inoculated with fresh disc of the test pathogen, *S.rolfsii*. Control plates were maintained simultaneously and per cent inhibition was calculated.

Mycelial interactions between *Trichoderma* spp. and *S. rolfsii*

Growth of S. rolfsii in Petri-dish dual-culture was suppressed by all the three Trichoderma spp. (T. viride, T. harzianum and T. hamatum). Highest inhibition was recorded in case of T. harnatum (52.22%) followed by T. viride (44.11%) and T. harzianum (38.87%) (Table 1). Inhibition zone was observed in the case of T. viride and T. hamatum followed by mycoparasitism whereas only mycoparasitism was noticed in case T. harzianum. In the case of T. hamatum, a yellow halo prevailed up to 10 days of interaction followed by mycoparasitism. In all the three cases, the S. rolfsii mycelium did not grow when transferred on to fresh media indicating its death. Microscopic observations made from the mycelial interaction zone revealed frequent adpressing zones of Trichoderma spp. mycelia on S. rolfsii mycelia.

Viability of Sclerotial bodies

Viability studies carried out on sclerotial

Biocontrol agent	Mycelial growth of <i>S. rolfsii</i> (mm)	Per cent inhibition of S. rolfsii mycelial growth	Mode of action	
T. viride	50.3	44.11 ^b	Antibiosis followed by mycoparasitism	
T. harzianum	55.0	38.87ª	Mycoparasitism	
T. hamatum	43.0	52.22°	Antibiosis (Yellow halo formation) followed by mycoparasitism	
Control	90	-	-	

Table 1. Dual-culture studies between Trichoderma spp. and S. rolfsii

* Numbers in each column followed by the different letters are significantly different (P=0.05).

bodies of *S. rolfsii* revealed that sclerotial body germination was affected when parasitised by *Trichoderma* spp. in dual culture studies. Sclerotial bodies gave rise to respective *Trichoderma* spp. mycelia when subjected to germination tests after 10 days of parasitisation and above. However, *T. hamatum* was found to be superior among the other *Trichoderma* spp., since parasitised sclerotial bodies from *T. hamatum* vs *S. rolfsii* dual culture plates gave rise to *T. hamatum* mycelia when subjected to germination tests after 7 days.

Inhibition by volatile metabolites of 0, 15 and 25 day old cultures of *Trichoderma* spp.

Among the Trichoderma spp., T. harzianum proved very effective in producing volatile antibiotics specific against S. rolfsii at all the three stages of exposure. This was followed by T. hamatum and T. viride, which were also effective in producing volatile substances against S. rolfsii. An increase in reduction was evident with an increase in the age of Trichoderma spp. cultures (Table 2). The results indicated the production of effective volatile antibiotics by all the antagonistic species of Trichoderma. Hyphae from the exposed cultures of S. rolfsii when transferred to fresh medium did not grow. Hence, the volatile metabolites produced by the Trichoderma spp. i.e., T. viride, T. harzianum and T. hamatum were both fungicidal and fungistatic (Claydon et al., 1987). Sawant and Mukhopadhyay (1990) while working on damping off of sugarbeet reported that older cultures of T. *harzianum* had a greater inhibitory effect on the mycelial growth of *Pythium aphanidermatum* as compared to that of younger cultures.

Inhibition by non-volatile metabolites

Culture or cell free filtrates of all the three Trichoderma spp., viz. T. viride, T. harzianum and T. hamatum were suppressive to the radial growth of S. rolfsii (Table 2). With an increase in the concentration of the culture filtrate of the Trichoderma spp., a corresponding increase in per cent inhibition of the mycelial growth of S. rolfsii was noticed. Similar results were noted with an increase in concentration of the culture filtrate of T. harzianum and inhibition of Pythium aphanidermatum on tobacco by Devaki et al. (1992). However, cell-free filtrates of T. hamatum were found to be highly effective at both the 50 per cent and 100 per cent concentration under study thus inhibiting the S. rolfsii mycelium completely. Narasimha Rao and Kulkarni (2003) reported that T. viride and T. harzianum are very effective in reducing the radial growth of S. rolfsii and also in the production of volatile and non-volatile antibiotics against the pathogen.

From the present investigations, it is evident that all the three *Trichoderma* spp., viz. *T. viride*, *T. harzianum* and *T. hamatum* are very effective against *S. rolfsii* under *in-vitro* conditions. Further, all the *Trichoderma* spp. are found to produce substantial quantities of volatile and non-volatile

Antagonist		Inhibition of S. rolfsii (%)					
	A	Volatile meta tge of antagon	bolites ist (days)	Non-volatile metabolites Concentration of culture filtrate (%)			
	0	15	25	50	100		
T. viride	4.44ª	20.00ª	46.67ª	45.00ª	65.00ª		
T. harzianum	28.89ª	55.56 [⊳]	73.33°	61.67 ^b	71.66 ^b		
T. hamatum	5.56ª	33.33°	51.11 ^b	100.00°	100.00°		

 Table 2. Effect of volatile and non-volatile metabolites of Trichoderma spp. on S. rolfsii under in vitro conditions

• Numbers in each column followed by the same letter are not significantly different.

compounds, which are detrimental to the growth of *S. rolfsii*. Moreover, all the *Trichoderma* spp. replaced the sclerotial contents of the test pathogen within 7 to 10 days by parasitization. Going by the present results, the collar-rot pathogen in elephant foot yam can be effectively checked by application of these antagonists to the soil either singly or in combination.

REFERENCES

- Abd-El Moity, H. and Shatla, M. N. 1981. Biological control of white rot disease of onion. (Sclerotium cepivorum) by Trichoderma harzianum. Phytopathologische Zeitschrift, 100: 29-35.
- Chet, I., Hadar, Y., Elad. Y., Katan, J. and Henis, Y. 1979
 Biological control of soil borne plant pathogens by Trichoderma harzianum, pp. 585-591. In: B.
 Schippers and W. Gams (Eds.). Soil borne Plant Pathogens. Academic Press, London.
- Claydon, N., Allan, M., Hanson, J. R. and Avent, A. G. 1987. Antifungal alkyl pyrones of *Trichoderma* harzianum. Transactions of the British Mycological Society, **88**: 503-513.
- Denis, C. and Webster, J. 1971. Antagonistic properties of species- growth of *Trichoderma*-I. Production of non-volatile antibiotics. *Transactions of the British Mycological Society*, **57**: 25-39.
- Devaki, N. S., Bhat, S. S., Bhat, S. G and Manjunatha, K. R. 1992. Antagonistic activities of *Trichoderma* harzianum against *P. aphanidermatum* and *P.*

myriotylum on tobacco. *Journal of Phytopathology*, **136**: 82-87.

- Elad, Y., Chet, I., Katan, J. 1980. Trichoderma harzianum: A biocontrol agent effective against Sclerotium rolfsii and Rhizoctonia solani. Phytopathology, 70: 119-121.
- Elad, Y., Chet, I., Boyle, P. and Henis, Y. 1983 Parasitism of *Trichoderma* spp on *Rhizoctonia solani* and *Sclerotium rolfsii* - scanning electron microscopy and fluorescence microscopy. *Phytopathology*, **73**: 418-422.
- Gams, W., Vander A. A., Vander Plaats-Niterink, A. J., Samson, R. A. and Stalpers, J. A. 1980.*CBS Course* of Mycology, second edition. Centraalbureau voor Schimmelcultures. Baarn, The Nethelands.
- Hadar, Y., Chet, I., and Henis, Y. 1979. Biological control of *Rhizoctonia solani* damping off with wheat bran cultures of *Trichoderma harzianum*. *Phytopathology*, **69**: 64-68.
- Khara, H. A. and Hadwan, H. A 1990. In vitro studies on antagonism of Trichoderma spp, against Rhizoctonia solani the causal agent of damping off of tomato. Plant Disease Research, 5: 144-147.
- Mathur, S. B. and Sarbhoy, A. K. 1978. Biological control of sclerotium root-rot of sugarbeet. *Indian Phytopathology*, **31**: 365-367.
- Morton, D. J. and Stroube, W. H. 1955. Antagonistic and stimulatory effects of micro-organisms upon *Sclerotium rolfsii. Phytopathology*, **45**: 417-420.

- Narasimha Rao. S. and Kulkarni, S. 2003. Effect of *Trichoderma* spp. on the growth of *Sclerotium rolfsii* Sacc. *Journal of Biological Control*, 17: 181-184
- Rifai, M. A. 1969. A revision of the genus *Trichoderma*. *Mycological papers*, no.116. Commonwealth Mycological Institute, Association of Applied Biologists, Kew, Surrey, England.
- Sawant, I. S. and Mukhopadhyay, A. N. 1990. Integration of metalaxyl with *Trichoderma*

harzianum for the control of Pythium damping-off in sugarbeet. Indian Phytopathology, **43**: 535-541.

- Upadhyay, J. P and Mukhopadhyay, A. N. 1983. Effects of non-volatile and volatile antibiotics of *Trichoderma harzianum* on the growth of *Sclerotium rolfsii*. *Indian Journal of Mycology and Plant Pathology*, **13**: 232-233
- Wells, H. D., Bell. D. K. and Jaworski. C. A. 1972. Efficacy of *Trichoderma harzianum* as a biological control for *Sclerotium rolfsii*. *Phytopathology*, 62: 442-447.